

# *In vitro* Methods used in Cytotoxicity Assays

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## Description

Cytotoxicity is well-defined as the toxicity initiated due to the action of chemotherapeutic agents on active cells. Cytotoxicity tests are very significant for nanoparticles as they aid in the determination of the recommended biomedical use. The technique for determination of cytotoxicity and cell viability contains dyes such as Trypan Blue, Alamar Blue, neutral red, and Coomassie Blue. The technique distinguishes the various cells in terms of colour. The cells are separated based on the ratio of the colour application of both living and dead cells. The other techniques for examining cytotoxicity include the tritium-labeled thymidine uptake assay, the MTT method, the WST assay, and the dehydrogenase-based assay. The cytotoxicity of titanium dioxide microbeds, the TiO<sub>2</sub> +5%Gd MBs, and TiO<sub>2</sub> +10% Gd MBs was estimated by using the resazurin assay that deals with the mitochondrial activity of cells. The mitochondrial action of MG-63 was unaffected by the management of all used microbeds. Our results show that all the used MBs is noncytotoxic, a significant requirement for particles proposed for biomedical use. Though we did not identify cytotoxicity, there is still a need to estimate additional categories of toxicity like immunotoxicity and genotoxicity, and to measure cytotoxicity to other cell types. Cytotoxicity and sensitization are the two simple tests that are related to all medical devices that come into interaction with the body. Cytotoxicity is defined as an *in vitro* test to distinguish whether the medical device may lead to cell death due to discharge of toxic substances or direct interaction. Once you have determined that it does not cause cell death, the next stage is to check whether the substance may cause any allergic reaction in the body due to a latent

leachable chemical. This process is carried out *in vivo* even though an *in vitro* model may first be used to monitor the product prior to *in vivo* trials, particularly for skin application. The irritation test is the next most extensively conducted test, and it applies to the organ that is affected by the medical device. A cytotoxicity test, which goes to early testing, is an economical method, with the benefits of a relatively simple testing method, high replicability, accurate results, and large-scale assessment as an outcome of standardization. For the cytotoxicity calculation, various cell lines are used in unity with the target application. For the evaluation of bone implant materials, marrow cells and osteoblastlike cells are commonly used, while for the evaluation of materials used within blood vessels, endothelial cells and human smooth muscle cells are frequently used. Direct and indirect contact assays are the two main techniques and are separated by culturing cells on the samples or in their extracts. After incubation for a suggested period, a microscope and/or a microplate reader are generally used to illustrate cell morphology and viability/cytotoxicity. For example, the cytotoxicity of the HA-coated Mg-4Zn-1Ca-0.6Zr alloy has been estimated using an indirect assay. The cytocompatibility study is to examine the interactions between magnesium and tissue cells at an initial stage. Because Mg and its alloys are mainly to be used as hard tissue alternates and stent materials, complete *in vitro* biocompatibility studies, containing hemocompatibility, cytotoxicity tests, and antibacterial effects, are required.

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