

Vaccines and Neuroinflammation

Giannotta G1* and Giannotta N2

¹Basic Pediatrician, Provincial Health Authority of Vibo Valentia, Vibo Valentia, Italy ²Università Magna Grecia, Medical and Surgery Sciences, Catanzaro, Italy

Abstract

Giannotta and Giannotta, Int J Pub Health Safe 2018, 3:3

Background: Post-vaccination adverse reactions (AEs) are a reason of strong debate among scientists. Unfortunately, we often make the mistake of discussing just the epidemiology but not the molecular biology. The action mechanism of the vaccines is still not fully known despite the fact that aluminum adjuvants have been used for about 100 years.

Hypothesis: We hypothesized a link between vaccinations and neuroinflammation. The peripheral proinflammatory cytokines (IL-1 β , IL-6, and TNF- α), expressed after the injection of the vaccines can reach the brain and can cause neuroinflammation after microglia activation. Elevated pro-inflammatory cytokines, particularly TNF- α , have been described in studies regarding the cytokines profile in autistic children. IL-1 β represents a cytokine that controls the local pro-inflammatory cascade and thereby affects the balance between protective immunity and destructive inflammation. A subgroup of children with ASD (Autism Spectrum Disorder) has developed neuroinflammation. Several postmortem studies have confirmed the activation of microglia and neuroinflammation. A recent study has shown the presence of aluminum in the brain of individuals with autism and this aluminum was also found in microglia cells. Aluminum from vaccines is redistributed to numerous organs, including brain, where it accumulates. Each vaccine adds to this tissue different level of aluminum. Aluminum, like mercury, activates microglia leading to chronic brain inflammation and neurotoxicity.

Conclusion: The molecular mechanisms presented here demonstrate how peripheral cytokines, expressed after vaccination, can cause neuroinflammation in some subjects, after microglia activation, depending on the immunogenetic background and the innate immune memory.

Keywords: Neuroinflammation; Vaccines; Microglia activation; ASD (Autism Spectrum Disorder); HPV vaccine AEs; Immune innate memory; Vaccines and aluminum; Peripheral cytokines

Introduction

Vaccines are an important health policy tool and have changed the history of infectious diseases. In recent years, the number of vaccines inject to infants has increased, and many doses are administered during the first year of life, when the immune system and the central nervous system have yet to complete their development. Furthermore, the efficacy of the vaccination program is affected by the the possible presence of maternal antibodies in the infant, and by the degree of environmental chemical pollution. Some vaccines contain the specific antigen associated with aluminum, such as Infanrix-Hexa (Combined Diphtheria-Tetanusacellular Pertussis, Hepatitis B, inactivated Poliovirus and Haemophilus influenzae type b Vaccine), MenC (Meningococcal conjugate vaccines), MenB (Serogroup B meningococcal vaccines), and pneumococcal conjugate vaccine (Prevenar). Aluminum performs the essential task of activating the innate immune system. Other vaccines contain alive virus, such as MMR (Measles, Mumps, Rubella), and MMRV (Measles, Mumps, Rubella, and Varicella), and do not contain aluminum. Each injection of vaccine, regardless of the type of vaccine, is followed by the production of variable amounts of pro-inflammatory cytokines, which exert both local effects and at a distance from the production site. Postvaccination adverse events (AEs) are a reason of strong debate among scientists. Unfortunately, we often make the mistake of discussing just the epidemiology but not the molecular biology. The action mechanism of the vaccines is still not fully known despite the fact that aluminum adjuvants have been used for about 100 years.

Since the peripheral cytokines, produced after the injection of the vaccines, are able to reach the central nervous system, we hypothesize that these cytokines can have effects on the microglia (macrophages of

the central nervous system), and that these effects can be facilitated by repeated vaccinations to infants during the first year of life. In this paper, we studied the molecular biology of vaccines and present the putative mechanisms that link the injection of vaccines to neuroinflammation, which can occur as the effect of microglial activation and its subsequent pro-inflammatory orientation (M1).

Aluminum

Since equating the aluminum introduced into the body with food and water (essentially extracellular) to the one injected with the vaccines (exclusively intracellular) is not a matter of science (because of the total diversity in kinetics and bio-dynamics), the Mitkus et al. [1] estimates are not consistent with the significant study of Priest [2], and with the others mentioned in this paper. Indeed, Mitkus et al. [1] state that the burden of aluminium accumulated in the body from vaccines and diet, throughout an infant's first year of life, is significantly less than the corresponding safe level of aluminium burden modeled using the regulatory MRL. They conclude by stating that: Those episodic exposures to vaccines that contain aluminum adjuvant continue to be extremely low risk to infants and that the benefits of using vaccines

*Corresponding author: Girolamo Giannotta, Basic Pediatrician, Provincial Health Authority of Vibo Valentia, Vibo Valentia, Italy, Tel: +39096341930; E-mail: girolamo.giannotta@inwind.it

Received September 06, 2018; Accepted September 18, 2018; Published September 21, 2018

Citation: Giannotta G, Giannotta N (2018) Vaccines and Neuroinflammation. Int J Pub Health Safe 3: 163.

Copyright: © 2018 Giannotta G, et al. This is an open-access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

containing aluminum adjuvant outweigh any theoretical concerns".

Unfortunately, many studies published on the hypothetical safety of aluminum injected with vaccines are not conclusive, and there are not randomized controlled trials (RCTs) on the safety of aluminum injected with vaccines. No significant change in levels of urinary or serum aluminum were seen after vaccination of preterm infants with vaccines containing a total of 1200 μ g of aluminum [3]. Also this study confirms that the aluminum injected with the vaccines is not found in the serum of the vaccinated subjects, but does not show that the vaccine aluminum is safe. Mateusz et al. [4] have shown that infant blood-aluminum and hair-aluminum varied considerably but did not correlate with their immunization history. The aluminum injected with the vaccines cannot correlate with that of the blood and/or hair because it is not found free in the blood, as repeatedly said.

In 2018, it should already be quite clear that the aluminum injected with the vaccines cannot be measured in the serum because it is only found in the cells of the monocyte/macrophage lineage. Besides, intramuscular injection of alum-containing vaccine in mice was associated with the appearance of aluminium deposits in distant organs, such as spleen and brain, where they were still detected one year after the injection [5]. Nanomaterials can be transported by monocytelineage cells to draining lymph nodes, blood and spleen, and, similarly to HIV, may use CCL2-dependent mechanisms to penetrate the brain. In normal conditions, this occurs at a very low rate, thus explaining good overall tolerance of alum despite its strong neurotoxic potential. However, continuously escalating doses of this poorly biodegradable adjuvant in the population may become insidiously unsafe, especially in the case of over immunization or immature/altered blood brain barrier or high constitutive CCL-2 production [5]. Aluminum oxyhydroxide (Alhydrogel®) is a nano-crystalline compound forming aggregates that has been introduced in vaccine for its immunologic adjuvant effect in 1926. Although generally well tolerated on the short term, it has been suspected to occasionally cause delayed neurologic problems in susceptible individuals [6]. Concerns linked to the use of alum particles emerged following the recognition of their causative role in the socalled macrophagic myofasciitis (MMF), lesion detected in patients with myalgic encephalomyelitis/chronic fatigue syndrome (ME/ CFS). MMF revealed an unexpectedly long-lasting biopersistence of alum within immune cells in presumably susceptible individuals, thus stressing the fundamental misconception of its biodisposition [7].

With regard to aluminium introduced with food and water, available studies indicate that the oral bioavailability of aluminium in humans and experimental animals from drinking water is in the range of 0.3%, whereas the bioavailability of aluminium from food and beverages generally is considered to be lower, about 0.1% [8]. The Joint Food and Agriculture Organization/World Health Organization Expert Committee on Food Additives (JECFA) established a Provisional Tolerable Weekly Intake (PTWI) for aluminium of 1 mg/kg body weight in 2006 [8]. Five years later, the committee re-evaluated the safety of aluminium and proposed a PTWI of 2 mg/kg body weight in June 2011. The PTWI applies to all aluminium compounds in food, including food additives [9]. Based on the maximum reported concentration of aluminium in breast milk, exposures of exclusively breastfed infants may be up to 6% of the PTWI of 2 mg/kg bw (2,000 μ g/kg bw), with the highest exposure in high level consumers aged 0-3 months. In infants fed exclusively with ready-to-consume formulae, exposures to aluminium may be 4% of the PTWI. In those fed exclusively with powdered formulae, the exposure to aluminium could be up to 8, 11 and 21% of the PTWI, respectively from cows' milk, goats' milk and soya based products. In addition, the water used to reconstitute the infant formula could give further exposure of up to some 248 μ g/kg bw/ week aluminium (12% of the PTWI), resulting in total exposure of up to 33% of the PTWI.

The 33% of 2,000 μ g/kg bw is 660 μ g/kg bw. Since the absorption of aluminum from food is low, generally 0.5% or less [10], 0.5% of 660 μ g/kg bw is equal to 3.30 μ g/kg bw per week. As a year to 52 weeks, the total amount of aluminum theoretically absorbed in the first year of life should be 171.6 μ g/kg (52 × 3.30 μ g/kg). Moreover, assuming that at one year of life the weight of a child is on average 10 kg, it should have absorbed totally 1,716 μ g of aluminum (Table 1).

As shown by the data presented in Table 1, the amount of aluminum injected in the first year of life, for Italian infants, is 2.52 times greater, compared to the maximum amount absorbed at one year from the diet [11-13]. However, most aluminium that enters the blood is excreted in urine within a few days or weeks and the gastrointestinal tract provides an effective barrier to aluminium uptake [2], while the aluminum administered with the vaccines is internalized by the cells of the monocyte/macrophage lineage, and for this reason it is intracellular and cannot be eliminated by the kidney.

All adjuvants modulated a common set of 168 genes and promoted antigen-presenting cell recruitment. Alum regulated 312 genes [14]. A number of *in vitro* experiments [15] have shown that alum activates the NLRP3 inflammasome in macrophages which, in turn, activates caspase-1 and consequent production of interleukin- (IL-) 1 β (IL-1 β). Upon activation, members of the Nod-Like Receptors (NLR) family, such as NLRP3, form complexes with ASC and pro-caspase-1. The complex formed by these molecules is referred to as the inflammasome. The NLRP3 inflammasome is activated by a number of materials, including alum. Whatever the cause of inflammasome activation, the consequences include production of active caspase-1 thus conversion of inactive precursor cytokines of the IL-1 family, including IL-1 β , IL-18 and IL-33, to their active forms [16].

In summary, the aluminum salts injected as vaccine adjuvants are taken by the innate immunity cells (especially from dendritic cells), they engage a receptor called NLR (NLRP3), which together with other proteins is organized into an intracellular macromolecular complex that activates the enzyme caspase-1. This enzyme converts pro-IL-1 β and pro-IL-18 into their active forms (IL-1 β , and IL-18). The role of caspase-1 is not limited to the conversion of pro IL-1ß to IL-1ß alone, but it strongly affects the secretion of proinflammatory cytokines: IL-1β, IL-1α, IL-6, TNF-α, IL-18 and IFN-γ. IL-1 is a primary regulator of inflammatory and immune responses. Via its type I receptor it activates specific protein kinases, including the nuclear factor kappa-light-chainenhancer (NF-KB) inducing kinase (NIK) and three distinct mitogenactivated protein (MAP) kinase cascades. These modulate a number of transcription factors including NF-KB, AP1 and CREB, each of which regulate a plethora of immediate early genes central to the inflammatory response [17]. Therefore, each injection of the vaccine produces a proinflammatory response. An immune response to the vaccine antigens

| Vaccines | Aluminum | Aluminum Doses | | |
|-------------------------------------|--------------------------------------|----------------|----------|--|
| Bexero [11] | 500 µg 3 | | 1,500 µg | |
| Prevenar 13 [12] | 125 µg | 3 | 375 µg | |
| Infanrix-Hexa [13] | 820 µg | 3 | 2,460 µg | |
| Total aluminum inject | - | | | |
| Maximum amount absorbed at one year | For infant with 10 kg of body weight | | 1,716 µg | |

Table 1: Aluminum in vaccines (Italian schedula) and diet.

(to the quantities currently present in the vaccines), is non-possible without a pro-inflammatory response, which is produced by adjuvants.

Vaccines

Vaccines have drastically reduced infant death and disability caused by preventable diseases in the United States [18]. However, some vaccines may not achieve the desired goal, or they can cause serious AEs that alter the benefit-risk balance by shifting the balance in the direction of risk.

Vaccine 4CMenB (Bexero)

This vaccine has two major problems: First, it is strongly reactogenic; secondly, it provides little individual and collective long-term protection. The incidence of potentially vaccine-related, acute serious AEs in individuals receiving 4CMenB was low (5.4 per 1000 individuals), but was significantly higher than routine vaccines (1.2 per 1000 individuals). Long-term immunogenicity against strain NZ98/254 (Bexero) remains suboptimal [19]. Soeters et al. [20] have investigated MenB-FHbp impact on meningococcal carriage. Carriage prevalence on campus remained stable, suggesting MenB-FHbp does not rapidly reduce meningococcal carriage or prevent serogroup B carriage acquisition. In a university setting, the majority of meningococcal carriage was due to nongroupable strains, followed by serogroup B [21]. MenB-FHbp and MenB-4C do not have a large, rapid impact on meningococcal carriage and are unlikely to provide herd protection in the context of an outbreak response [22].

Human papillomavirus vaccines (HPV Vaccines)

HPV vaccines are neither safe nor effective as claimed by so much scientific literature. These vaccines are anti-virus vaccines, but they are not anti-tumor vaccines. In fact, they can prevent the infection (even if it is not always so), that before producing precancerous lesions (CIN2/3), it must become persistent. There are 3 licensed HPV vaccines (Gardasil 4, Gardasil 9 and Cervarix). All HPV vaccines are virus-like particles (VLPs) based on the major HPV capsid protein L1. Antigenically the vaccines are very similar but they are produced in different systems and contain different adjuvants. The Gardasil® vaccine is adjuvanted with aluminum hydroxyphosphate sulfate. The Cervarix® vaccine is formulated with AS04, which contains aluminum hydroxide salts and the Toll-Like Receptor 4 (TLR4) agonist MPL (3-O-desacyl-4'-monophosphoryl lipid A). Elevated levels of circulating plasma cytokine/chemokines were observed post first vaccination in Gardasil® recipients and proinflammatory cytokines were elevated following 1st and 3rd Cervarix® vaccinations [23]. The new Cochrane Review [24] on the HPV vaccine for cervical cancer prevention in girls and women, included studies that were not truly randomized, double-blind, placebo-controlled studies (RCTs) because in several paper they labelled aluminium salts as a placebo. As previously reported, aluminium regulates 312 genes [14] and for this reason it is not a placebo. This can be considered a scientific oxymoron.

In the VRBPAC Background Document [25] two important concerns were identified during the course of the efficacy review of this BLA. One was the potential for Gardasil[™] to enhance disease among a subgroup of subjects who had evidence of persistent infection with vaccine-relevant HPV types at baseline. The other concern was the observations of CIN 2/3 or worse cases due to HPV types not contained in the vaccine. The results of exploratory subgroup analyses for study 013 suggested a concern that subjects who were seropositive and PCR-positive for the vaccine-relevant HPV types had a greater number of CIN 2/3 or worse cases [25]. Fischer et al. [26] have hypothesized that there

may be a continuous change in the prevalence of HPV types following vaccination. Vaccinated young women had a higher prevalence of high-risk non-vaccine types [27]. Ultimately, the vaccination can induce evolutionary responses of viruses to vaccines, and they may appear several years after the introduction of such control measures.

Post-vaccination AEs can be added to this complicated scenario. In Japan, the period of HPV vaccination overlapped with that of the development of HPV vaccine-related symptoms in the vaccinated patients, including chronic regional pain syndrome (CRPS) and autonomic and cognitive dysfunctions. Moreover, 28 months have passed since the recommendation for HPV vaccination was withdrawn, and new HPV vaccine-related symptoms have not been observed during 14-month follow-up period. The sequence of these events suggests that HPV vaccination is temporally related to the development of these symptoms in Japanese adolescent girls [28].

Post-vaccination inflammatory syndrome: A new syndrome?

Giannotta [29] hypothesized that several vaccine AEs may be determined by the excessive production of proinflammatory cytokines, determined by the injection of these vaccines. He hypothesized that these post-vaccination reactions fall into the ASIA syndrome, but represent a sub-group of clinical syndromes determined by the excessive expression and secretion of pro-inflammatory cytokines. To elaborate this hypothesis, he started from two considerations: 1- all the girls affected by important adverse reactions, all around the world, experience an almost identical neurological symptomatology after the injection of the vaccines; 2 - if these AEs are attributable to vaccines it is necessary to understand how the vaccines is able to produce symptoms, essentially neurological in nature. In 2013, a number of safety signals arose for HPV vaccines: CRPS in Japan, postural orthostatic tachycardia syndrome (POTS) in Denmark, and long-lasting fatigue in the Netherlands [30-32]. The European Medicines Agency (EMA) reported a review of the safety concerns of POTS and CRPS in November 2015. The conclusion was that the current evidence does not suggest a causal association between HPV vaccines and POTS or CRPS [33]. Anyway, high levels of circulating plasma cytokine/chemokines were observed in post-vaccination time with HPV vaccines [23].

Once you have arrived at this point, the question arises: Can a peripheral immune stimulation produce a central nervous system response? Actually, there is a natural condition that certifies its existence: Sick behavior. Elevated levels of proinflammatory cytokines drive most if not all aspects of the sickness response either directly or indirectly [34]. Proinflammatory cytokines, including IL-1, IL-6, and tumor necrosis factor- α (TNF- α) activate the production and/or release of secondary inflammatory mediators [35], such as prostaglandins (PGs) and nitric oxide (NO). Proinflammatory cytokines directly stimulate numerous neurohormonal systems. Proinflammatory cytokines can directly interact with microglia and astrocytes in the glia limitans. Once microglia are activated, astrocytes are recruited leading to further activation of neuroinflammatory signals [36].

Having verified that peripheral cytokines can produce microglial activation, only the putative link between some AEs and the cytokines released after injection of these vaccines remains.

CRPS type I in Japan

Every pain syndrome has an inflammatory profile consisting of the inflammatory mediators that are present in the pain syndrome. The biochemical mediators of inflammation include cytokines, neuropeptides, growth factors and neurotransmitters. Irrespective of the type of pain whether it is acute or chronic pain, peripheral or central pain, nociceptive or neuropathic pain, the underlying origin is inflammation and the inflammatory response [37]. Table 2 lists a number of neurological syndromes (painful and painless) and their relationship to proinflammatory cytokines.

The immune system via peripheral and central release of proinflammatory cytokines contributes significantly to pain modulation [38]. There is abundant evidence that certain pro-inflammatory cytokines such as IL-1β, IL-6, and TNF-a were involved in the process of pathological pain [39] IL-1β was one of the first cytokines to be implicated in peripheral nerve injury-induced neuropathic pain mechanisms in rodents, and TNF- α is critical for the development of neuropathic pain [40]. For instance, nociceptors are known to be IL-1β sensors and IL-1β can directly activate nociceptors to generate action potentials and induce hyperalgesia [41]. CRPS describes an array of painful conditions (nociceptive pain in CRPS type I) that are characterized by a continuing (spontaneous and/or evoked) limb pain that is seemingly disproportionate in time or degree to the usual course of any known trauma or other lesion. The pain is regional (not in a specific nerve territory or dermatome) and usually has a distal predominance [42]. Symptoms of CRPS-I include spontaneous pain ("burning" pain referred to the skin, and "aching" pain referred to deep tissues), and a variety of stimulus-evoked abnormal pain sensations, including mechano-hyperalgesia, mechano-allodynia, cold-allodynia and sometimes heat-hyperalgesia. Other symptoms include disorders of vasomotor and sudomotor regulation; trophic changes in skin, hair, nails, and bone; and dystonia and other motor abnormalities [43]. Thus, the most prominent mechanism appears to be the inflammatory process because all the classic signs of inflammation (oedema, redness, hyperthermia, and impaired function) are conspicuous in the early stages of CRPS [44].

High levels of the proinflammatory cytokines (TNF- α and IL-6) have been found in skin blister fluid of the affected limbs versus the unaffected limbs of CRPS patients [45]. In patients with CRPS, the levels of IL-1 β and IL-6 were significantly increased in cerebrospinal fluid (CSF), compared to other subjects [46,47]. In the blood of subjects with painful neuropathy, TNF- α levels were doubled, compared to healthy subjects and those with non-painful neuropathy [48]. IL-1 β can modulate the transmission of sensory neurons because it increases the release of substance P [49,50].

Thus, CRPS type I is associated with high levels of IL-1 β and IL-6 in CSF, and high levels of TNF- α in the blood. Furthermore, these proinflammatory cytokines are strongly expressed after the injection of HPV vaccines (Figure 1). It is now evident that the pro-inflammatory response to the injection of the vaccine is identical, under the common citokines substrate, to the inflammatory profile of the CRPS type I. Certainly, individual predisposition and other possible interfering factors have determined who should get sick and who did not, while

| Neurologic clinic syndrome | Pro-inflammatory cytokines | | | | |
|---------------------------------|----------------------------|-------|------|-------|------|
| | IL-1β | TNF-α | IL-6 | IFN-γ | IL-8 |
| Pathological pain [39] | Yes | Yes | Yes | Yes | - |
| Peripheral neuropatic pain [40] | Yes | Yes | - | - | - |
| Hyperalgesia [41] | Yes | - | - | - | - |
| CRPS [46-48] | Yes | Yes | Yes | - | Yes |
| Chronic fatigue [55,56] | Yes | Yes | Yes | - | - |
| CFS/ME [55,56] | Yes | Yes | - | - | - |
| POTS [54] | - | - | Yes | - | - |

Table 2: Neurologic clinic syndrome and pro-inflammatory cytokines.

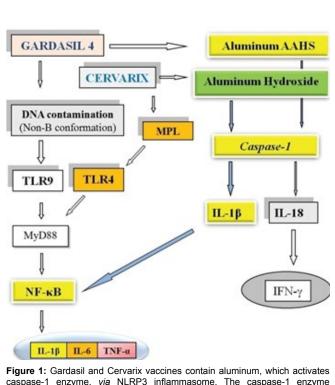


Figure 1: Gardasil and Cervarix vaccines contain aluminum, which activates caspase-1 enzyme, via NLRP3 inflammasome. The caspase-1 enzyme converts the pro-interleukins 1 β and 18 in their active forms. IL-18 determines the production of IFN-y. IL-1 β represents a cytokine that controls the local pro-inflammatory cascade and contributes to activate the transcription factor NF-kB. The Cervarix adjuvant AS04 contains Aluminum Hydroxide and MPL. The second one stimulates the TLR4. Gardasil 4 vaccine is contaminated with foreign DNA in non-B conformation [145], which activates TLR9. TLR9. TLR9 act through the adapter protein MyD88 that acts by increasing the activity of NF-kB, which then increases the expression and secretion of IL-1 β , IL-6 and TNF- α .

expressing (both categories of subjects) high levels of proinflammatory cytokines after injection of HPV vaccines.

POTS in Denmark

A number of safety signals, CRPS, POTS, and chronic fatigue syndrome (CFS), have emerged with HPV vaccines, which share a similar pattern of symptomatology [51]. POTS is a heterogeneous disorder of the autonomic nervous system [52] in which a change from the supine position to an upright position causes an abnormally large increase in heart rate or tachycardia (30 bpm within 10 minutes of standing or head-up tilt). Brinth et al. [53] report the characteristics of a number of patients with a syndrome of orthostatic intolerance, headache, fatigue, cognitive dysfunction, and neuropathic pain starting in close relation to HPV vaccination. Furthermore, the diagnosis CFS/ ME may be suitable in patients with suspected side effects to the Q-HPV vaccine [31]. Sympathetic activation and parasympathetic withdrawal in POTS patients were associated with increased serum IL-6 [54].

Long-lasting fatigue in the Netherlands

Lareb [32] has received a substantial number of reports concerning long-lasting AEs after vaccination with Cervarix[®]. The most frequently reported long-lasting AE was fatigue. The relation between elevated proinflammatory cytokines and fatigue and fatiguability is well documented [55]. In patients with CFS/ME, proinflammatory cytokines, including IL-1 β and TNF- α , are elevated and are significantly associated with the severity of fatigue, a flu-like malaise, sadness and autonomic symptoms [55]. Furthermore, other authors have found high levels of pro-inflammatory cytokines, such as IL-6 and TNF- α , in CFS/ME patients [56]. IL-1 β , IL-6 and TNF- α are strongly expressed after the injection of HPV vaccines, and in patients with CFS/ME, the same proinflammatory cytokines are elevated [55,56].

Microglia

It is now accepted that microglia are derived from mesodermal/ mesenchymal tissues, primarily myeloid cells from the bone marrow. These cells migrate to the Central Nervous System (CNS) during the first trimester of pregnancy and throughout the early part of the second trimester in humans [57]. A recent study by Paolicelli et al. [58] demonstrated a central role of microglia in synaptic pruning and circuit development in the developing embryonic brain. Microglia cells are heterogenous in their distribution [59]. Microglia cells along brain-blood vessels are often found in an activated state and form a particular immunological barrier for the brain in conjunction with the blood-brain barrier (BBB). Microglia are also concentrated in sites of incomplete BBB function, such as the circumventricular organs (CVOs), or the organum vasculosum of the lamina terminalis, subcommissural organ, subfornical organ, area postrema, posterior pituitary, median eminence, pineal, and choroid plexus, as these are entry sites of bloodborne invaders and even larger molecules [60]. Microglia contain receptors for a number of cytokines, both proinflammatory and antiinflammatory. One of the critical types of cytokine receptors for microglia activation is the IL-1 β receptors, which includes the subtypes IL1RI, IL-1RII, and IL-RIII [61]. IL-1 β activates brain microglia. The BBB has an energy-dependent, saturable, carrier-mediated transport system for cytokines, primarily IL-1, IL-6, and TNF- α [62,63]. When endothelial cells making up the BBB come into contact with these peripheral cytokines, they secrete various immune molecules into the brain parenchyma, including NO, prostaglandin E2, IL-1, and IL-6, all proinflammatory cytokines known to affect neurological function [64]. It is now accepted that peripheral inflammation and immune activation secondarily effect brain function during the infectious process [65]. Microglial activation is quite rapid following systemic immune activation, usually within minutes and results in immunoexcitotoxicity. This secondary immune process has been named sickness behavior and is characterized by anorexia, hypersomnia, lethargy, reduced social interaction, reduced cognitive function, and weakness.

Microglia primed

Microglia are very long-living cells [66,67]. Microglia can switch from a resting phenotype to a primed state by an initial immune stimulus that is not excessively intense. For example, a mild head injury or episode of hypoxia can switch microglia from its resting state to a functional condition in which the enzymes and genetic activation is upregulated, but the active immune molecules, primarily proinflammatory cytokines and chemokines, are not released [57]. NADPH oxidase is essential for microglial priming. NADPH oxidase primarily produces oxygen radicals and inducible nitric oxide synthase (iNOS) generates nitrogen radicals, which when combined forms the very powerful reactive nitrogen species (RNS) peroxynitrite [68].

With a second immune stimulus, these primed microglia began to release proinflammatory cytokines and chemokines in much higher concentrations than that of not primed microglia. Systemic immune stimulation can prime brain microglia, which means that either subsequent brain disturbances or systemic immune activation would trigger a magnified immune response within the brain. Immune events throughout life, exposure to neurotoxic metals, exposure to pesticides/ herbicides and fungicides, head injury, and other factors, can cause episodes associated with microglia priming and activation, leading to a progressive loss of neurons in the most vulnerable parts of the CNS, such as the hypothalamus, temporal lobes (hippocampus, striatal area, amygdala, and entorhinal cortex) and prefrontal cortex [57].

In the infant or small child, the priming event may come from a number of sources, such as vaccination of the mother during pregnancy or with intrauterine or early post birth infections [69,70]. In other instances, the priming event may occur with the first vaccine inoculation, usually at birth (hepatitis B). Once primed, subsequent vaccinations, especially within months of the previous inoculation, will trigger full microglial activation and in the developing brain can result in abnormal pathways development [71-74]. While natural infections can also produce this neurodestructive response, vaccinations produce higher levels of immune activation and the immune response can persist longer than natural infections – sometimes lasting years [57].

It is well-established that inflammation in the periphery can prompt immune responses in the brain [75]. Contrary to the longheld assumption that immunological memory exists only in cells of the adaptive immune system, recent evidence has indicated that also myeloid cells display memory effects [76,77]. For example, certain immune stimuli train blood monocytes to generate enhanced immune responses to subsequent immune insults [78,79]. By contrast, other stimuli induce immune tolerance/suppression of inflammatory responses to subsequent stimuli [78,80]. Innate immune memory lasts for several days in vitro and for up to three months in circulating monocytes in vivo and is mediated by epigenetic reprogramming in cultured cells, with chromatin changes also apparent in vivo [78,81,82]. However, while training may be beneficial in the periphery, owing to enhanced pathogen elimination [82-84], and tolerance may be detrimental owing to higher rates of infection resulting from immune suppression [80], training promotes, while tolerance alleviates neuropathology [85].

In summary, innate immune memory is a vital mechanism of myeloid cell plasticity that occurs in response to environmental stimuli and alters subsequent immune responses. Two types of immunological imprinting can be distinguished, training and tolerance. These are epigenetically mediated and enhance or suppress subsequent inflammation, respectively. Peripherally applied inflammatory stimuli induce acute immune training and tolerance in the brain and lead to the differential epigenetic reprogramming of microglia that persists for at least six months. Individual cytokines applied peripherally may also elicit immune memory effects in the brain [85].

Autism Spectrum Disorders (ASD) and Neuroinflammation

ASD is a pervasive neurodevelopmental condition characterized by variable impairments in communication and social interaction as well as restricted interests and repetitive behaviors. The latest estimate of the prevalence of autism in the US refers to those born in 2006 [86], for whom the overall prevalence of ASD was 16.8 per 1,000 (one out of 59). In recent years, many studies indicate that children with an ASD diagnosis have brain pathology suggestive of ongoing neuroinflammation or encephalitis in different regions of their brains. Evidence of neuroinflammation or encephalitis in ASD includes: Microglial and astrocytic activation, a unique and elevated proinflammatory profile of cytokines, and aberrant expression of NF- κ B of activated B cells. A conservative estimate based on the research suggests that at least 69% of individuals with an ASD diagnosis have microglial activation or neuroinflammation [87]. For a subgroup of children, parents report that their child had normal or near-normal

Investigations of immune system problems in ASD, including aberrations in cytokine profiles and signaling, have been increasing in recent times and are the subject of ongoing interest [95]. The immune system and the nervous system are intricately interconnected. The functional status of the immune system affects a multitude of biological processes, including brain function and development, which can be affected when the innate and adaptive immune responses are dysregulated [96]. In addition, a systematic review evaluating proinflammatory markers in almost 4,000 children and adolescents with neuropsychiatric and neurodevelopmental disorders, including ASD, identified preliminary evidence of the role of inflammation in these conditions and an association with a pro-inflammatory state [97]. Altered cytokine profiles have been consistently linked to ASD in children in the postnatal period [98]. Cytokines may influence behavior through effects on neurotransmitter function, neuroendocrine activity, neurogenesis, and alterations to brain circuitry [99]. For example, cytokines have shown to increase release and decrease reuptake of the excitatory neurotransmitter glutamate, which can result in the pathological process of excitotoxicity [100]. This evidence for abnormal cytokine profiles in ASD suggests that immune system disturbances may be active and continuous contributors to the presentation of ASD. This accumulation of evidences has acted as the catalyst for efforts to characterize possible subgroups of ASD patients who are presented with immune system abnormalities or dysfunction and altered patterns of symptom presentation [101,102].

Peripheral cytokine signals are thought to access the brain through three pathways: Humoral (with antibody involvement), neural, and cellular [95,99]. These communication pathways involve at least five mechanisms: (1) passage of cytokines through leaky regions of the blood-brain barrier; (2) active transport *via* saturable cytokinespecific transport molecules on brain endothelium; (3) activation of endothelial cells, which release second messengers within the brain parenchyma; (4) transmission of cytokine signals *via* afferent nerve fibers, including the vagus; and (5) entry into the brain parenchyma of peripherally-activated monocytes which release cytokines. An alternate communication pathway has recently been proposed. It is based on the groundbreaking work by Louveau and colleagues [103] who identified functional lymphatic vessels, in the CNS, that carry fluid and immune cells from the cerebrospinal fluid, and in doing so, they discovered a pathway for immune cells to exit the CNS.

The entry of peripheral cytokines into the brain determines different effects. The brain recognizes cytokines such as the pro-inflammatory cytokines IL-1 α , IL-1 β , TNF- α , and IL-6 as molecular signals of sickness [104]. Furthermore, TNF- α , IL-6, and IL-1 β can cross the blood-brain barrier and act on the hypothalamus where they promote fever and sickness behavior [105]. Elevated IL-1 β and IL-6 have been associated with increased stereotypical behaviors [106]. Dysregulation of IL-1 β , a pro-inflammatory cytokine expressed early in an immune response, is implicated in impairments in memory and learning [107]. IL-1 β induces and inhibits neural progenitor cell proliferation in the CNS, which can contribute to region-specific deviant brain growth in ASD [108]. Children and adults with autism have increased plasma IL-1 β levels [109-111]. Compared to monocytes of control subjects, monocytes from subjects with ASD produce excessive amounts of IL-1 β after exposure to lipopolysaccharides [112,113]. IL-1 β induces

proliferation of neural progenitor in some brain areas, while inhibiting it in others [114]. This ability may contribute to the genesis of the observed areas of excessive growth and reduced growth in the brains of individuals with autism [107]. IL-1 is involved in the most sophisticated brain processes, its induction occurs in the hippocampus during learning processes and is essential to maintain long-term potentiation (LTP), but at higher doses, as encountered in pathological conditions, IL-1 inhibits LTP [115]. Both hypersecretion and reduced secretion of IL-1 β are associated with impaired memory and language [116-118].

In summary, IL-1 β participates in neurological processes and plays key role in the pathology and healing of the central nervous system. Normal levels of IL-1 β and its IL-1ra receptor antagonist are necessary to achieve normal development and normal brain function.

 $TNF-\alpha$ is a central regulator of inflammation and is elevated in the cerebrospinal fluid of children with ASD [119].

High level of IL-6 in ASD, both centrally and peripherally, has been frequently reported [106,120,121]. IL-6 is typically regarded as a proinflammatory cytokine and has been identified as a cytokine the brain recognizes as a molecular signal of sickness [104]. However, it also has regenerative or anti-inflammatory activity, and is involved in the regulation of metabolic and neural processes [122]. In a mouse model with elevated IL-6 in the brain, Wei and colleagues [123] have shown that IL-6 can modulate autism-like behaviors through impairments of synapse formation, dendritic spine development, and neuronal circuit balance. Immunocytochemical studies have identified marked activation of microglia and astroglia associated with the increased production of two cytokines by neuroglia, macrophage chemoattractant protein (MCP)-1, and TGF-B1 [124]. In addition, a unique profile of pro-inflammatory cytokines has been identified in cerebrospinal fluid [124]. Another postmortem study demonstrated also significant increases in pro-inflammatory and Th1 cytokines relative to matched controls [120].

Altogether, ASD is recognized as having an inflammatory component. There is an association between ASD and neuroinflammation in anterior regions of the neocortex [124-126] resulting from activation of microglia and astrocytes [127]. Gene networks involved in immune processes are overexpressed in the brain of individuals with ASD [128,129]. Postmortem brain samples from patients with ASD display neuroglial activation and inflammatory markers in cerebrospinal fluid.

NF-kB mediates regulation of immune response by inducing the expression of inflammatory cytokines and chemokines, establishing a feedback mechanism that can produce chronic or excessive inflammation. NF-KB is aberrantly expressed in orbitofrontal cortex in patients with ASD, as part of a putative molecular cascade leading to inflammation, especially of resident immune cells in brain regions associated with the behavioral and clinical symptoms of ASD [130]. The implication of the NF-KB signalling pathway in ASD further supports a potential role for neuroinflammation [131]. Immune pathways are activated by proinflammatory cytokines such as TNF-a and IL-6 that stimulate the nuclear translocation of various transcription factors, including NF-KB that subsequently results in the potentiation of the immune response [132]. Cytokines, chemokines, and reactive oxygen species are among a number of key mediators that induce NF-KB by activating IkB kinases [133]. These phosphorylate IkBa, leading to its poly ubiquitination and degradation [134], allows NF-KB to migrate to the nucleus, where it activates the transcription of various proinflammatory genes.

Aberrant levels of proinflammatory cytokines (IL-6, TNF-a, and

MCP-1), not only in brain specimens and cerebrospinal fluid [135], but also in amniotic fluid [136], indicate an active inflammatory process both in children and adults with ASD. Cytokines and chemokines are pleiotrophic proteins that coordinate the host response to infection as well as mediate normal, ongoing communication between cells of nonimmune tissues, including the nervous system [137]. As a consequence of this dual role, cytokines induced in response to adverse stimuli (i.e. maternal infection or prenatal hypoxia) can profoundly impact fetal neurodevelopment. Microglia play a critical role in the pruning of synapses, thus providing a potential bridge between the atypical synaptic pruning and the immune dysregulatory hypotheses of ASD [138].

Ultimately, the pro-inflammatory cytokines produced by the activation of the peripheral immune system, also determine effects in central nervous system and in many cases of ASD there is evidence of neuroinflammation. Pro-inflammatory cytokines, including IL-1 β , IL-6, and TNF- α , appear to be at the forefront in the communication between the immune and the nervous system, playing dual roles in mediating physiological and neuroprotective roles in normal brain function or being detrimental and associated with brain diseases, especially when present at elevated concentrations [139].

Since in the first year of life, in the industrialized countries, the immune system is activated more by the incoming number of vaccinations than by the number of infections, we must begin to think that the pro-inflammatory cytokines released after vaccine injections can produce microglia activation which can cause neuroinflammation.

Discussion

Cytokines, together with neurotransmitters and hormones, are signaling molecules possessing unique immunomodulatory functions. Virtually, they can influence every physiological system including neuroendocrine interactions, neurotransmitter metabolism and neuroplasticity, thereby affecting behavioral and cognitive functioning [140]. Cytokines take center stage in orchestrating immune responses [141]. Injection of the vaccines results in a strong expression of proinflammatory cytokines. Macrophages secrete pro-inflammatory cytokines such as IL-1 β , TNF- α , and IL-6, when activated. It seems that variations in vaccine-induced cytokine responses are modulated by genetic polymorphisms in many cytokine and cytokine receptor genes [142]. Cytokines are cell-to-cell messangers similar to hormones with the stronger activity in the microenvironmental of the cells that secrete them [143]. They act in most cases at shorter distances (with exceptions such as IL-1, IL-6 and TNF). However, cytokines penetrate most tissues, being delivered by migration of white blood cells of the hematopoietic tissue, which virtually permeates all other tissues in vertebrates. Furthemore, the immune competent cells are one of the largest sources of cytokines, that being capable to migrate in almost all tissues of the body, represent moving regulators of the local microenvironment [143].

We hypothesized a link between vaccinations and neuroinflammation. The peripheral pro-inflammatory cytokines (IL-1 β , IL-6, and TNF- α), expressed after the injection of all vaccines, can reach the brain and can cause neuroinflammation after microglia activation. Depending on the age and type of vaccines, neuroinflammation may produce AEs, such as those following HPV vaccination. Elevated pro-inflammatory cytokines, particularly TNF- α , have been described in studies regarding the cytokines profile in autistic children. IL-1 β represents a cytokine that controls the local pro-inflammatory cascade and thereby affects the balance between protective immunity and destructive inflammation. A subgroup of children with ASD have

Page 7 of 11

developed neuroinflammation. Several postmortem studies have confirmed the activation of microglia and neuroinflammation. A recent study shows the presence of aluminium in brain tissue in ASD. Besides, aluminium was also found in microglia cells [144]. Aluminium from vaccines is redistributed to numerous organs including brain, where it accumulates. Each vaccine adds to this tissue different level of aluminium. Aluminum, like mercury, activates microglia leading to chronic brain inflammation and neurotoxicity.

Gardasil and Cervarix vaccines (Figure 1) contain aluminum, which activates caspase-1 enzyme, *via* NLRP3 inflammasome. The caspase-1 enzyme converts the pro-interleukins 1 β and 18 in their active forms. IL-18 determines the production of IFN- γ . IL-1 β represents a cytokine that controls the local pro-inflammatory cascade and contributes to activate the transcription factor NF- κ B. The Cervarix adjuvant AS04 contains Aluminum Hydroxide and MPL. The second one stimulates the TLR4. Gardasil 4 vaccine is contaminated with foreign DNA in non-B conformation [145], which activates TLR9. TLRs act through the adapter protein MyD88 that acts by increasing the activity of NF- κ B, which then increases the expression and secretion of IL-1 β , IL-6 and TNF- α .

Thus, there is a strong immune stimulation and a strong production of pro-inflammatory cytokines, including IL-1 β IL-6 and TNF- α , which are capable of exerting effects at a distance from the production site.

In Figure 2, the mechanism of action of the aluminum is always represented, but a new anti-meningococcal B vaccine produces the activation of the TLR- 2 and 4. The OMV vesicles contain lipoproteins that activate the TLR2, and LPS that activate TLR4. The strong production of peripheral pro-inflammatory cytokines is capable of producing microglia activation and neuroinflammation.

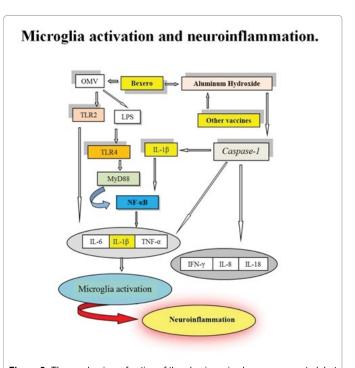


Figure 2: The mechanism of action of the aluminum is always represented, but a new anti-meningococcal B vaccine produces the activation of the TLR- 2 and 4. The OMV vesicles contain lipoproteins that activate the TLR2, and LPS that activate TLR4. The strong production of peripheral pro-inflammatory cytokines is capable of producing microglia activation and neuroinflammation.

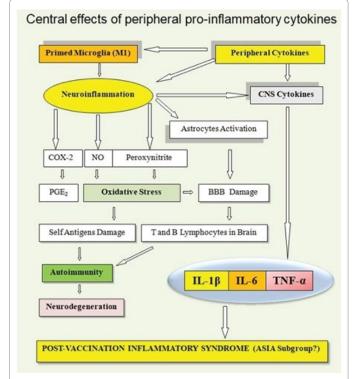


Figure 3: On the right side, you can see that the peripheral pro-inflammatory cytokines, expressed after the injection vaccines, can reach the brain and, apart from neuroinflammation, can cause a post-vaccination inflammatory syndrome [29], as in the case of HPV vaccines. If a neuroinflammation is present, it could be followed by autoimmune reactions and neurodegeneration. Peripheral cytokines can produce primed microglia and the inflammatory phenotype M1 participating in the neuroinflammation. The neuroinflammation increases production of pro-inflammatory cytokines, activates astrocytes, produces an oxidative stress and astrocytes activation cause a rupture of the BBB that eases the entry of the T and B lymphocytes in the brain. Oxidative stress also produces damage to self-antigens and may help to produce autoimmunity and neurodegenerative diseases.

On the right side of Figure 3, you can see that the peripheral proinflammatory cytokines, expressed after the injection vaccines, can reach the brain and, apart from neuroinflammation, can cause a postvaccination inflammatory syndrome [29], as in the case of HPV vaccines. If a neuroinflammation is present, it could be followed by autoimmune reactions and neurodegeneration. Peripheral citokines can produce primed microglia and the inflammatory phenotype M1 participating in the neuroinflammation. The neuroinflammation increases production of pro-inflammatory cytokines, activates astrocytes, produces an oxidative stress and increases the production of prostaglandins in the brain. Oxidative stress and astrocytes activation cause a rupture of the BBB that eases the entry of the T and B lymphocytes in the brain. Oxidative stress also produces damage to self-antigens and may help to produce autoimmunity and neurodegenerative diseases.

Conclusion

The existence of extensive lines of communication between the nervous system and immune system represents a fundamental principle underlying neuroinflammation. Immune memory in the brain is an important modifier of neuropathology. Systemic inflammation generates signals that communicate with the brain and lead to changes in metabolism and behavior, with microglia assuming a pro-inflammatory phenotype. Two types of immunological imprinting can be distinguished: Training and tolerance. These are epigenetically mediated and enhance or suppress subsequent inflammation, respectively. Peripherally applied inflammatory stimuli induce acute immune training and tolerance in the brain and lead to differential epigenetic reprogramming of brain-resident macrophages (microglia) that persists for at least six months.

The molecular mechanisms presented here demonstrate how peripheral cytokines, expressed after vaccination, can cause neuroinflammation in some subjects, after microglia activation, depending on the immunogenetic background and the innate immune memory.

The effects produced by the activation of the microglia, and the subsequent neuroinflammation, are diversified according to age: before the first two years of life they can contribute to producing ASD (in some subjects with ASD there is neuroinflammation and aluminium accumulation in the brain); while a different neurological symptomatology can arise in girls vaccinated with HPV vaccines. A post-vaccination inflammatory syndrome can explain the adverse reactions to HPV vaccines (ASIA Subgrup?). Indeed, IL-1 β causes pathological pain and fatigue and is elevated in peripheral neuropathic pain and CRPS type I. Increased levels of IL-1 β , IL-6, and TNF- α in the brain cause cognitive impairment, sleep disorders, and reduced motor activity.

Regarding the possible relationships between HPV vaccines and CRPS type I, at the molecular level, the cytokines typical of CRPS type I environment, with high levels of IL-1 β , IL-6, and TNF- α [46-48], it is faithfully reproduced by the injection of HPV vaccines. Whereas in patients with POTS, sympathetic activation and parasympathetic withdrawal were associated with increased serum IL-6 [54]. IL-1 β , IL-6 and TNF- α are strongly expressed after the injection of HPV vaccines, and in patients with CFS/ME, the same proinflammatory cytokines are elevated [55,56].

Suzuki and Hosono [146] did not find an association between HPV vaccine and reported post-vaccination symptoms in Japanese young women. The survey tool was an anonymous postal questionnaire.

As is evident, epidemiology uses different investigative tools than molecular biology. In the case of HPV vaccine AEs, molecular biology demonstrates what epidemiology does not detect.

Disclosure of Potential Conflicts of Interest

The authors declare that there are no competing interests regarding the publication of this paper.

References

- Mitkus RJ, King DB, Hess MA, Forshee RA, Walderhaug MO (2011) Updated aluminum pharmacokinetics following infant exposures through diet and vaccination. Vaccine 29: 9538-9543.
- Priest ND (2004) The biological behaviour and bioavailability of aluminium in man, with special reference to studies employing aluminium-26 as a tracer: Review and study update. J Environ Monit 6: 375-403.
- Movsas TZ, Paneth N, Rumbeiha W (2013) Effect of routine vaccination on aluminum and essential element levels in preterm infants. JAMA Pediatr 167: 870-872.
- Karwowski MP, Stamoulis C, Wenren LM, Faboyede GM, Quinn N, et al. (2018) Blood and hair aluminum levels, vaccine history, and early infant development: A cross-sectional study. Acad Pediatr 18: 161-165.
- Khan Z, Combadière C, Authier FJ, Itier V, Lux F, et al. (2013) Slow CCL2dependent translocation of biopersistent particles from muscle to brain. BMC Medicine 11: 99.

- Gherardi RK, Aouizerate J, Cadusseau J, Yara S, Authier FJ (2016) Aluminum adjuvants of vaccines injected into the muscle: Normal fate, pathology and associated disease. Morphologie 100: 85-94.
- 7. Gherardi RK, Eidi H, Crépeaux G, Authier FJ, Cadusseau J (2015) Biopersistence and brain translocation of aluminium adjuvants of vaccines. Front Neurol 6: 4.
- EFSA (European Food Safety Authority) (2008) Safety of aluminium from dietary intake, scientific opinion of the panel on food additives, flavourings, processing aids and food contact materials (AFC) EFSA J 6: 754.
- Yang M, Jiang L, Huang H, Zeng S, Qiu F, et al. (2014) Dietary exposure to aluminium and health risk assessment in the residents of shenzhen, China. PLoS ONE 9: e89715.
- 10. Committee on toxicity of chemicals in food, consumer products and the environment, UK.
- 11. Bexero vaccine (2013), European Union.
- 12. Prevenar 13 vaccine (2017), Italy.
- 13. Infanrix-Hexa vaccine (2017), Italy.
- Mosca F, Tritto E, Muzzi A, Monaci E, Bagnoli F, et al. (2008) Molecular and cellular signatures of human vaccine adjuvants. Proc Natl Acad Sci USA 105: 10501-10506.
- 15. McKee AS, Munks MW, MacLeod MK, Fleenor CJ, Van Rooijen N, et al. (2009) Alum induces innate immune responses through macrophage and mast cell sensors, but these sensors are not required for alum to act as an adjuvant for specific immunity. J Immunol 183: 4403-4414.
- Pétrilli V, Dostert C, Muruve DA, Tschopp J (2007) The inflammasome: A danger sensing complex triggering innate immunity. Curr Opin Immunol 19: 615-622.
- Stylianou E, Saklatvala J (1998) Interleukin-1. Int J Biochem Cell Biol 30: 1075-1079.
- CDC (Center for Disease Control and Prevention), (2017) Vaccine site, Atlanta, Georgia.
- Flacco ME, Manzoli L, Rosso A, Marzuillo C, Bergamini M, et al. (2018) Immunogenicity and safety of the multicomponent meningococcal B vaccine (4CMenB) in children and adolescents: A systematic review and meta-analysis. Lancet Infect Dis 18: 461-472.
- Soeters HM, Whaley M, Alexander-Scott N, Kanadanian KV, MacNeil JR, et al. (2017) Meningococcal carriage evaluation in response to a serogroup b meningococcal disease outbreak and mass vaccination campaign at a collegerhode island, 2015-2016. CID 64: 1115-1122.
- Breakwell L, Whaley M, Khan UI, Bandy U, Alexander-Scott N, et al. (2018) Meningococcal carriage among a university student population-United States, 2015. Vaccine 36: 29-35.
- 22. McNamara LA, Thomas JD, MacNeil J, Chang HY, Day M, et al. (2017) Meningococcal carriage following a vaccination campaign with MenB-4C and MenB-FHbp in response to a university Serogroup B meningococcal disease outbreak-Oregon, 2015-2016. J Infect Dis 216: 1130-1140.
- Herrin DM, Coates EE, Costner PJ, Kemp TJ, Nason MC, et al. (2014) Comparison of adaptive and innate immune responses induced by licensed vaccines for human papillomavirus. Hum Vaccines Immunother 10: 3446-3454.
- 24. Arbyn M, Xu L, Simoens C, Martin-Hirsch PP (2018) Prophylactic vaccination against human papillomaviruses to prevent cervical cancer and its precursors. Cochrane Database of Systematic Reviews 5: 1-252.
- 25. VRBPAC (2006) Background Document. Gardasil™ HPV Quadrivalent Vaccine, VRBPAC Meeting, USA.
- Fischer S, Bettstetter M, Becher A, Lessel M, Bank C, et al. (2016) Shift in prevalence of HPV types in cervical cytology specimens in the era of HPV vaccination. Oncol Lett 12: 601-610.
- Guo F, Hirth JM, Berenson AB (2015) Comparison of HPV prevalence between HPV-vaccinated and non-vaccinated young adult women (20-26 years). Hum Vaccines Immunother 11: 2337-2344.
- Ozawa K, Hineno A, Kinoshita T, Ishihara S, Ikeda SI (2017) Suspected adverse effects after human papillomavirus vaccination: A temporal relationship between vaccine administration and the appearance of symptoms in japan. Drug Saf 40: 1219-1229.
- Giannotta G (2016) Post-vaccination inflammatory syndrome: A new syndrome. 10th international congress on autoimmunity, Leipzig, Germany.

- 30. Kinoshita T, Abe RT, Hineno A, Tsunekawa K, Nakane S, et al. (2014) Peripheral sympathetic nerve dysfunction in adolescent Japanese girls following immunization with the human papillomavirus vaccine. Intern Med 53: 2185-2200.
- Brinth LS, Pors K, Theibel AC, Mehlsen J (2015) Orthostatic intolerance and postural tachycardia syndrome as suspected adverse effects of vaccination against human papilloma virus. Vaccine 33: 2602-2605.
- 32. Lareb (2015) Long-lasting adverse events following immunization with Cervarix®, Netherlands.
- European Medicines Agency assessment report. Review under Article 20 of Regulation (EC) No 726/200411.
- Viljoen M, Panzar A (2004) Sickness behaviour: Causes and effects. SA Fam Pract 45: 15-18.
- Marty V, El Hachmane M, Amedee T (2008) Dual modulation of synaptic transmission in the nucleus tractus solitarius by prostaglandin E2 synthesized downstream of IL-1beta. Eur J Neurosci 27: 3132-3150.
- Hwang SY, Jung JS, Kim TH, Lim SJ, Oh ES, et al. (2006) Ionizing radiation induces astrocyte gliosis through microglia activation. Neurobiol Dis 21: 457-467.
- 37. Omoigui S (2007) The biochemical origin of pain: The origin of all pain is inflammation and the inflammatory response. part 2 of 3, Inflammatory profile of pain syndromes. Med hypotheses 69: 1169-1178.
- Zouikr I, Bartholomeusz MD, Hodgson DM (2016) Early life programming of pain: Focus on neuroimmune to endocrine communication. J Transl Med 14: 123.
- Zhang JM, An J (2007) Cytokines, inflammation and pain. Int Anesthesiol Clin 45: 27-37.
- Clark AK, Old EA, Malcangio M (2013) Neuropathic pain and cytokines: Current perspectives. J Pain Res 6: 803-814.
- Binshtok AM, Wang H, Zimmermann K, Amaya F, Vardeh D, et al. (2008) Nociceptors are interleukin-1beta sensors. J Neurosci. 28: 14062-14073.
- Sebastin SJ (2011) Complex regional pain syndrome. Indian journal of plastic surgery : Official publication of the association of plastic surgeons of India. 44: 298-307.
- Veldman PH, Reynen HM, Arntz IE, Goris RJ (1993) Signs and symptoms of reflex sympathetic dystrophy: prospective study of 829 patients. Lancet 342: 1012-1016.
- 44. Schwarzer A, Maier C (2010) Complex regional pain syndrome. In: Kopf A, Patel N, editors. Guide to pain managment in low resource settings. Seattle: IASP Press pp. 249-254.
- 45. Huygen FJ, De Bruijn AG, De Bruin MT, Groeneweg JG, Klein J, et al. (2002) Evidence for local inflammation in complex regional pain syndrome type 1. Mediators Inflamm 11: 47-51.
- Alexander GM, Van Rijn MA, Van Hilten JJ, Perreault MJ, Schwartzman RJ (2005) Changes in cerebrospinal fluid levels of pro-inflammatory cytokines in CRPS. Pain 116: 213-219.
- Parkitny L, McAuley JH, Di Pietro F, Stanton TR, O'connell NE, et al. (2013) Inflammation in complex regional pain syndrome: A systematic review and meta-analysis. Neurology 80: 106-117.
- Taha R, Blaise GA (2012) Update on the pathogenesis of complex regional pain syndrome: Role of oxidative stress. Can J Anaesth 59: 875-881.
- Malcangio M, Bowery NG, Flower RJ, Perretti M (1996) Effect of interleukin-1beta on the release of substance P from rat isolated spinal cord. Eur J Pharmacol 299: 113-118.
- Inoue A, Ikoma K, Morioka N, Kumagai K, Hashimoto T, et al. (1999) Interleukin-1beta induces substance P release from primary afferent neurons through the cyclooxygenase-2 system. J Neurochem 73: 2206-2213.
- Chandler RE, Juhlin K, Fransson J, Caster O, Edwards IR, et al. (2017) Current safety concerns with human papillomavirus vaccine: A cluster analysis of reports in vigibase[®]. Drug Saf 40: 81-90.
- Grubb BP, Kanjwal Y, Kosinski DJ (2006) The postural tachycardia syndrome: A concise guide to diagnosis and management. J Cardiovasc Electrophysiol 17: 108-112.
- Brinth L, Pors K, Hoppe AAG, Badreldin I, Mehlsen J, et al. (2015) Is chronic fatigue syndrome/myalgic encephalomyelitis a relevant diagnosis in patients

with suspected side effects to human papilloma virus vaccine? IJVV 1: 00003.

- 54. Okamoto LE, Raj SR, Gamboa A, Shibao CA, Arnold AC, et al. (2015) Sympathetic activation is associated with increased IL-6, but not CRP in the absence of obesity: lessons from postural tachycardia syndrome and obesity. Am J Physiol Heart Circ Physiol 309: H2098-H2107.
- 55. Maes M, Twisk FN, Kubera M, Ringel K et al. (2012) Evidence for inflammation and activation of cell-mediated immunity in myalgic encephalomyelitis/chronic fatigue syndrome (ME/CFS): Increased interleukin-1, tumor necrosis factor-α, PMN-elastase, lysozyme and neopterin. J Affect Disord 136: 933-939.
- Fletcher MA, Zeng XR, Barnes Z, Levis S, Klimas NG (2009) Plasma cytokines in women with chronic fatigue syndrome. J Transl Med 7: 96.
- 57. Blaylock RL (2013) Immunology primer for neurosurgeons and neurologists part 2: Innate brain immunity. Surg Neurol Int 4: 118.
- Paolicelli RC, Bolasco G, Pagani F, Maggi L, Scianni M, et al. (2011) Synaptic pruning by microglia is necessary for normal brain development. Science 333: 1456-1458.
- Kettenmann H, Hanisch UK, Noda M, Verkhratsky A et al. (2011) Physiology of microglia. Physiol Rev 91: 461-553.
- Bechmann I, Galea I, Perry VP (2002) What is the blood-brain barrier (not)? Trend Immunol 28: 5-11.
- Lee YB, Nagal A, Kim SU. (2002) Cytokiness, chemokines, and cytokine receptors in human microglia. J Neurosci Res 69: 94-103.
- Banks WA, Kastin AJ (1991) Blood to brain transport of interleukin links the immune and central nervous system. Life Sci 48: 117-121.
- Gutierrez EG, Banks WA, Kastin AJ (1993) Murine tumor necrosis factor alpha is transported from blood to brain in the mouse. J Neuroimmunol 47: 169-176.
- 64. Fabry Z, Fitzsimmons KM, Herlein JA, Moninger TO, Dobbs MB et al. (1993) Production of the cytokines interleukin 1 and 6 by murine brain microvessel endothelium and smooth muscle pericytes. J Neuroimmunol 47: 23-34.
- 65. Dantzer R, O'Connor JC, Freund GG, Johnson RW, Kelley KW, et al. (2008) From inflammation to sickness and depression: When the immune system subjugates the brain. Nat Rev Neurosci 9: 46-56.
- Tay TL, Mai D, Dautzenberg J, Fernández-Klett F, Lin G, et al. (2017) A new fate mapping system reveals context-dependent random or clonal expansion of microglia. Nat Neurosci 20: 793-803.
- 67. Füger P, Hefendehl JK, Veeraraghavalu K, Wendeln AC, Schlosser C et al. (2017) Microglia turnover with aging and in an Alzheimer's model via long-term in vivo single-cell imaging. Nat Neurosci 20: 1371-1376.
- Choi SH, Aid S, Kim HW, Jackson SH, Bosetti F (2012) Inhibition of NADPH oxidase promotes alternative and anti-inflammatory microglial activation during neuroinflammation. J Neurochem 120: 292-301.
- Blaylock RL (2012) Immunoexcitotoxicity as a central mechanism of autism spectrum disorders, Bentham Science Publishers, UAE.
- Blaylock RL, Strunecka A (2009) Immune-glutamatergic dysfunction as a central mechanism of the autism spectrum disorders. Curr Med Chem 16: 157-70.
- Blaylock RL (2008) A possible central mechanism in autism spectrum disorders, Part I. Altern Ther Health Med 14: 46-53.
- Blaylock RL (2009) A possible central mechanism in autism spectrum disorders, Part 2: Immunoexcitotoxicity. Altern Ther Health Med 15: 60-67.
- Aarum J, Sandberg K, Budd-Haeberlein SL, Persson MAA et al. (2003) Migration and differentiation of neural precursor cells can be directed by microglia. Proc Natl Acad Sci USA. 100: 15983-15988.
- 74. Schlett K (2006) Glutamate as a modulator of embryonic and adult neurogenesis. Curr Top Med Chem 6: 949-960.
- Perry VH, Cunningham C, Holmes C (2007) Systemic infections and inflammation affect chronic neurodegeneration. Nat Rev Immunol 7: 161-167.
- Netea MG, Latz E, Mills KH, O'neill LA (2015) Innate immune memory: A paradigm shift in understanding host defense. Nat Immunol 16: 675-679.
- Netea MG, Joosten LA, Latz E, Mills KH, Natoli G, et al. (2016) Trained immunity: A program of innate immune memory in health and disease. Science 352: 1098.
- 78. Saeed S, Quintin J, Kerstens HH, Rao NA, Aghajanirefah A, et al. (2014) Epigenetic programming of monocyte-to-macrophage differentiation and

trained innate immunity. Science 345: 1251.

- Cheng SC, Quintin J, Cramer RA, Shepardson KM, Saeed S, et al. (2014) mTOR-and HIF-1α-mediated aerobic glycolysis as metabolic basis for trained immunity. Science 345: 1250.
- Biswas SK, Lopez-Collazo E (2009) Endotoxin tolerance: New mechanisms, molecules and clinical significance. Trends Immunol 30: 475-487.
- Novakovic B, Habibi E, Wang SY, Arts RJ, Davar R, et al. (2016) β-Glucan reverses the epigenetic state of LPS-induced immunological tolerance. Cell 167: 1354-1368.
- Kleinnijenhuis J, Quintin J, Preijers F, Joosten LA, Ifrim DC, et al. (2012) Bacille Calmette-Guerin induces NOD2-dependent nonspecific protection from reinfection via epigenetic reprogramming of monocytes. Proc Natl Acad Sci 109: 17537-17542.
- Kaufman E, Sanz J, Dunn JL, Khan N, Mendonça LE, et al. (2018). BCG educates hematopoietic stem cells to generate protective innate immunity against tuberculosis. Cell 172: 176-190.
- 84. Arts RJ, Moorlag SJ, Novakovic B, Li Y, Wang SY, et al. (2018) BCG vaccination protects against experimental viral infection in humans through the induction of cytokines associated with trained immunity. Cell host microbe 23: 89-100.
- Wendeln AC, Degenhardt K, Kaurani L, Gertig M, Ulas T, et al. (2018) Innate immune memory in the brain shapes neurological disease hallmarks. Nature 556: 332.
- 86. Baio J, Wiggins L, Christensen DL, Maenner MJ, Daniels J, et al. (2018) Prevalence of Autism spectrum disorder among children aged 8 years autism and developmental disabilities monitoring network, 11 Sites, United States. MMWR Surveill Summ 67(6):1-23.
- Kern JK, Geier DA, Sykes LK, Geier MR (2016) Relevance of neuroinflammation and encephalitis in autism. Front Cell Neurosci 9: 519.
- Goldberg WA, Osann K, Filipek PA, Laulhere T, Jarvis K, et al. (2003) Language and other regression: Assessment and timing. J Autism Dev Disord 33: 607-616.
- Davidovitch M, Glick L, Holtzman G, Tirosh E, Safir MP (2000) Developmental regression in autism: Maternal perception. J Autism Dev Disord 30: 113-119.
- Hoshino Y, Kaneko M, Yashima Y, Kumashiro H, Volkmar FR, et al. (1987) Clinical features of autistic children with setback course in their infancy. Jpn J Psychiatry Neurol 41: 237-245.
- Kurita H (1985) Infantile autism with speech loss before the age of thirty months. J Am Acad Child Psychiatry 24:191-196.
- Lord C (1995) Follow up of two year olds referred for possible autism. J Child Psychol Psychiatry 36: 1365-1382.
- Rogers S, DiLalla D (1990) Age of symptom onset in young children with pervasive developmental disorders. J Am Acad Child Adolesc Psychiatr 29: 863-872.
- Tuchman RF, Rapin I (1997) Regression in pervasive developmental disorders: Seizures and epileptiform electroencephalogram correlates. Pediatrics 99: 560-566.
- Masi A, Glozier N, Dale R, Guastella AJ (2017) The immune system, cytokines, and biomarkers in autism spectrum disorder. Neuroscience bulletin 33: 194-204.
- 96. Filiano AJ, Gadani SP, Kipnis J (2015) Interactions of innate and adaptive immunity in brain development and function. Brain Res 1617: 18-27.
- Mitchell RH, Goldstein BI (2014) Inflammation in children and adolescents with neuropsychiatric disorders: A systematic review. J Am Acad Child Adolesc Psychiatry 53: 274-296.
- Goines P, Van de Water J (2010) The immune system's role in the biology of autism. Curr Opin Neurol 23: 111-117.
- Capuron L, Miller AH (2011) Immune system to brain signaling: Neuropsychopharmacological implications. Pharmacol thera 130: 226-238.
- 100. Tilleux S, Hermans E (2007) Neuroinflammation and regulation of glial glutamate uptake in neurological disorders. J Neurosci Res 85: 2059-2070.
- 101.McDougle CJ, Landino SM, Vahabzadeh A, O'Rourke J, Zurcher NR, et al. (2015) Toward an immune-mediated subtype of autism spectrum disorder. Brain Res 1617: 72-92.
- 102. Mead J, Ashwood P (2015) Evidence supporting an altered immune response

in ASD. Immunol Lett 163: 49-55.

- 103.Louveau A, Smirnov I, Keyes TJ, Eccles JD, Rouhani SJ, et al. (2015) Structural and functional features of central nervous system lymphatic vessels. Nature 523: 337-341.
- 104.Dantzer R (2009) Cytokine, sickness behavior, and depression. Immunol Allergy Clin North Am 29: 247-264.
- 105.Dantzer R (2001) Cytokine-induced sickness behavior: Where do we stand? Brain Behav Immun 15: 7-24.
- 106. Ashwood P, Krakowiak P, Hertz-Picciotto I, Hansen R, Pessah I, et al. (2011) Elevated plasma cytokines in autism spectrum disorders provide evidence of immune dysfunction and are associated with impaired behavioral outcome. Brain Behav Immun 25: 40-45.
- 107. Goines PE, Ashwood P (2013) Cytokine dysregulation in autism spectrum disorders (ASD): Possible role of the environment. Neurotoxicol Teratol 36: 67-81.
- 108. Courchesne E (2004) Brain development in autism: Early overgrowth followed by premature arrest of growth. Ment Retard Dev Disabil Res Rev 10: 106-111.
- 109. Ashwood P, Krakowiak P, Hertz-Picciotto I, Hansen R, Pessah IN, et al. (2011) Altered T cell responses in children with autism. Brain Behav Immun 25: 840-849.
- 110. Ashwood P, Anthony A, Torrente F, Wakefield AJ (2004) Spontaneous mucosal lymphocyte cytokine profiles in children with autism and gastrointestinal symptoms: Mucosal immune activation and reduced counter regulatory interleukin-10. J Clin Immunol 24: 664-673.
- 111. Suzuki K, Matsuzaki H, Iwata K, Kameno Y, Shimmura C, et al. (2011) Plasma cytokine profiles in subjects with high-functioning autism spectrum disorders. PLoS One 6: e20470.
- 112. Jyonouchi H, Sun S, Le H (2001) Proinflammatory and regulatory cytokine production associated with innate and adaptive immune responses in children with autism spectrum disorders and developmental regression. J Neuroimmunol 120: 170-179.
- Enstrom AM, Onore CE, Van de Water JA, Ashwood P (2010) Differential monocyte responses to TLR ligands in children with autism spectrum disorders. Brain Behav Immun 24: 64-71.
- 114. de la Mano A, Gato A, Alonso MI, Carnicero E, Martín C, et al. (2007) Role of interleukin-1beta in the control of neuroepithelial proliferation and differentiation of the spinal cord during development. Cytokine 37: 128-137.
- 115. Ross FM, Allan SM, Rothwell NJ (2003) A dual role for interleukin-1 in LTP in mouse hippocampal slices. J Neuroimmunol 144: 61-67.
- 116. Barrientos RM, Frank MG, Hein AM, Higgins EA, Watkins LR, et al. (2009) Time course of hippocampal IL-1 beta and memory consolidation impairments in aging rats following peripheral infection. Brain Behav Immun 23: 46-54.
- 117. Labrousse VF, Costes L, Aubert A, Darnaudéry M, Ferreira G, et al. (2009) Impaired interleukin-1beta and c-Fos expression in the hippocampus is associated with a spatial memory deficit in P2X(7) receptor-deficient mice. PLoS One 4: e6006.
- 118. Goshen I, Kreisel T, Ounallah-Saad H, Renbaum P, Zalzstein Y, et al. (2007) A dual role for interleukin-1 in hippocampal-dependent memory processes. Psychoneuroendocrinology 32: 1106-1115.
- Chez MG, Dowling T, Patel PB, Khanna P, Kominsky M, et al. (2007) Elevation of tumor necrosis factor-alpha in cerebrospinal fluid of autistic children. Pediatr Neurol 36: 361-365.
- 120.Li X, Chauhn A, Shiekh AM, Patil S, Chauhn V, et al. (2009) Elevated immune response in the brain of autistic patients. J Neuroimmunol 207: 111-116.
- 121. Wei H, Zou H, Sheikh AM, Malik M, Dobkin C, et al. (2011) IL-6 is increased in the cerebellum of autistic brain and alters neural cell adhesion, migration and synaptic formation. J Neuroinflammation 8: 52.
- 122. Scheller J, Chalaris A, Schmidt-Arras D, Rose-John S (2011) The pro and anti-inflammatory properties of the cytokine interleukin-6. Biochim Biophys Acta 1813: 878-888.
- 123. Wei H, Chadman KK, McCloskey DP, Sheikh AM, Malik M, et al. (2012) Brain IL-6 elevation causes neuronal circuitry imbalances and mediates autism-like behaviors. Biochim Biophys Acta 1822: 831-842.
- 124. Vargas DL, Nascimbene C, Krishnan C, Zimmerman AW, Pardo CA, et al. (2005) Neuroglial activation and neuroinflammation in the brain of patients with autism. Ann Neurol 57: 67-81.

- 125.Pardo CA, Vargas DL, Zimmerman AW (2005) Immunity, neuroglia and neuroinflammation in autism. Int Rev Psychiatry 17: 485-495.
- 126.Zimmerman AW, Jyonouchi H, Comi AM, Connors SL, Milstien S, et al. (2005) Cerebrospinal fluid and serum markers of inflammation in autism. Pediatr Neurol 33: 195-201.
- 127.Anderson AA, Ushakov DS, Ferenczi MA, Mori R, Martin P, et al. (2008) Morphoregulation by acetylcholinesterase in fibroblasts and astrocytes. J Cell Physiol 215: 82-100.
- Voineagu I, Wang X, Johnston P, Lowe JK, Tian Y, et al. (2011) Transcriptomic analysis of autistic brain reveals convergent molecular pathology. Nature 474: 380-384.
- 129. Voineagu I, Eapen V (2013) Converging pathways in autism spectrum disorders: Interplay between synaptic dysfunction and immune responses. Front Hum Neurosci 7: 738.
- 130. Young AMH, Campbell E, Lynch S, Suckling J, Powis SJ, et al. (2011) Aberrant NF-KappaB expression in autism spectrum condition: A mechanism for neuroinflammation. Frontiers in Psychiatry 2: 27.
- 131. Young AMH, Chakrabarti B, Roberts D, Lai MC, Suckling J, et al. (2016) From molecules to neural morphology: Understanding neuroinflammation in autism spectrum condition. Mol Autism 7: 9.
- Perkins ND (2004) NF-kappaB: Tumor promoter or suppressor? Trends Cell Biol 14: 64-69.
- Pahl HL (1999) Activators and target genes of Rel/NF-kappaB transcription factors. Oncogene 18: 6853-6866.
- 134.Ghosh S, Karin M (2002) Missing pieces in the NF-kappaB puzzle. Cell 109: S81-S96.
- 135. Schwarz E, Guest PC, Rahmoune H, Wang L, Levin Y, et al. (2011) Sexspecific serum biomarker patterns in adults with Asperger's syndrome. Mol Psychiatry 16: 1213-1220.
- 136.Abdallah MW, Larsen N, Grove J, Bonefeld-Jørgensen EC, Nørgaard-Pedersen B, et al. (2013) Neonatal chemokine levels and risk of autism spectrum disorders: Findings from a Danish historic birth cohort follow-up study. Cytokine 61: 370-376.
- 137. Deverman BE, Patterson PH (2009) Cytokines and CNS development. Neuron 64: 61-78.
- Paolicelli RC, Gross CT (2011) Microglia in development: Linking brain wiring to brain environment. Neuron Glia Biol 7: 77-83.
- 139. Strunecka A, Blaylock RL, Patocka J, Strunecky O (2018) Immunoexcitotoxicity as the central mechanism of etiopathology and treatment of autism spectrum disorders: A possible role of fluoride and aluminum. Surg Neurol Int 9: 74.
- 140.Di Benedetto S, Müller L, Wenger E, Düzel S, Pawelec G, et al. (2017) Contribution of neuroinflammation and immunity to brain aging and the mitigating effects of physical and cognitive interventions. Neurosci Biobehav Rev 75: 114-128.
- 141.Essen C (2016) Envinronmental influences on the immune system. Springer pp: 1-17.
- 142. Ovsyannikova IG, Haralambieva IH, Kennedy RB, Pankratz VS, Vierkant RA, et al. (2012) Impact of cytokine and cytokine receptor gene polymorphisms on cellular immunity after smallpox vaccination. Gene 510: 59-65.
- 143. Dembic Z (2015) The cytokines of the immune system. The role of cytokines in disease related to immune responce. Academic Press pp: 143-239.
- 144. Mold M, Umar D, King A (2018) Aluminium in brain tissue in autism. J Trace Med Biol 46: 76-82.
- 145.Lee SH (2012) Detection of human papillomavirus L1 gene DNA fragments in postmortem blood and spleen after Gardasil[®] vaccination. A case report. Adv Biosci Biotech 3: 1214-1224.
- 146. Suzuki S, Hosono A (2018) No association between HPV vaccine and reported post-vaccination symptoms in Japanese young women: Results of the Nagoya study. Papillomavirus Res 5: 96-103.