Toxicological Evaluations of the Crude Extracts and Fractions of *Moringa stenopetala* Leaves in Liver and Kidney of Rats

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Abstract

**Background:** *Moringa stenopetala* is a medicinal plant that has been used in Ethiopian traditional medicine as a remedy for treatment of hypertension, diabetes, malaria and stomach pain.

**Objective:** The aim of this study was to assess the safety of the extracts and fractions of *Moringa stenopetala* leaves in liver and kidney of female Wistar rats.

**Methods:** For acute toxicity study, the female Wistar rats were randomly divided into control and treatment groups that received distilled water and oral administration of single dose of 5000 mg/kg (n=5) extract, respectively and followed for 14 days. For repeated daily administration of extract, the female Wistar rats were Female Wistar rats were randomly divided into control and treatment groups (n=6). Treatment groups were given daily extracts (250, 500, and 1000 mg/kg) orally. Whereas, normal control received distilled water *ad libitum* for 15 days. On the 16th day the blood was sampled to evaluate the effect on liver and kidney using clinical chemistry analyzer.

**Results:** The acute toxicity study found no signs of toxicity; hence LD50 was greater than 5000 mg/kg. The biochemical test revealed that extracts produced a rise in liver in a dose dependent manner but no effect on kidney function indicators compared with normal control.

**Conclusion:** These findings revealed that the extracts of *Moringa stenopetala* are toxic to liver but not to kidney.

Keywords: Toxicity, *Moringa stenopetala*, Liver; Kidney

Abbreviations:

ALT: Alanine Transaminase; AST: Aspartate Transaminase; ALP: Alkaline Phosphatase; LD: Lactate Dehydrogenase; GGT: Gamma-Glutamyl Transpeptidase; BUN: Blood Urea Nitrogen; EPHE: Ethiopian Public Health Institute; EtOH: Ethanol; AQ: Aqueous; EtAc: Ethyl Acetate; OECD: Organization for Economic and Co-operation and Development; D1: Frist day; D5: Fifth day; D7: Seventh day; D10: Tenth day; D14: Fourteenth day; D15: Fifteenth day; LD50: Lethal dose-50

Introduction

Plants provided effective sources of traditional medicines against many ailments since ancient times. Peoples of all continents, especially in Africa and Asia, with its diverse culture and rich plant flora, used folklore medicine for their health needs [1]. Medicinal plants contain various pharmacologically active compounds which have useful therapeutic applications [2] and many are utilized in the development of the drug industry [3]. About thirty percent of the drugs sold worldwide contain compounds derived from plants [4].

Although there is increased acceptance and consumption of herbal remedies worldwide, care must be taken not to consume harmful plants or high doses of plant extracts that could have deleterious effects on vital body organs either in short term or long term. Concerns by medical personnel indicate that herbal medicines may be harmful to vital organs such as liver and kidneys [5]. Toxic effects due to herbal medicine may manifest in a number of organs such as kidney, liver, stomach, nervous system and blood. The liver is a vital organ for maintaining of metabolic functions and detoxification from exogenous and endogenous substances like xenobiotics, drugs and viral infections [6,7].

When the liver is exposed to such substances, its protective mechanisms are overpowered due to cellular necrosis and increase in serum levels of biochemical parameters like ALT and AST [8,9]. Determination of efficacy and safety of herbal remedies is necessary as many people use them for self-medication [10,11]. For majority of herbal products in use, very little is known about their active and/or toxic constituents. Evaluating the prolonged effects of medicinal plant extracts used in humans is useful in assessing the potential toxic effects. This increases the confidence in their safety to humans, particularly for use in the development of pharmaceuticals. The provision of safe and effective herbal therapies could, thus, become a critical tool to increase access to health care [12].

*Moringa stenopetala* (Baker f.) Cudof is one of these medicinal plants widely used for treatment of variety of diseases including hypertension. *M. stenopetala* (Baker f.) Cudof. belongs to the family Moringacae represented only by a single genus *Moringa*. The genus is...
represented by 14 species to which *M. stenopetala* (Baker f.) Cudof. belongs. It is a branched tree that grows 6 to 10 m tall, thick at base bark with white to pale gray or silvery coloration [13]. It grows abundantly in south western Ethiopia at an altitude range of 1000 to 1800 m, where the leaves are eaten as vegetable. The species is known by different vernacular names such as “Shiferaw” in Amharic, “Aleko” in Wolaytga and Gamunga and “Cabbage tree” in English [14]. The indigenous knowledge and use of *Moringa* is referenced in more than 80 countries and it’s known in over 200 local languages.

The history of *Moringa* dates back to 150 B.C. The family Moringaceae is a monotypic family of single genera with around 33 species of which 4 is accepted, 4 are synonym and 25 are not investigated. Out of these, 13 species, native of old world tropics are documented [15]. *M. stenopetala* (Baker f.) Cudof. is among various types of Moringa species which is native to Ethiopia, Northern Kenya and Eastern Somali and is the most economically important species after *Moringa oleifera* [16]. *M. stenopetala* (Baker f.) Cudof. is a multipurpose plant [13]. It has been reported that, *M. stenopetala* (Baker f.) Cudof. has hypotensive [17], antihyperglycemic and hypoglycemic [18-22], antileishmenial and antiinflammatory [14], treatment of stomach pain, anti-spasmodic and to expel retained placenta following birth [23], antimicrobial activity [24] and also has a nutritional value [13]. The *in vitro* study demonstrated the beneficial biochemical effects of *M. stenopetala* (Baker f.) Cudof. by inhibiting intestinal α-glucosidase, pancreatic cholesterol esterase and pancreatic lipase activities [22]. The objective of the present study is, therefore, to investigate the safety of extracts and fractions of *M. stenopetala* (Baker f.) Cudof. leaves in rats.

**Materials and Methods**

**Drugs and chemicals**

Ethyl Acetate (lot no: 8114/4, Park Scientific Limited, Northampton, UK), Absolute Ethanol (lot no: E35070/2, WINLAB, UK) and Petroleum Ether (lot no: V2B668282B, Scientific limited, UK) were used in the study. All the drugs, chemicals and reagents used complied with the required standard and were of analytical grade.

**Instruments and apparatus**

Lyophilizer/ Freeze dry system (Labconco, 12 L Console Freeze Dry 230v-60 (7754040), Freeze Dry System, USA), Centrifuge (Rotant 98, Hettich, Zentrifugen, UK), Clinical chemistry analyzer (Cobas-e-411, HITACHI, ROCHE, Germany).

**Plant materials**

The fresh *M. stenopetala* (Baker f.) Cudof. leaves were collected from Southern Ethiopia around Arbaminch, about 500 km far from Addis Ababa on September 2014. The plant material was authenticated by a taxonomist in the EPH and a voucher number AL-001 was deposited in the herbarium for future reference.

**Experimental animals**

The experiments were performed on adult, healthy female Wistar rats (*Rattus norvegicus*) weighing 150-200 g bred and obtained from the EPH. All the animals used for this study were kept in standard animal cages and maintained under laboratory conditions of temperature (22 ± 30°C), relative humidity (40-70%) and 12 hour day-12 hour night and had free access to food (standard pellet diet) and water *ad libitum*. The animals were treated humanely throughout the study period and were kept in a well-controlled area according to the guideline for use and care of animals [25].

**Plant material preparation and extraction**

Fresh *M. stenopetala* (Baker f.) Cudof. leaves were garbled, chopped, dried under shade (at room temperature), grinded to powder using mortar and pestle and stored in cool and dry place. Weighed amounts of 1.208 Kg and 2.130 Kg powdered leaves were kept in Erlenmeyer flasks and macerated with water (distilled and deionized) and 70% ethanol at room temperature under a rotator shaker until exhaustion for 4 and 72 hours, respectively. The 70% EtOH extract was filtered using cotton gauze and then with Whatman filter paper No.1. The filtrate was concentrated under reduced pressure using Rota vapor. The semidried residue was kept on a water bath at 400°C overnight and then with a Lyophilizer to completely remove the solvent residue. The AQ crude extract was filtered using a Whatman filter paper No.1, kept in refrigerator overnight to freeze and lyophilized to remove the water. The total yield of the AQ and 70% EtOH extract were calculated 17.1% and 4.9% (w/w), respectively. About 178.95 g of dried AQ crude extract were defatted by petroleum ether and partitioned with EtAc, the solvent was removed using Rota vapor and Lyophilizer to obtain EtAc (18.6% w/w yield) and AQ residue (35% w/w yield), respectively. The dried extracts were kept in a refrigerator until used for the experiment.

**Evaluation of effects on liver and kidney**

Female Wistar rats were randomly divided into groups with 6 animals (n=6). Normal control rats (Group 1) received distilled water *ad libitum* and treatment rats (Group 4-13) received different extracts (250, 500, 1000 mg/kg/day) for 15 days. During the experiment, the animals were observed for detection of clinical signs of toxicity.

On the 16th day, the blood was collected in vacutainer tube by cardiac puncture from night fasted cervical dislocated rats. The serum was separated after centrifugation at 3000 rpm for 10 minutes. The liver function indicators (ALT, AST, ALP and GGT) and kidney function indicators ((BUN and creatinine) were assayed using methods described by the manufacturer (Roche diagnostics, Germany) using COBA-e-411 Clinical chemistry analyzer instrument.

**Acute toxicity study**

The extracts of *M. stenopetala* (Baker f.) Cudof. leaves (AQ crude, 70% EtOH crude, EtAc fraction and AQ residue of AQ crude extracts) were evaluated for possible toxic effect in female Wistar rats at a dose of 5000 mg/kg body weight according to OECD guidelines No. 425 [26].

On the basis of oral reports of high consumption of the leaves for diet mixed with cultural foods, the chosen dose of extract was highest (5000 mg/kg body weight). The animals were deprived of food for 18 hours with free access to water. Immediately after administration of the extract, the animals were carefully observed continuously for the first 4 hours for any overt signs of toxicity and death and then for the next 24 hours. Thereafter, they were kept under close observation up to 14 days to monitor the presence of any signs of morbidity or mortality. The weight of each animal was recorded at the 1st, 7th, and 14th day of administration to verify any unexpected weight change that might have occurred. During the experiment, the animals were observed for detection of clinical signs of toxicity. Finally, after cervical dislocation,

the rats were dissected at the 14th day to observe gross pathology of the vital organs such as liver, kidney, spleen and pancreas.

Statistical analysis

Data were analyzed using the SPSS version 16 software package for ANOVA and p < 0.05 was considered as the level of significance.

Results and Discussion

This study investigated the safety of different extracts and fractions of *M. stenopetala* (Baker f.) Cudof. leaves in rats through physical and gross pathological observation as well as biochemical assessment of liver and kidney in rats.

Effect on body weight

All groups showed an increase in weight throughout the experiment. All groups did not showed a significant difference in percent weight gain on D5 and D10 except the group that took 500 mg/kg/day AQ residue of AQ extract (P < 0.05), whereas, in D15 the percent weight gain is high in groups that took 500 mg/kg/day followed by 250 mg/kg/day and 1000 mg/kg/day of extracts in similar manner in succession. All treatment groups that administered highest dose (1000 mg/kg/day) of extracts showed a significant difference (P < 0.001) in percent weight gain in D15 of the study compared with normal control (Graph 1).

All groups of rats in this study showed increase in body weights which is in agreement with the study done on sub-acute administration of AQ crude extract of *M. stenopetala* (Baker f.) Cudof. leaves in mice [27]. However, the percentage of body weight gain was reduced in the highest doses of all extracts (1000 mg/kg/day). This reduction was significant compared to normal control in the D15 of the experiment. This finding is in line with that of the previous study on sub-acute administration of AQ crude extracts of *M. oleifera* in rats [28-30].

Effect on liver enzymes and kidney metabolites

The extracts and fractions increased ALT, AST, ALP and GGT levels in a dose dependent manner compared with normal control. All treatment groups did not show significant difference in creatinine compared with normal control (Tables 1 and 2).

<table>
<thead>
<tr>
<th>Substance administered</th>
<th>Dose (mg/kg)</th>
<th>ALP (U/L)</th>
<th>ALT (U/L)</th>
<th>AST (U/L)</th>
<th>GGT (U/L)</th>
</tr>
</thead>
<tbody>
<tr>
<td>AQ crude</td>
<td>250</td>
<td>166.03 ± 10.79</td>
<td>62.88 ± 3.65</td>
<td>162.57 ± 10.85</td>
<td>2.80 ± 0.21</td>
</tr>
<tr>
<td></td>
<td>500</td>
<td>211.57 ± 10.85</td>
<td>77.87 ± 6.01</td>
<td>238.58 ± 42.96</td>
<td>3.06 ± 0.43</td>
</tr>
<tr>
<td></td>
<td>1000</td>
<td>239.33 ± 8.43</td>
<td>80.50 ± 0.99</td>
<td>246.78 ± 14.08</td>
<td>3.58 ± 0.15</td>
</tr>
<tr>
<td>70% EIOH crude</td>
<td>250</td>
<td>179.18 ± 7.92</td>
<td>63.75 ± 3.55</td>
<td>176.15 ± 12.81</td>
<td>2.55 ± 0.18</td>
</tr>
<tr>
<td></td>
<td>500</td>
<td>219.57 ± 15.24</td>
<td>64.10 ± 2.39</td>
<td>193.73 ± 27.64</td>
<td>2.71 ± 0.35</td>
</tr>
<tr>
<td></td>
<td>1000</td>
<td>227.08 ± 15.62</td>
<td>66.67 ± 1.54</td>
<td>214.15 ± 9.84</td>
<td>3.31 ± 0.18</td>
</tr>
<tr>
<td>EtAc fraction of AQ crude</td>
<td>250</td>
<td>195.47 ± 11.59</td>
<td>65.03 ± 3.25</td>
<td>199.47 ± 35.39</td>
<td>2.850 ± 0.56</td>
</tr>
<tr>
<td></td>
<td>500</td>
<td>204.93 ± 25.91</td>
<td>65.93 ± 2.38</td>
<td>244.32 ± 20.88</td>
<td>3.5500 ± 0.49</td>
</tr>
<tr>
<td></td>
<td>1000</td>
<td>222.02 ± 11.19</td>
<td>68.13 ± 1.17</td>
<td>251.33 ± 7.37</td>
<td>4.3000 ± 0.61</td>
</tr>
<tr>
<td>AQ residue of AQ crude</td>
<td>250</td>
<td>193.23 ± 13.29</td>
<td>57.18 ± 2.02</td>
<td>186.57 ± 19.75</td>
<td>3.1333 ± 0.17</td>
</tr>
<tr>
<td></td>
<td>500</td>
<td>215.67 ± 4.11</td>
<td>64.20 ± 0.91</td>
<td>196.73 ± 5.55</td>
<td>3.5333 ± 0.39</td>
</tr>
<tr>
<td></td>
<td>1000</td>
<td>237.53 ± 18.38</td>
<td>65.23 ± 3.02</td>
<td>200.27 ± 25.00</td>
<td>3.7500 ± 0.29</td>
</tr>
<tr>
<td>Distill. water</td>
<td><em>ad libitum</em></td>
<td>174.47 ± 8.64</td>
<td>66.63 ± 3.54</td>
<td>194.75 ± 18.18</td>
<td>3.9667 ± 1.16</td>
</tr>
</tbody>
</table>

Table 1: Effect of crude extracts and fractions of *M. stenopetala* on liver of rats (Results are expressed as mean ± SEM (n = 6), * = Compared to normal control; 1 = P < 0.05; 2 = P < 0.01; 3 = P < 0.001)

Effect on liver enzymes

The liver is vulnerable to various environmental toxicants which may cause structural and functional abnormalities. If there is mild inflammation and tissue damage to these organs, the permeability of the cell membrane will increase and release cytoplasmic enzymes such as LD, ALP, and AST, while necrosis will release mitochondrial ALT as well as AST leaking into the blood and increase in levels. Testing of AST and ALT are used for hepatocellular (damage to liver cells and liver disease) evaluation while ALP is used for hepatobiliary (cholestasis or hyperbilirubinemia) evaluation which is used to determine potential target organs toxicity and associated time courses of the damage without the need of biopsy or necropsy samples [31-33].

From the present work all treatment groups elicited an increase in AST, ALT, ALP and GGT in a dose dependent manner though not significant. The largest serum ALP, AST, ALP and GGT was observed for groups that took the maximum dose (1000 mg/kg/day) of AQ crude extract. The results are in line with those of the studies done on extract of *M. stenopetala* (Baker f.) Cudof. leaves in HEPG2 cells *in vitro* [34], sub-chronic administration of 70% EIOH crude extract of *M. stenopetala* (Baker f.) Cudof. leaves in mice [20], sub-acute
administration of AQ crude extract of *M. stenopetala* (Baker f.) Cudof. leaves in mice [27] and seven days administration of EtOH extract of *M. oleifera* leaves in rats [35].

The liver and kidney are major organs of early screening efforts in the preclinical research and a major target organ in the repeated-dose nonclinical safety studies used to support clinical trials [29-30].

**Effect on kidney metabolites**

Toxic agents may affect kidneys and impair their physiological functions. The effect on kidney can be investigated by cross checking the normally expected function, such as in excreting waste product like BUN and creatinine [36].

In this study renal toxicity was assessed by measuring BUN and creatinine and the results showed that there was no significant increase in the plasma level of BUN and creatinine in comparison with normal control which indicated nontoxic effect of the extract on the kidney. This finding is in agreement with those of the studies done on sub-chronic administration of 70% EtOH crude extract of *M. stenopetala* (Baker f.) Cudof. leaves in mice [20], sub-acute administration of AQ crude extract of *M. stenopetala* (Baker f.) Cudof. leaves in mice [27] and sub-acute administration of AQ crude extract of *M. oleifera* leaves in rats [28].

<table>
<thead>
<tr>
<th>Substance administered</th>
<th>Dose (mg/kg)</th>
<th>Kidney parameters</th>
<th>Creatinine (mg/dl)</th>
<th>BUN (mg/dl)</th>
</tr>
</thead>
<tbody>
<tr>
<td>AQ crude</td>
<td>250</td>
<td></td>
<td>0.39 ± 0.02*2</td>
<td>19.60 ± 1.01*3</td>
</tr>
<tr>
<td></td>
<td>500</td>
<td></td>
<td>0.34 ± 0.01</td>
<td>22.03 ± 1.44*3</td>
</tr>
<tr>
<td></td>
<td>1000</td>
<td></td>
<td>0.32 ± 0.00</td>
<td>23.33 ± 0.42*2</td>
</tr>
<tr>
<td>70% EtOH crude</td>
<td>250</td>
<td></td>
<td>0.30 ± 0.01</td>
<td>24.00 ± 0.81*1</td>
</tr>
<tr>
<td></td>
<td>500</td>
<td></td>
<td>0.32 ± 0.01</td>
<td>22.55 ± 1.76*2</td>
</tr>
<tr>
<td></td>
<td>1000</td>
<td></td>
<td>0.34 ± 0.02</td>
<td>21.15 ± 0.73*3</td>
</tr>
<tr>
<td>EtAc fraction of AQ crude</td>
<td>250</td>
<td></td>
<td>0.39 ± 0.01</td>
<td>21.81 ± 1.39*3</td>
</tr>
<tr>
<td></td>
<td>500</td>
<td></td>
<td>0.35 ± 0.01</td>
<td>18.56 ± 0.73*3</td>
</tr>
<tr>
<td></td>
<td>1000</td>
<td></td>
<td>0.32 ± 0.00</td>
<td>24.48 ± 0.44*1</td>
</tr>
<tr>
<td>AQ residue of AQ crude</td>
<td>250</td>
<td></td>
<td>0.40 ± 0.01</td>
<td>17.83 ± 0.89*3</td>
</tr>
<tr>
<td></td>
<td>500</td>
<td></td>
<td>0.30 ± 0.01</td>
<td>18.06 ± 1.21*3</td>
</tr>
<tr>
<td></td>
<td>1000</td>
<td></td>
<td>0.34 ± 0.01</td>
<td>22.33 ± 0.80*3</td>
</tr>
<tr>
<td>Distill. water</td>
<td>ad libitum</td>
<td></td>
<td>0.33 ± 0.00</td>
<td>29.80 ± 0.92</td>
</tr>
</tbody>
</table>

**Table 2:** Effect of crude extracts and fractions of *M. stenopetala* (Baker f.) Cudof. on kidney of rats (Results are expressed as mean ± SEM (n = 6), *= Compared to normal control; 1 = P < 0.05; 2 = P < 0.01; 3 = P < 0.001)

**Acute oral toxicity**

The acute toxicity study indicated that the extract did not show any signs of toxicity with close follow up for 4 hours and cause no mortality within 24 hours and 14 days at a dose of 5000 mg/kg body weight. After 14 days of observation of the experimental rats, no body weight reduction as well as over increase was observed. Gross physical and behavioral observation also revealed no visible signs of toxicity. Additionally, there was no gross pathological alteration (color, size and texture) of the vital organs. Therefore, the LD50 of these extracts was greater than 5000 mg/kg (Graph 2).

**Graph 1:** Effect of different extract and solvent fractions of *M. stenopetala* (Baker f.) Cudof. on body weight in rats

**Graph 2:** Effect of single dose (5000 mg/kg) of different extracts and solvent fractions of *M. stenopetala* (Baker f.) Cudof. leaves in body weight of rats

Aside from being highly revered for its medicinal value, *M. stenopetala* (Baker f.) Cudof. has also served the community as a source of food for centuries. With this prior information of safety, limit test was found to be the appropriate test for assessing the acute toxicity profile with high dose (5000 mg/kg body weight) as per the OECD guideline [26].
The acute toxicity study of oral administration of the limit dose of 5000 mg/kg of the AQ crude extract and its solvent fractions (EtAc fraction and AQ residue) and 70% EtOH crude extract of M. stenopetala (Baker f.) Cudof. leaves indicated that there was no mortality observed in all test animals during the course of the study period. Furthermore, no overt behavioral and physical signs of toxicity were discerned at this dose. The observation that there was no sign of morbidity as well as mortality at this high dose, it would in fact allow making suggestions that the oral LD50 of the extract could be greater than 5000 mg/kg.

All experimental groups showed an increase in body weight and a reasonable percent weight gain in D7 and D14 of the experiment compared with normal control. This finding is in line with the study done on 70% EtOH extract of M. stenopetala (Baker f.) Cudof. leaves at 5000 mg/kg in mice [18], n-butanol fraction of 70% EtOH extract M. stenopetala (Baker f.) Cudof. leaves at 5000 mg/kg in mice [19] and AQ extract of M. oleifera leaves at 2000 mg/kg in rats [28]. Therefore, the outcome of the acute toxicity test could support the plant for folkloric medicinal use by the community.

Conclusion

Findings in the acute toxicity test suggest that the aqueous leaf extract of M. stenopetala are practically non-toxic to the female Wistar rats when administered orally. Whereas, the repeated oral daily administration has revealed potential damage to liver in a dose dependent manner but not to the kidney. Further studies, however, need to be done to confirm this.

Acknowledgement

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