

The Impacts of Microplastics to Environment

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In today's world, plastic litter has been positioned in the expanded list of worldwide threats, counting climate change and ozone depletion [1]. Plastic debris is regularly separated into two categories: macro plastics and micro plastics. Macro plastic is a well-known worldwide issue potentially causing negative impacts on both the life forms and environment, counting trap, ingestion, retention of harmful chemicals, and transportation of obtrusive species. Macro plastics are known threat to human society and the economy [2,3], while micro plastics are regularly received less consideration. In recent years, it has taken note that impacts caused by microplastics are comparatively more critical than those caused by macro plastics, so there have been increasing environmental concern about tiny plastics.

Microplastics are defined to be the plastic particles between 1 nm to <5 mm in diameter. They are originated from fragmented macro plastics by mechanical abrasion and UV exposure [4], man-made fibers for textiles releasing from domestic washing [5], microplastics used in consumer and cosmetic products, e.g. facial cleaners, toothpaste and etc [6]. These debris resulting from the disposal and breakdown of consumer products, and industrial waste are found in oceans, estuaries, bodies of freshwater and even in the tap water is now well established [7-13]. Due to its light weight, plastic litter transported by winds and currents, and recirculates between seawater and beach sediments. Polymer density is an important determinant for microplastics circulation [14]. Microplastics would cause entanglement and ingestion by a range of marine organisms such as zooplankton, fish, seabirds, sea turtles, crustaceans and mammals has been documented [15]. Smaller microplastics were found to cause higher toxicity to algae [16]. The adsorption of toxic polycyclic aromatic hydrocarbon [17], heavy metals [18] and pathogens [19] on microplastics promoted the negative impacts to marine organisms, probably via the increase in oxidative stress [20] and reduction of nutrient uptake [21]. Microplastics introduce harmful impacts at the tissue and cellular level, and meddled with energy reallocation, reproductive success, and sibling execution [22,23], which pose a threat to biodiversity and environments [2]. Although identified as an emerging environmental threat to the freshwater ecosystems and its ecological consequence. Wastewater treatment plant effluents represent an important point source for micro plastic particles for freshwater environments [24,25].

Presently, there are no standardized protocols for surveying, measuring and monitoring micro plastics in natural ecosystems [26,27]. A standardized method for microplastics measurement is required for data comparability, which should be in low cost and with capability for high volume throughput with acceptable accuracy. Current common approaches to quantify microplastics from sediment would involve multiple steps, including drying to reduce volume, followed by separation, and confirmed by analytic equipment.

Separation could be carried out by density separation, filtration, and visual sorting [28]. Visual sorting is time consuming and introduces a lot of false identification. As microplastics are light, floatation techniques using super saturated NaCl [29], sodium nitrate/sodium thiosulfate (SNT) solution [30], Zinc chloride solution [25] were commonly used. Filtration using discfilter with rapid sand filtration and air floatation was introduced in the final stage of municipal wastewater treatment [31]. Additional pre-separation digestion with enzyme [32], acids or chemicals [33] to remove attached organic material without damaging the microplastics would improve the extraction and analysis. The isolated microplastics would then be analyzed by advanced analytical equipment, such as micro-Fourier-transform infrared (micro-FT-IR) spectroscopy, focal plane array (FPA)-based transmission micro-FT-IR imaging [25], raman spectroscopy [34], pyrolysis gas chromatography/mass spectrometry [35], thermal desorption gas chromatography mass spectrometry [36], field-portable-X-ray fluorescence (FP-XRF) spectrometry [18], Nuclear Magnetic Resonance (NMR) [37] and TGA-DSC [38]. They have been employed to detect microplastics in marine habitats by identifying the molecular construction of different plastic types from other materials, which would reduce 22 to 90% of false identification [39]. To use alkaline and wet peroxide oxidation chemical digestion techniques to remove microplastics and followed by looking at the loss of signal in analysis provided an alternative approach to quantify Microplastics [40]. Apart from using advanced analytical equipment, simple method using florescence dye e.g. Nile red was reported to have comparable result of FTIR with 98% recovery rate [41].

More than 300 million tons of plastic are made each year worldwide. This includes polyethylene terephthalate (PET), Polyethylene (PE), Polyvinyl Chloride (PVC), Polystyrene (PS). Equally it is hardly biodegradable, although their degradation can be speed up by UV exposure [42]. However, the process of photo aging is slow, approximately release 3% of content after 2000 hour of photo aging [43]. PE and PS are relatively easier to be degraded in natural environment [44]. However, they can be found even decades later as plastic litter and micro plastic, especially PET is mainly found in all forms of plastic bottles and promotional material. Approximately 51 million tons of PET was produced worldwide in 2014 [45].

According to Microbial degradation of plastics has been provided by [46-48]. There is very little earlier study on the biological degradation of plastic litter or its utilization to support microbial growth. This brings out a whole range of both terrestrial and marine microbial species capable of the degradation activity. Several species of bacteria and fungi have been set apart, showing degradation properties of different cases of plastic polymers. Rare examples include members of the filamentous fungi *Fusarium oxysporum* and *Fusarium solani*, which have been shown to grow on a mineral medium containing PET yarns [49]. Recent reports have found marine fungus *Zalerion maritimum* is capable to utilize PE [37], while bacterial isolates of

Bacillus cereus and *Bacillus gottheilii* degrade UV-treated microplastics [50]. Consequently, plastics being resistant to degradation need certain pre-treatment like photo-oxidation or hydrolysis or enzymatic degradation by microorganisms, before the polymers can be metabolized by the beings. Thus, microbes are known to initiate the process of degradation of marine plastic through the organization of a biofilm and secretion of extracellular enzymes to aid in breaking down of plastic polymers (Table 1).

In conclusion, a huge body of knowledge exists for degradation capabilities by microbes; there is even a lack of technological and real time applications of these biological processes in the surroundings.

Plastic bioremediation studies suffer from a major limitation, the recalcitrant nature of plastic polymers, which need extra discussion. This treatment could be either chemical or physical methods that could develop down the polymer chains and help speed up the biological processes. Such treatments can generally be enzyme responsible for this degradation may lead to cost-effective and environmentally conscious method for degrading micro plastic. A framework of standard for "Ecocyclable in natural carbon cycle" related to toxicity, bioaccumulation and degradation/assimilation is highly suggested [51-53].

Type of Plastic	Species	Reference
Polyethylene terephthalate	Ideonella sakaiensis	Yoshida et al., [54]
Polycaprolactone	Pseudozyma jejuensis	Seo et al., [55]
Polyethylene	Pseudomonas sp	Sudhakar et al., [56]
Polyethylene	Enterobacter asburiae & Bacillus sp. Zalerion maritimum	Jun Yang et al., [57] Paço et al., [36]
Bisphenol A (BPA)	Pseudomonas sp	Artham & Doble et al., [58]
Low Density Polyethylene	Aspergillus versicolor	Pramila & Ramesh [59]
Low Density Polyethylene	Chamaeleomyces viridis	Anudurga Gajendiran et al., [60]
polyester	Geomyces pannorum	Cosgrove et al., [61]
Polyester polyurethane	Pestalotiopsis microspore	Russell et al., [62]
Polyethylene	Bacillus cereus	Sudhakar et al., [63]
Polyethylene	Brevibacillus borstelensis	Hadad et al., [64]
Polystyrene	Rhodococcus ruber	Mor and Sivan [65]

Table 1: Biological degradation of plastics.

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