Pub: Probing the Interaction of Human Salivary Alpha-Amylase and Amylase Binding Protein A (AbpA) of Streptococcus gordonii

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Abstract
Amylase binding protein A (AbpA) of Streptococcus gordonii serves as a major receptor for human salivary α-amylase (HSAmy), the predominant enzyme in human salivary secretions, to bind to the bacterial cell surface. On enamel surfaces, the binding of AbpA to HSAmy renders S. gordonii act as a scaffold for other oral bacteria to attach to the acquired enamel pellicle leading to complex bacterial communities and invasion of host tissues. While the role of AbpA in adhesion, starch metabolism and biofilm formation in influencing the ecology of the oral biofilms has been established, the structure-function relationships of AbpA are yet to be defined. Since distally located aromatic residues of HSAm are involved in the interaction with AbpA, we hypothesized that AbpA might use separate structural regions to bind to HSAm. To test this, several deletion mutants of AbpA were constructed and studied to correlate the effect of deletions in binding to HSAm in the following: 1) their secondary structural features through circular dichroism studies; 2) their ability to alter the capacity of HSAmy to hydrolyze starch and several oligosaccharides after complex formation and 3) their binding to HSAm using surface plasmon resonance spectroscopy. Our results showed that a) AbpA does not bind at the active site of salivary α-amylase; b) both the N-terminal region (24-56 residues) and a central region (residues 124-165) are required for binding to HSAmy; and c) the C-terminal end residues (166-195) are not necessary for binding. These results clearly show that AbpA binding to HSAmy encompasses distinct and distal regions of the structure.

Keywords: α-amylase; Mutagenesis; Hydrolysis; Bacterial binding; Amylase-binding

Abbreviations: HSAmy: Human Salivary α-amylase; AbpA: Amylase Binding Protein A; SPR: Surface Plasmon Resonance; RU: Resonance Unit

Introduction
Human salivary α-amylase (HSAmy), a multifunctional enzyme present in the oral cavity of humans, plays a significant role in saliva-bacteria interactions and participates in the establishment of dental biofilm [1]. HSAmy is consistently present in the enamel pellicle [2] where it acts as a receptor to a group of streptococci known as the amylase-binding streptococci (ABS), which include S. gordonii, S. mitis, S. parasanguis, S. cristatus and S. salivarius [3-6]. Among these bacteria, S. gordonii, a pioneer colonizer present in substantial quantity in the early enamel pellicle partly due to its ability to attach to HSAm in the pellicle. This initial attachment to HSAm might be a critical step in the development of dental biofilms since it provides a scaffold for non-amylase binding streptococci. Nutritional support for the dental biofilm bacteria is provided by HSAmy through its ability to hydrolyze α-1,4-glucosidic bonds in starch and other polysaccharides during food intake. Thus, each of the three functions of the enzyme, namely, hydrolysis of starch, adherence to enamel and binding to viridans streptococci together might play a role affecting the formation of oral biofilms.

The ability of S. gordonii to bind to HSAmy has been attributed to the presence of low- molecular weight protein (AbpA) and high molecular weight protein (AbpB) [7-9]. Recent studies have shown that several streptococci express a number of proteins of different sizes ranging from 20 to 87 kDa with a common property of binding to HSAm. Amongst these amylase-binding proteins, the low molecular weight protein AbpA has been extensively studied to establish its role in S. gordonii biofilm formation [10-12]. Several characteristics of AbpA have led to a detailed study of this protein including its small size, its role in biofilm formation of S. gordonii, its ability to bind HSAm to the bacterial surface and thus enhance starch-induced growth [11]. Its ability to increase the enzyme activities with Gifs of both S. gordonii and S. mutans in the presence of HSAm have led to suggest that HSAm: AbpA complex may modulate the bacterial adhesion and colonization in ways that is not completely understood [1,12]. The second protein AbpB, due to its enzymatic property of being a dipeptidase, although important for bacterial colonization, is somewhat limited in the binding of HSAm to the bacterial surface [1]. Interestingly, mutation of abpB did not abolish the binding of HSAm whereas mutation of abpA led to the absence of HSAm binding to S. gordonii [13]. Unlike AbpA, the role of AbpB has been in bacterial colonization suggested to be through nutrient acquisition pathways [1].

While significant progress has been made in the identification of various amylase-binding proteins of oral streptococci and study of their functions, especially AbpA, little is known about the structural basis for these proteins to interact with HSAmy. In this regard, several studies have been reported on the structural aspects of HSAm and the role of several residues in the active site as well as in the secondary saccharide binding sites [14-20]. Interestingly, our earlier study suggested that the AbpA interacts with amylase over an extended surface region with two distinct sites both of which have aromatic residues [20]. The binding occurs in such a way that the binding of smaller oligosaccharides to the active site in HSAmy is not blocked but hinders the binding of starch suggesting that secondary saccharide binding sites of HSAmy (site involving W203 and W284) in the vicinity of the active site were being used by AbpA [20]. Nevertheless, for a complete understanding of the binding interaction between HSAm and AbpA, structure-function...
studies on AbpA are warranted. In this regard, it was necessary to understand the structural features of AbpA that could provide such an extended interaction with α-amylase. Hence, in this study, we have focused on the structure function relationships of AbpA and its interaction with HSAmy through molecular modeling, selective deletion mutagenesis and biophysical studies.

**Materials and Methods**

**General procedures**

All the chemicals used herein were obtained from Sigma Chemical Co. Oligonucleotides used in this study were obtained from Integrated DNA Technologies. The core facilities at the University of Medicine Co. Oligonucleotides used in this study were obtained from Integrated DNA Technologies. The core facilities at the University of Medicine

**Bacterial strains, plasmids and growth conditions**

*S. gordonii* G9B was used for cloning of *abpA* gene, *E. coli* DH5α (NEB10-β), *E. coli* Rosetta™ (DE3) and vector pET29b were used for standard cloning and protein expression. Streptococci were maintained on tryptic soy broth supplemented with 0.5% yeast extract (TSBY) and were incubated at 37°C for 12 to 16 hours. *E. coli* strains were grown under aerobic conditions with shaking for 12 to 16 h at 37°C in Luria-Bertani (LB) broth and maintained on LB agar. *E. coli* strains containing recombinant clones were plated on LB agar and supplemented as required with kanamycin (30 µg/mL). The *abpA* gene was PCR amplified from the *S. gordonii* G9B genomic DNA using a forward primer that introduced an NdeI restriction site and a reverse primer containing a KpnI restriction site and a stop codon. The PCR product was digested with enzymes NdeI and KpnI, ligated into the pET29b vector (Novagen) between its NdeI and KpnI restriction sites to yield the plasmid pCR4. The resulting plasmid encodes the full-length mature protein (AbpA<sub>1-195</sub>) with a C-terminal His<sub>6</sub> tag. Selective deletion of N and C terminal domain was done to generate four variants of AbpA. The full length AbpA without the signal sequence was named PG1 (24-195), N terminal deletion variant was named PG2 (54-195), C terminal deletion was named PG3 (24-165), a truncated mutant with the 1-100 residues of the mature protein was named PG4 (24-124) and a fifth mutant with residues encompassing the C-terminal region (residues 124-195) was named PG5. The primers used for construction of the full length AbpA and the variants are listed in Table 2.

**Expression and purification of AbpA**

The protein AbpA was expressed using *E. coli* Rosetta™ (DE3) cells transformed with pCR4 using procedures described before [20]. Typically, 1 L of Luria-Bertani broth supplemented with 30 µg/mL of kanamycin per mL was inoculated with 10 mL of an overnight culture of *E. coli* strain Rosetta™ (DE3) (Novagen) [21] transformed with the plasmid pCR4. After incubation at 37°C for 4 h or until the optical density of the culture (OD 600) reached 0.6, protein expression was induced by the addition of isopropyl-β-D-1-thiogalactopyranoside to a final concentration of 0.1 mM. The cells were incubated post-induction for 5 h at 37°C and then harvested by centrifugation at × 6000 g. The cell pellet was resuspended in 20 mL of lysis buffer (20 mM Tris-Cl, pH 8.0, 500 mM NaCl, 1 mM phenylmethylsulfonyl fluoride and 0.1% Nonidet P-40). The cell suspension was then sonicated on ice for 30 s (x5 with 2 min intervals) at 30% capacity with a 30% duty cycle by using a Branson model 450 sonicator equipped with a microprobe. The cell debris was pelleted by centrifugation (15,000 g for 20 min; 4°C), and the supernatant was filtered using 0.45 µm syringe filter (Corning) and passed through a 0.2 µm filter (Millipore). The supernatant was applied to a column of a 5 mL Ni<sup>2+</sup>-NTA agarose (Qiagen) equilibrated with 20 mM Tris-Cl, pH 8.0, 500 mM NaCl, 1 mM phenylmethylsulfonyl fluoride and 0.1% Nonidet P-40. Upon 100% binding of the His<sub>6</sub>-tagged AbpA, the column was washed with wash buffer (20 mM Tris-Cl, pH 8.0, 500 mM NaCl, 1 mM phenylmethylsulfonyl fluoride and 0.1% Nonidet P-40) and then the eluted protein was dialyzed against 50 mM sodium phosphate, pH 7.4.

**Table 1:** Kinetic parameters for the binding of AbpA to HS Amy enzymes derived from surface plasmon resonance studies.

<table>
<thead>
<tr>
<th>Analyte</th>
<th>k&lt;sub&gt;s&lt;/sub&gt; (1/Ms)</th>
<th>k&lt;sub&gt;d&lt;/sub&gt; (1/s)</th>
<th>K&lt;sub&gt;a&lt;/sub&gt; (1/M)</th>
<th>K&lt;sub&gt;d&lt;/sub&gt; (M)</th>
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<tr>
<td>PG1</td>
<td>5.7 × 10&lt;sup&gt;4&lt;/sup&gt;</td>
<td>8.0 × 10&lt;sup&gt;4&lt;/sup&gt;</td>
<td>7.1 × 10&lt;sup&gt;4&lt;/sup&gt;</td>
<td>1.4 × 10&lt;sup&gt;10&lt;/sup&gt;</td>
</tr>
<tr>
<td>PG2</td>
<td>1.6 × 10&lt;sup&gt;4&lt;/sup&gt;</td>
<td>2.0 × 10&lt;sup&gt;2&lt;/sup&gt;</td>
<td>8.2 × 10&lt;sup&gt;4&lt;/sup&gt;</td>
<td>1.2 × 10&lt;sup&gt;8&lt;/sup&gt;</td>
</tr>
<tr>
<td>PG3</td>
<td>2.1 × 10&lt;sup&gt;4&lt;/sup&gt;</td>
<td>1.4 × 10&lt;sup&gt;4&lt;/sup&gt;</td>
<td>1.4 × 10&lt;sup&gt;4&lt;/sup&gt;</td>
<td>6.9 × 10&lt;sup&gt;10&lt;/sup&gt;</td>
</tr>
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**Table 2:** Primers used in this study to generate AbpA variants. The underlined sequences correspond to the restriction sites (Ndel in forward and KpnI in reverse). All constructs have a His<sub>6</sub> tag sequence at the 3′ end.
variants were incubated for 30 minutes before G7-PNP was added. This 100 µl) consisted of HSAmy (60 nM), AbpA variants (480 nM), Boehringer Mannheim). Each reaction mixture (total volume of

used in the dinitrosalicylic acid assay [Bernfeld, 1955]. Similarly, to test phosphate buffer (pH 6.9) containing 6 mM NaCl. Prior to use, the baseline, and measured in the same cells. These results were normalized from 260 nm to approximately 200 nm (with a step size of 0.5 nm

a protein assay kit (Bio-Rad) and nanodrop. Spectra were collected and 0.01 M sodium phosphate (pH 7.4) was transferred into quartz cells with 0.1 cm path length. Concentrations were determined using a protein assay kit (Bio-Rad) and nanodrop. Spectra were collected from 260 nm to approximately 200 nm (with a step size of 0.5 nm and a bandwidth of 1.5 nm) at room temperature, corrected for buffer baseline, and measured in the same cells. These results were normalized to mean residue ellipticities ([θ]).

Circular dichroism

CD spectra were recorded on an AVIV model 400 spectrophotometer equipped with a five-position thermostated cell holder controlled by a Hewlett-Packard Peltier temperature controller µg/mL in 0.15 M NaCl and 0.01 M sodium phosphate (pH 7.4) was transferred into quartz cells with 0.1 cm path length. Concentrations were determined using a protein assay kit (Bio-Rad) and nanodrop. Spectra were collected from 260 nm to approximately 200 nm (with a step size of 0.5 nm and a bandwidth of 1.5 nm) at room temperature, corrected for buffer baseline, and measured in the same cells. These results were normalized to mean residue ellipticities ([θ]).

Enzyme activity and other in vitro assays

The AbpA and its variants with their truncations might be expected to exhibit differences in their ability to bind to HSamy. Previous studies [20] have shown that residual HSamy activity could be determined from the supernatants of mixtures in which HSamy was incubated with AbpA on ice for 30 minutes. Typical experiments were carried out using 1% soluble starch as substrate at 25°C for 5 min in 20 mM phosphate buffer (pH 6.9) containing 6 mM NaCl. Prior to use, the HSamy solution (150 nM) was incubated with various concentrations of the AbpA variants (0.1-6.6 µM) for 30 minutes on ice to allow for complex formation after which an aliquot of the reaction mixture was used in the dinitrosalicylic acid assay [Bernfeld, 1955]. Similarly, to test for the residual activity in the presence of smaller oligosaccharides, enzymatic reactions were carried out using maltotetraose (G7; Boehringer Mannheim). Each reaction mixture (total volume of 100 µl) consisted of HSamy (60 nM), AbpA variants (480 nM), maltotetraose (5 nM). As with the starch assay, HSamy and AbpA variants were incubated for 30 minutes before G7-PNP was added. This reaction mixture was injected onto the HPLC column 5 min after the oligosaccharide was added and the product distribution of amylase and amylase bound to AbpA variants were determined by HPLC analyses using a chromatography system (Varian) as described before [20]. The amount of residual G7 was determined using the area under each peak and converting these into concentrations using the standard curves. The product profile was analyzed based on retention times of standards run under similar conditions without the addition of the enzyme. The concentrations of the HSamy and the AbpA variants were determined by a protein assay kit (Bio-Rad) using bovine serum albumin (BSA; Sigma Chemical Co.) as the standard. All experiments were performed in triplicate and repeated twice.

BIAcore assays

Surface plasmon resonance experiments were performed on a BIAcore X 2000 instrument equipped with research-grade CM5 sensor chips as detailed before [20] with some minor modifications. The amylase binding protein constructs PG1 through PG4 were immobilized on the chip using the amine coupling kit from BIAcore. Briefly, the sensor chips were activated with a solution containing 0.1 M N-hydroxysuccinimide and 0.4 M 1-ethyl-3-(3-dimethylaminopropyl) carbodiimide (1:1). Typically, AbpA was immobilized to a maximum value 1000 RU wherein the change in refractive index induced by the association of compounds interacting with the sensor surface is expressed in RU. Unreacted residual N-hydroxysuccinimide esters were inactivated using 1 M ethanolamine. Real-time binding analyses to AbpA variants were performed at 25°C using HSamy in a buffer containing 10 mM HEPES (pH 6.9), 150 mM NaCl and 2.5 mM CaCl2 at varying concentrations ranging from 0.125 to 1 µM at a flow rate of 20 µL/min at 25°C. After each experiment, the chip surface was regenerated using 10 mM HCl. The binding parameters (kon, koff, Ka, Kd) for the interaction were calculated using BIAevaluation 3.2 software. The curves were fitted according to a simple 1:1 interaction. The fitting residuals did not exhibit any significant systematic deviations and were minimal (-1 to 1).

Results

Model of AbpA and design of the AbpA mutants

The mature AbpA is 172 residues long (24-195 numbering) excluding the signal sequence. Since the crystal structure of AbpA is not yet available and has been difficult to achieve, we resorted to the molecular modeling analysis. The homology model of AbpA obtained through the HHpred server revealed a structure with a large content of α-helical segments (Figure 1). It should be emphasized that the model presented here may not be perfectly accurate since it is based on bioinformatics approach. However, our approach has some merits to further the structural knowledge of AbpA in the absence of a definitive structure of AbpA through either x-ray crystallographic techniques or NMR. The significant feature in this model is the preponderance of the helical structure adopted by the molecule especially at the N-terminal end of the molecule. The C-terminal end of the molecule (residues 165-195) is comprised of both helix as well as β-strands. The model obtained, with a caveat that it could be somewhat less reliable due to paucity of significant homologous structures, could be easily tested for the preponderance of helical regions using circular dichroism, which is described below. Using this tentative model, the truncated mutants were designed with the following rationale. The mutant PG2, which lacks the first 30 residues (residues 24-53) were designed to test the role of the unordered N-terminal segment and the first small helical segment shown in red (Figure 1). The mutant PG3 is lacking the last 30 residues (residues 165-195), which also has a helical segment (blue


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99% purity, with a cell culture yield of 10–15 mg/L. (data not shown). AbpA mutant proteins were obtained at greater than also showed corresponding reduction in sizes in the SDS-PAGE gels calculated mass of the protein. Similarly, the other truncated variants Blue showed a single band of approximately 20 kDa consistent with the SDS-PAGE gels of the resulting purified protein stained with Coomassie obtained from the Oralgen database (http://www. stdgen. lanl. gov/) nm. The increase in helicity in these two variants could have arisen by the homology modeling since both PG1 and PG2 have very similar secondary structure content (residues 24-123), which represents a large fraction of the α-h helical protein. The CD spectrum of PG5, however, resembles the characteristics of mature AbpA (PG1) with ordered secondary structures reminiscent of the helical structures. Notably, all these variants except PG5, has the characteristic double minima of a typical α-h helical protein. The CD spectrum of PG5, however, resembles a typical unordered structure, likely due to the large deletion in the sequence (residues 24-123), which represents a large fraction of the helical structures. These CD spectra partly validate the model obtained by the homology modeling since both PG1 and PG2 have very similar spectra whereas PG3 and PG4 have a larger negative minimum at 222 nm. The increase in helicity in these two variants could have arisen from the removal of either unordered regions (PG3) or a higher ratio of helix vs. other structures in PG4. The estimation of the secondary structure content from the spectra of these variants was analyzed using the program suite CDPRO. The values estimated by the individual programs were comparable and suggest that these variants are mostly helical structures.

**Starch hydrolysis studies**

Previous studies have shown that AbpA forms a tight complex with HS Amy and the formation of the complex reduces the HS Amy’s ability to hydrolyze starch [20]. To determine whether or not the binding of AbpA variants also results in the reduction of starch hydrolyzing activity of HS Amy, the four variants along with the full length AbpA were tested. After binding, the starch hydrolysis by unbound Amylase was studied. The results are shown in figure 3. As has been observed previously, the full length AbpA, PG1 showed the highest binding as the starch hydrolysis is reduced by increasing concentration of AbpA. Interestingly, partial deletion of the C-terminal domain, as in PG3, did not have any effect in the starch hydrolysis. The residual activity of PG3: HS Amy complex mirrored that of the PG1: HS Amy complex suggesting that the last 30 residues may or may not bind to HS Amy in a way that restricts starch binding. In cases of PG2, PG4 and PG5, the reduction in the starch hydrolysis with increasing concentration of the protein is not as drastic as in cases of PG1 and PG3. In the case of PG2, a small reduction compared to PG1 implies that the first 30 N terminal residues of AbpA may bind to HS Amy in one or more of secondary saccharide binding sites of HS Amy. Moreover, starch binding is not affected in the presence of PG4 and PG5 since there is very little effect in the reduction in the residual activity. The starch hydrolysis data from PG1 and PG4 variants suggest that some of the residues in the C-terminal domain bind to HS Amy at sites where starch also binds. The absence of these residues did not affect starch binding and hence the hydrolytic efficiency of HS Amy is restored. Similarly, while the C-terminal residues might be binding to starch binding sites on the surface of HS Amy, the presence of only this domain (PG5) is ineffective to exhibit a reduction in starch hydrolytic activity. This is highly likely due to absence of any ordered secondary structures in PG5.

**Hydrolysis of oligosaccharides**

Unlike starch, which might bind to HS Amy at multiple secondary...
saccharide binding sites, smaller oligosaccharides such as G5, G6 and G7 bind to the active site of HSAmy completely [16]. Previous studies have shown that HSAmy possesses seven glucose-binding subsites [16], which completely fill the active site. In order to test whether or not the AbpA binds to one or more of the subsites of the active site, we tested the hydrolytic ability of HSAmy against maltoheptaose in the presence of AbpA and its variants. The standard hydrolysis curve of maltoheptaose was compared to the hydrolysis of maltoheptaose with the reaction mixture of AbpA and HSAmy (Figure 4). The hydrolysis pattern of maltoheptaose with HSAmy was identical to the pattern obtained for the mixtures of HSAmy and AbpA variants. These results suggest that AbpA binds to HSAmy by leaving the active site open for the hydrolysis of G7. Comparison of the hydrolytic abilities of HSAmy showed that G7 was hydrolyzed to the same extent in the presence of PG1, PG2 and PG3 while HSAmy hydrolyzed G7 to a larger extent in the presence of PG4 and PG5.

Binding of AbpA variants to HSAmy using surface plasmon resonance spectroscopy

A representative sensorgram depicting the binding of HSAmy (0.5 μM) to the AbpA variants is shown in Figure 5. The binding efficiency was observed in the order PG1 >> PG3 > PG2 > PG4. Clearly, the sensorgram corresponding to PG4 shows that there is very little binding. Because of the unordered nature of PG5 and insensitivity of HSAmy to the presence of PG5 in starch hydrolysis (Figure 3), PG5 was not used in the Biacore experiments. In addition, the binding of HSAmy to the immobilized AbpA variants resulted in a dose-dependent increase in HSAmy binding for the mutants PG1, PG2 and PG3 (data not shown). Analysis of the binding data (BIAevaluation 3.2 software) indicated a weak affinity in the binding to PG2 coupled with a faster dissociation rate and a slower association rate (Table 1). In sharp contrast, the Kd for HSAmy binding to PG1 and PG3 were comparable (same order of magnitude) and suggests a very tight binding between HSAmy: PG1 and HSAmy: PG3. Interestingly, the absence of the C-terminal residues in PG3 does not affect the binding to HSAmy whereas the absence of the N-terminal residues in PG2 (residues 24-53) does affect the binding. However, PG4 which lacks the C-terminal domain (last 60 residues) completely lost its ability to bind HSAmy suggesting that residues 124-165 are essential to bind to HSAmy. Taken together, the results of these experiments suggest that some or all of residues 24-53 at the N-terminus and residues 124-165 at the C-terminal end might play a role in HSAmy interaction.

Discussion

CD spectra of AbpA variants

The CD spectra in the far-UV region are well known to reflect the secondary structure of proteins, although the tertiary structures have been known to influence the spectrum sometimes due to the interaction among aromatic residues. The mature sequence of AbpA does not contain many aromatic residues except a few in the N-terminal end and a very few at the C-terminal end. Thus, the observation of the typical spectrum corresponding to an α-helical protein is representative of the structure adopted by the molecule. Successive deletion of residues from the C-terminal end appears to increase the ratio of the helix vs. strand and other structures as the ellipticity value at 222 nm, a measure sometimes used to represent helicity increases in PG3. The structural
model obtained for AbpA (Figure 1) is somewhat limited in the absence of a definitive three-dimensional structure of AbpA. Nevertheless, it could be argued that the hypothetical model derived for AbpA has some validity since the results obtained through CD analyses support the preponderance of the helical nature obtained for the model.

Significance of the designed selective deletions and interaction between AbpA and HSAmy

Earlier structural studies have shown that HSAmy possesses multiple secondary oligosaccharide binding sites, which influence the binding to AbpA [20]. It was also noted that there exists similarity between bacterial binding sites and substrate-binding sites [26]. An interesting outcome of the combination of these studies is that both starch binding and AbpA binding to HSAmy might use multiple saccharide binding sites located on the surface of HSAmy. A further outcome of the recent study [20] on the role of aromatic residues in bacterial binding was that two aromatic residues (W203 and W284) might play a role in bacterial binding activity. Our results suggesting that both the N-terminal and the C-terminal helices in the hypothetical model of AbpA might be involved in binding to HSAmy is particularly interesting in this regard. If the model of AbpA is relevant for this discussion, it should be noted that the helix consisting of residues 34-45 (NDGAYLQFT) has two Tyr residues. When this segment is absent (PG2), then the SPR data suggest that there is weak affinity to HSAmy. It is likely that aromatic residues might dominate the interaction between HSamy and AbpA. In this regard, PG4 also has lost its ability to bind HSamy when the C-terminal residues (138-150) are absent (Figure 5). This segment contains two Tyr residues (TYR129 and TYR142), one of which occurs in a β-strand (TYIQ) and the second one in a helical segment (ANARYLKRYGAAN) further suggesting that aromatic residues in AbpA might play a role in the binding to HSamy.

It is tempting to speculate that these spatially well-separated aromatic residues in AbpA might be suitably juxtaposed to bind to the secondary saccharide sites harboring W203 and W284 in HSAmy. However, proof for this requires the validation of the model by x-ray structure analysis or other types of three-dimensional structure determination.

Conclusion

We show here that the AbpA might adopt a mostly helical structure and two segments in AbpA when absent decrease the binding to HSamy. The presence of aromatic residues in these segments points to the potential role of these residues in the binding interaction paralleling the role of aromatic residues on the surface of HSamy in binding to AbpA.

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