Prolonged Immunodeficiency in a Patient with Burkitt’s Lymphoma after Treatment with Rituximab and Chemotherapy

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Abstract

Common-variable immunodeficiency (CVID) is a heterogeneous disorder characterized by recurrent bacterial infections, hypogammaglobulinemia, and impaired antibody responses. Patients with CVID can remain undiagnosed for many years but carry an increased risk for developing B cell derived non-Hodgkin’s lymphomas, such as Burkitt’s lymphoma. Screening for CVID in patients diagnosed with Burkitt’s lymphoma is necessary before prescribing treatments, because certain regimens may significantly increase the risk for flare-up of CVID that could result in prolonged immunodeficiency. We present a patient who was diagnosed with Burkitt’s lymphoma and sustained prolonged severe immunodeficiency after receiving a complete course of rituximab and chemotherapy treatment. This case report illustrates the need for an immunological work-up in lymphoma patients before initiation of treatment because of the increased incidence of lymphoma in patients with immunodeficiencies. Patients with lymphoma should also be monitored longitudinally for immunodeficiencies after treatment with rituximab with or without chemotherapy.

Keywords: CVID; Burkitt’s lymphoma; Rituximab; Chemotherapy

Introduction

Burkitt’s lymphoma, a non-Hodgkin’s B cell lymphoma, is caused by translocation and subsequent deregulation of the c-myc oncogene on chromosome 8 [1]. Rituximab, a common treatment for Burkitt’s lymphoma, is an anti-CD20 chimeric antibody deprecating CD20-expressing B cells in lymphoid tissues and circulation. Adjuvant monoclonal antibody therapy with rituximab has successfully achieved improved outcomes for patients. Treatment with rituximab results in 90% reduction of CD20+ B cells. Its application is intended to eliminate malignant B cell clones [2]. The degree and duration of B cell depletion and its effects on immunoglobulin (Ig) levels in Burkitt’s lymphoma patients is not fully understood. There is even evidence that rituximab interventions may modulate the T cell compartment as well [3].

There is an increased risk of developing non-Hodgkin’s B cell lymphomas in patients with CVID. Patients with CVID often acquire inflammatory and autoimmune disorders, malignancies and systemic granuloma formations. Most of the malignancies affect the B cell compartment, with the development of non-Hodgkin’s B cell lymphomas posing a major risk [4]. Other patients develop symptoms of non-classic CVID such as defects in T cell development and secondary B cell deficiency [5]. Autoimmune manifestations are found in the form of cytopenias (usually acting as the initial presentation of CVID), such as immune thrombocytopenic purpura (ITP) and autoimmune hemolytic anemia (AIHA). A small proportion of CVID patients have defects in CD20 B cell surface protein. It has been suggested that CD20 dissociation from the BCR signalsome during early B cell activation might predispose individuals to CVID [6].

Acquired hypogammaglobulinemia is frequently concomitant with B cell derived non-Hodgkin’s lymphomas. Treatment of lymphomas with cytoreductive chemotherapy or drugs that can cause amplified protein loss can lead to acquired hypogammaglobulinemia [7]. Immunosuppressive drugs targeting B lymphocytes can additionally result in similar clinical effects. It is not well known whether rituximab, chemotherapy, or combined treatment leads to prolonged immunodeficiency. We present a patient with Burkitt’s lymphoma who developed severe and prolonged immunodeficiency after undergoing rituximab and chemotherapy treatment.

Case Presentation

A 43-year-old woman reported swelling in her right inguinal lymph node approximately five years ago, prompting a lymph node biopsy. A CT scan revealed a 5 cm soft tissue mass, and final pathology confirmed a diagnosis of Burkitt’s lymphoma nine days later. Prior to the start of her treatment regimen, her medical history was not significant for any instances of pneumonia or sinus infections and she had normal levels of white blood cells (5.8×10E3/μL), platelets (219×10E3/μL), absolute neutrophils (4.1×10E3/μL) and lymphocytes (1.2×10E3/μL) (Table 1). A comprehensive immunology laboratory assessment including immunoglobulins, however, was not administered before and after her diagnosis of Burkitt’s lymphoma.

After her diagnosis of lymphoma, the patient underwent a combination of chemotherapy and rituximab treatment for two-and-a-half months. Briefly, each treatment cycle consisted of CVP (cyclophosphamide-vincristine-prednisone) chemotherapy with high-dose cyclophosphamide (1500 mg/m²) and vincristine (1.4 mg/m²) on day, prednisone (100 mg) on days 1 through rituximab (375 mg/m²) on days 1 and 8, and high-dose methotrexate (3 g/m²) and leucovorin (25 mg) rescue on day 8. Additionally, filgrastim (G-CSF) (480 μg) was given.
given subcutaneously (SC) once daily beginning on day 3 and cytarabine intrathecally (IT) on days, 4 and 11. After two cycles, the patient continued with consolidation therapy consisting of rituximab and high-dose cyclophosphamide for 4 days, followed by rituximab once weekly and G-CSF SC daily until the end of treatment.

Approximately four months after her final treatment with rituximab, the patient began developing upper respiratory tract infections, discolored rhinorrhea and nasal congestion with a pattern characterized by brief, positive response to oral antibiotic followed by rapid relapse within two weeks. Over the next eight months, the patient had recurrent sinus infections every 2-4 weeks, requiring treatment with antibiotic courses. A CT scan done at that time revealed mucosal thickening in bilateral maxillary sinuses and thickening in her ethmoid cavity. She was then referred to our allergy and immunology clinic for an allergy and immunology work-up.

An allergy skin-prick test was positive for tree, grass, ragweed pollens and dust mites. Her initial immune work-up revealed low IgG (<60 mg/dL), IgA (<5 mg/dL), and IgM (<4 mg/dL). She demonstrated no response to all 14 serotypes of pneumococcal bacteria 4 weeks after vaccination with pneumovax. She was started on monthly intravenous immunoglobulin (IVIg) therapy (500 mg/kg). Her subsequent trough levels of IgG were normal, but as expected, IgA and IgM continued to be deficient (Table 1), because IVIg does not replace IgA and IgM. The patient continued with a significant deficiency in her white blood cell numbers (3.0×10E3/ μL), neutrophils (2.0×10E3/ μL), lymphocytes (0.7×10E3/μL) (Figure 1), platelets (112×10E3/ μL), and CD4 (266/μL) for a prolonged period of up to 42 months after finishing chemotherapy and rituximab treatment. The patient had a good response to the IVIg with an average of 1.5 sinus infections every year and no other major opportunistic infections. The results are summarized in Table 1.

![Figure 1: White blood cell, neutrophil, and lymphocyte counts in the patient before and after chemotherapy and rituximab.](image)

Normal ranges are as follows: WBC - 4.0-10.5×10E3/μL, Neutrophils - 1.8-7.8×10E3/ μL, Lymphocytes - 0.7-4.5×10E3/μL. BT refers to before treatment. X-axis numbers correspond to months after cessation of treatment.

### Table 1: Immunological laboratory values of the patient with Burkitt’s lymphoma before diagnosis and after cessation of treatment with chemotherapy and rituximab. Normal ranges are given as follows: 1(700-1600 mg/dL); 2(90-415 mg/dL); 3(40-230 mg/dL); 4(4.0-10.5 ×10E3/ μL); 5(150-415×10E3/μL); 6(1.8-7.8×10E3/μL); 7(0.7-4.5×10E3/μL); 8(359-1520 /μL). Month in leftmost column refers to elapsed time after ending treatment.

<table>
<thead>
<tr>
<th></th>
<th>IgG</th>
<th>IgA</th>
<th>IgM</th>
<th>WBC</th>
<th>Platelet</th>
<th>Neutrophils (abs)</th>
<th>Lymphocytes (abs)</th>
<th>CD4</th>
</tr>
</thead>
<tbody>
<tr>
<td>Before Initiation of Treatment</td>
<td>-</td>
<td>-</td>
<td>-</td>
<td>5.8</td>
<td>219</td>
<td>4.1</td>
<td>1.2</td>
<td>-</td>
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<tr>
<td>1 mo</td>
<td>-</td>
<td>-</td>
<td>-</td>
<td>3.1</td>
<td>69</td>
<td>2.4</td>
<td>0.29</td>
<td>-</td>
</tr>
<tr>
<td>13 mo</td>
<td>&lt;60</td>
<td>&lt;5</td>
<td>&lt;4</td>
<td>3.0</td>
<td>112</td>
<td>2.0</td>
<td>0.70</td>
<td>266</td>
</tr>
<tr>
<td>16 mo</td>
<td>27</td>
<td>&lt;5</td>
<td>&lt;4</td>
<td>2.0</td>
<td>103</td>
<td>1.7</td>
<td>0.5</td>
<td>206</td>
</tr>
<tr>
<td>23 mo</td>
<td>1097</td>
<td>11</td>
<td>4</td>
<td>1.8</td>
<td>119</td>
<td>1.0</td>
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<tr>
<td>27 mo</td>
<td>1279</td>
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<td>&lt;1</td>
<td>2.3</td>
<td>122</td>
<td>1.4</td>
<td>0.54</td>
<td>181</td>
</tr>
<tr>
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<td>1089</td>
<td>&lt;4</td>
<td>&lt;1</td>
<td>2.3</td>
<td>135</td>
<td>1.5</td>
<td>0.54</td>
<td>-</td>
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<tr>
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<td>&lt;4</td>
<td>2.0</td>
<td>129</td>
<td>1.0</td>
<td>0.60</td>
<td>-</td>
</tr>
</tbody>
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*Intravenous immunoglobulin (IVIg) treatment began.

## Discussion

Patients who have CVID are more prone to developing non-Hodgkin’s B cell lymphomas like Burkitt’s lymphoma, and incidences of lymphoma are more common in females than males [8]. Patients with CVID can go undiagnosed and remain asymptomatic for a number of years [9]. In the case presented, however, the patient likely developed the prolonged immunodeficiency secondary to treatment with Rituximab and / or chemotherapy and not due to preexisting CVID. The patient had no significant history of recurrent sinusitis, bronchitis or pneumonia prior to starting treatment with Rituximab and chemotherapy. This highlights the need for an immunological assessment in patients with lymphoma prior to their treatment with chemotherapy and/or rituximab to rule out preexisting CVID.

Rituximab typically does not lead to prolonged deficiencies in immunoglobulin levels because of the presence of long-lived plasma cells [10]. However, a study demonstrated prolonged
hypogammaglobulinemia after rituximab treatment in a subset of patients for up to 42 months [11]. In one case study, rituximab was found to aggravate underlying CVID in a 34-year-old male patient [12]. Routine baseline B-cell numbers and serum immunoglobulin levels before starting immunomodulatory therapy are required to help distinguish primary immunodeficiency diseases from secondary rituximab-induced, transient, and, at times, prolonged immune suppression [13]. It is unknown whether the addition of chemotherapy to rituximab can enhance the risk of developing prolonged hypogammaglobulinemia. Hence, patients treated with rituximab and/or chemotherapy should have serial laboratory assessments for earlier diagnosis of immunodeficiency. It possible that a subset of patients with a genetic predisposition may develop prolonged immunodeficiency after treatment with rituximab and/or chemotherapy.

This case report illustrates the need for an immunological work-up before initiation of treatment in all patients with lymphoma, because patients with CVID and other immunodeficiencies are at an increased risk for developing such malignancies. Our case report also demonstrates the need for serial immunological assessments in patients receiving rituximab with or without chemotherapy, because such treatments may lead to prolonged immunodeficiency in a subset of patients. Furthermore, our case report highlights the need for a larger, prospective study of patients receiving rituximab and/or chemotherapy for lymphomas or other autoimmune disorders to enhance our understanding on the risks of developing prolonged immunodeficiency due to such treatments.

References