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Photo-Bactericidal Property and Characterization of Cellulosic Fabric Treated with Two Tetra-Cationic Porphyrin Compounds

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Abstract

The cellulosic fabrics bearing the porphyrin as a photo-sensitizer were done in order to prepare efficient polymeric materials for antimicrobial applications. The obtained porphyrin-grafted cellulosic fabrics were characterized by ATR-FTIR, DRUV spectroscopy, TG and SEM image. The antimicrobial activity of the products was tested under visible light irradiation against *Staphylococcus aureus*, *Pseudomunas aeroginosa* and *Escherichia coli*.

Keywords: Cellulosic fabrics; Photo-bactericidal activity; Photodynamic Antimicrobial Chemotherapy; Tetra-cationic porphyrins; Thermo-gravimetric analysis; Zinc porphyrin compound.

Introduction

The photosensitizers (PS) such as porphyrin compounds have been intensively studied for their use as photo-bactericidal agents against both Gram negative and Gram positive bacteria in photodynamic antimicrobial chemotherapy (PACT) [1-5].

PACT relies on the intracellular accumulation of a (PS) upon illumination with visible light and produces singlet oxygen or generates free radicals. Singlet oxygen $({}^{1}O_{2})$ is able to react with almost every cellular ingredient, bringing about irreversible damage that ultimately leads to cell death [2]. This method has recently been studied against a wide range of clinically important bacteria, yeasts, fungi and viruses [6-9].

A number of water-soluble cationic porphyrins and phthalocyanine complexes of biocompatible metals (such as: Zinc, gallium and silicon), porphyrin compounds such as meso-tetrakis-(4-aminophenyl)-porphyrin, meso-tetrakis-(N-methylpyridyl)porphyrin and other photoactive dyes were used as biocompatible porphyrins for using in PACT. Photo-bactericidal cellulosic surfaces have been synthesized from cellulose and natural or synthetic porphyrins [1,2,10-13].

Due to the cellulose is a compatible molecule with no cytotoxic properties and perfect support for immobilization of bioactive molecules for application in medical and biological fields [14,15] we applied various concentrations of tetrakis(4-N,N,N-trimethylanilinium)porphyrin and its zinc metal ion complex to the cellulosic surface for the first time and characterized porphyrin-grafted cellulosic fabrics using various analysis methods (ATR-FITR, DRUV, SEM and TG). In addition, photo-bactericidal activity of these treated cellulosic fabrics was tested against *E. coli* (urinary tract infections [16]), *P. aeruginosa* (infection in patients with endotracheal and urinary cathetersis common [17]) and *S. aureus* (infection in patients

with prosthetic devices, venous catheters, and peritoneal dialysis catheters etc. [18]) under illumination with visible light.

Materials and Methods

All chemicals were purchased from Merck Company and used without further purification. The porphyrin, tetrakis(4-N,N,N-trimethylanilinium)porphyrin (TAPP) and its zinc ion complex (ZnTAPP) were synthesized as reported previously [19]. All fabrics were of plain (woven) construction, weighing 162.5 g/m², unfinished 100% cellulosic fabric, laundered, dried and the preparation of photoactive cellulosic fabrics and porphyrin treatment with them were done according to our previous article [20].

Bacterial strain and preparation of cultures

The bacterial strains *S. aureus, E. coli* and *P. aeruginosa* were obtained from microbiology laboratory of University of Guilan and were inoculated in liquid culture medium [nutrient broth (Source: Merck Company)] and incubated at 37° C overnight under aerobic conditions in an incubator. The stock suspensions of liquid culture medium were prepared approximately $\approx 10^{8}$ colony forming units/mL (CFU/mL). Antibacterial activity of photosensitive cellulosic fabric was done according to our previous article [20].

Irradiation system

All the experiments were carried out in a water-jacked reactor irradiated with a 100 W tungsten lamp (1250 lumen), as a visible light source with an average intensity of ~0.36 mWcm⁻² at a distance of 20 cm from the sample. To avoid light reflection, the reactor was placed in a dark room.

Sample characterization

UV-spectra were recorded on a UV-1700 pharma Spec (Shimadzu) with a quartz cuvette. ATR-FTIR spectra of untreated and treated cellulosic fabrics were recorded on Shimadzu FT-IR-8400S

spectrophotometer. DRUV spectra were prepared with a Shimadzu (MPC-2200) spectrophotometer. Thermo-gravimetric analyses of samples were carried out using a TGA V5.1A Dupont 2000 instrument with a heating rate of 10 $^{\circ}$ C/min in air and all samples were heated from 20-600 $^{\circ}$ C. Surface morphology of untreated and treated cellulosic samples was observed by SEM using a VEGA TESCAN scanning microscope. Electron micrographs of the sample were recorded at 600×magnification.

Results

The effect of photosensitizers on cellulose surface was confirmed by attenuated total reflectance Fourier transform infrared (ATR-FTIR) spectroscopy, diffuse reflectance UV-Vis (DRUV) spectroscopy, SEM and thermo-gravimetric analysis (TG).

ATR-FTIR spectra of treated and untreated cellulosic fabrics are presented in Figure 1a. Classic spectral data have been found: 3340 cm⁻¹ (OH stretching), 2850 and 2900 cm⁻¹ (symmetric and asymmetric C-H stretching), 1325, and 1045 cm⁻¹ (C-O stretching), (600-800) cm⁻¹ (alcoholic –OH out-of-plan bending, out-of-plan ring stretching in cellulose β -linkage) [21-24]. The samples of treated cellulose display a weak signal at 3340 cm⁻¹ and (600-800) cm⁻¹, corresponding to OH groups and the changes in the bands of (C-O stretching) at the range of (800-1325) cm⁻¹. These observations confirmed the linkage between porphyrins and the cellulosic fabric [1,25].

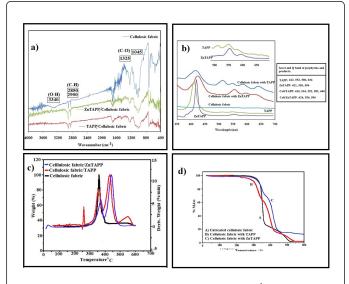


Figure 1: a) ATR-FTIR spectra (600-4000 cm⁻¹) of untreated cellulose, treated cellulose with TAPP⁴⁺ and treated cellulose with ZnTAPP⁴⁺; b) DRUV spectra (350-700 nm) of treated and untreated cellulose. Porphyrins spectra recorded at 2×10^{-6} M in water; c) TG and DTG thermograms of untreated and treated cotton fabric.

DRUV analysis of porphyrin solutions were shown Soret band near 412-424 nm and Q bands between 500-700 nm clearly show up (Figure 1b). Upon grafting the porphyrins onto cellulosic fabrics, Soret bands broadened that can be attributed to π -electron interaction with surface hydroxyl groups [9]. Therefore, DRUV analyses confirmed that porphyrins are either attached onto the surface via interaction between porphyrin and cellulose.

Thermo-gravimetric analyses (TG and DTG) were used to investigate the thermal properties of porphyrins attached to cellulose (Figure 1c and 1d). The pyrolysis of untreated cellulosic fabric comprises different major stages. The main pyrolysis stage of cellulosic fabric occurs in the temperature zone between 300-370°C [23,24].

At higher temperatures, treated cellulosic samples showed multistep mass loss due to decomposition of photo-sensitizers, or degradation of polymeric material or the backbone itself [25]. Also, residual mass percent of samples at 600°C were 15.08% and 1.74% respectively for ZnTAPP⁴⁺ and TAPP⁴⁺, whereas that for untreated cotton fabric was 1.63%.

The surface of untreated cellulose and porphyrin-grafted cellulose was examined by scanning electron microscopy (SEM) and did not show any destruction of cellulose fibers (similar diameter and structure) (Figure 2).

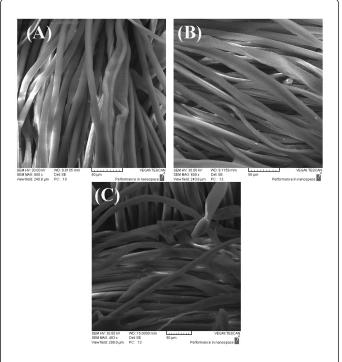
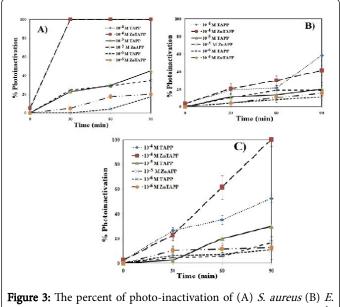


Figure 2: SEM photomicrographs (50 µm scale) of cellulose fibers. Untreated cellulosic fabric (a) and treated cellulosic fabric with TAPP4+ (b) ZnTAPP4+.

Photodynamic activity of treated cellulose was evaluated *in vitro* against bacteria. Extensive research on the effect of singlet oxygen on photo-inactivation of bacteria [11,26-29], indicates that singlet oxygen diffusion is an effective factor. Figure 3 shows the percentage of photo-inactivation against three bacterial strains. Untreated samples in the dark allow bacterial growth; but *S. aureus* growth under light irradiation for 90 min on untreated samples was reduced by about ~12.7%. Samples treated with ZnTAPP⁴⁺ and TAPP⁴⁺ (in the dark) allow bacterial growth and have little effect on reducing the number of bacteria; about 1-6%. Both types of treated cellulose have photobactericidal activities against *S. aureus*. At low concentrations, these compounds have photo-inactivation property against this strain. *E. coli* and *P. aeruginosa* are Gram-negative bacteria and are less susceptible to PACT than Gram-positive bacteria. ZnTAPP⁴⁺ ([PS]= 10^{-4} M, 90

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min) exhibits photo-bactericidal effect against *P. aeruginosa* whereas TAPP4+ exhibits 52.4% photo-inactivation. It seems that the Zinc atom in the TAPP⁴⁺ is clearly playing a synergistic inhibitory role in bacterial growth.



coli and (C) *P. aeruginosa* by treated cellulosic fabric with TAPP⁴⁺ and ZnTAPP⁴⁺.

Furthermore, both products have photo-inactivation effect on *E. coli.* The percentages of photo-inactivation for TAPP4+ were: 19%, 21.6% and 58.5% and for ZnTAPP⁴⁺ they were: 20.7%, 30% and 41.3% at a concentration of 10-4 M of [PS] after 30, 60 and 90 min illumination.

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References

- Ringot C, Sol V, Barrière M, Saad N, Bressollier P, et al. (2011) Triazinyl porphyrin-based photoactive cotton fabrics: preparation, characterization, and antibacterial activity. Biomacromolecules 12: 716-723.
- 2. Krouit M, Granet R, Krausz P (2009) Photobactericidal films from porphyrins grafted to alkylated cellulose-synthesis and bactericidal properties. Eur Polym J 45: 1250-1259.
- 3. Dosselli R, Tampieri C, Ruiz-González R, De Munari S, Ragàs X, et al. (2013) Synthesis, characterization, and photoinduced antibacterial activity of porphyrin-type photosensitizers conjugated to the antimicrobial peptide apidaecin 1b. J Med Chem 56: 1052-1063.
- 4. Nitzan Y, Ashkenazi H (2001) Photoinactivation of Acinetobacter baumannii and Escherichia coli B by a cationic hydrophilic porphyrin at various light wavelengths. Curr Microbiol 42: 408-414.
- Muñoz-Bonilla A, Fernández-García M (2012) Polymeric materials with antimicrobial activity. Prog Polym Sci 37: 281-339.
- 6. Grinholc M, Szramka B, Olender K, Graczyk A (2007) Bactericidal effect of photodynamic therapy against methicillin-resist- ant Staphylococcus aureus strain with the use of various porphyrin photosensitizers. Acta Biochim Pol 54: 665-670.

- 7. Cassidy CM, Donnelly RF, Tunney MM (2010) Effect of sub-lethal challenge with Photodynamic Antimicrobial Chemotherapy (PACT) on the antibiotic susceptibility of clinical bacterial isolates. J Photochem Photobiol B Biol 99: 62-66.
- 8. Parsons C, McCoy CP, Gorman SP, Jones DS, Bell SE, et al. (2009) Antiinfective photodynamic biomaterials for the prevention of intraocular lens-associated infectious endophthalmitis. Biomaterials 30: 597-602.
- Banfi S, Caruso E, Buccafurni L, Battini V, Zazzaron S, et al. (2006) Antibacterial activity of tetraaryl-porphyrin photosensitizers: an in vitro study on Gram negative and Gram positive bacteria. J Photochem Photobiol B Biol 85: 28-38.
- 10. Memmi A, Granet R, Aouni M, Bakhrouf A (2012) Synthesis of new photobactericidal polymers by " click chemistry " and a study of their biological activity. e-Polymers: 1-12.
- Krouit M, Granet R, Krausz P (2008) Photobactericidal plastic films based on cellulose esterified by chloroacetate and a cationic porphyrin. Bioorg Med Chem 16: 10091-10107.
- 12. Ringot C, Sol V, Granet R, Krausz P (2009) Porphyrin-grafted cellulose fabric: New photobactericidal material obtained by "Click-Chemistry" reaction. Mater Lett 63: 1889-1891.
- 13. Krouit M, Granet R, Branland P, Verneuil B, Krausz P (2006) New photoantimicrobial films composed of porphyrinated lipophilic cellulose esters. Bioorg Med Chem Lett 16: 1651-1655.
- Wang C, Venditti RA, Zhang K (2015) Tailor-made functional surfaces based on cellulose-derived materials. Appl Microbiol Biotechnol 99: 5791-5799.
- 15. Kang H, Liu R, Huang Y (2013) Cellulose derivatives and graft copolymers as blocks for functional materials. Polym Int 62: 338-344.
- 16. Nicolle LE (2005) Catheter-related urinary tract infection. Antimicrob Resist Infect Control 22: 627-639.
- Ortona L, Federico G, Fantoni M, Ardito F, Branca G, et al. (1985) A study on the incidence of nosocomial infections in a large university hospital. Eur J Epidemiol 1: 94-99.
- Dasgupta MK, Lam K, Ulan RA, Bettcher KB, Burns V, et al. (1998) An extracorporeal model of biofilm-adherent bacterial microcolony colonization for the study of peritonitis in continuous ambulatory peritoneal dialy- sis. Am J Nephrol 8: 118-122.
- 19. Krishnamurthy M (1997) Synthesis and characterization of a new watersoluble porphyrin. Indian J Chem 15: 964-966.
- Rahimi R, Fayyaz F, Rassa M (2016) The study of cellulosic fabrics impregnated with porphyrin compounds for use as photo-bactericidal polymers. Mater Sci Eng C 59: 661-668.
- 21. Allen A, Foulk J, Gamble G (2007) Preliminary Fourier-Transform Infrared Spectroscopy Analysis of Cotton Trash. J Cotton Sci 11: 68-74.
- 22. Chung C, Lee M, Choe EK (2004) Characterization of cotton fabric scouring by FT-IR ATR spectroscopy. Carbohydr Polym 58: 417-420.
- Hu S, Hu Y, Song L, Lu H (2012) The potential of ferric pyrophosphate for influencing the thermal degradation of cotton fabrics. J Therm Anal Calorim 109: 27-32.
- 24. Alongi J, Colleoni C, Rosace G, Malucelli G (2012) Thermal and fire stability of cotton fabrics coated with hybrid phosphorus-doped silica films. J Therm Anal Calorim 110: 1207-1216.
- Mbakidi JP, Herke K, Alvès S, Chaleix V, Granet R (2013) Synthesis and photobiocidal properties of cationic porphyrin-grafted paper. Carbohydr Polym 91: 333-338.
- Xing C, Xu Q, Tang H, Liu L, Wang S (2009) Conjugated polymer/ porphyrin complexes for efficient energy transfer and improving lightactivated antibacterial activity. J Am Chem Soc 131: 13117-13124.
- Feese E, Sadeghifar H, Gracz HS, Argyropoulos DS, R.A.G. (2011) Photobactericidal Porphyrin-Cellulose Nanocrystals: Synthesis, Characterization, and Antimicrobial Properties. Biomacromolecules 12: 3528-3539.
- Romanova NA, Brovko LY, Moore L, Pometun E, Savitsky AP, et al. (2003) Assessment of Photodynamic Destruction of Escherichia coli O

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157: H7 and Listeria monocytogenes by Using ATP Bioluminescence. Appl. Environ. Microbiol 69: 6393-6398. 29. Wainwright M (2000) Methylene blue derivatives-suitable photoantimicrobials for blood product disinfection? Int J Antimicrob Agents 16: 381-394.