

Post-translational Modifications of Proteins in Metabolic Syndrome

Jorge Suarez^{1*} and Julieta Díaz-Juárez²

¹Department of Medicine, University of California, San Diego, USA

²Department of Pharmacology, Instituto Nacional de Cardiología, "Ignacio Chávez", México

Metabolic syndrome is accompanied by central obesity, dyslipidemia, compromised fasting glucose, and hypertension [1]. Unfortunately, all of these factors contribute to damage the endothelium that in turn, will conclude in the development of multiple complications observed in the metabolic syndrome. Endothelial dysfunction is mainly caused by a decrease in nitric oxide (NO) availability due to reduced NO production and/or increase in oxygen-derived free radicals (ROS) that can react with NO and inactivate the active molecule [2]. NO production in endothelial cells is mainly mediated by the endothelial isoform of NO synthase (eNOS), therefore, studies that investigate regulatory mechanisms of this enzyme are essential. Currently, the influence of metabolic syndrome on eNOS regulation is incompletely investigated. Recently, Guterbaum et al. [3] published a paper in this journal that describes the effects of H₂O₂ on phosphorylation of the eNOS of endothelial cells pretreated with supra-physiologic glucose concentrations. Their findings demonstrated that H₂O₂, with the concomitant increase ROS production, resulted in an increase in Thr495 phosphorylation while phosphorylation of Ser1177 was reduced. Furthermore, these authors demonstrated that combination of high glucose concentration with H₂O₂ induces phosphorylation of Thr495 through the PKC pathway. These phosphorylation sites confere fine regulation of eNOS activity [4] and the findings by Guterbaum et al. provide bases to understand more the complexity of pathophysiologic mechanisms that characterize the metabolic syndrome.

Post-translational regulation of eNOS, including phosphorylation, is a growing field that increased the complexity of endothelial function and the maladaptive effects resulting from the metabolic syndrome. Furthermore, integrating other post-translational modifications of proteins can complicate the picture even more. O-GlcNAcylation of serine or threonine residues of nuclear, cytoplasmic and mitochondrial proteins is a dynamic and ubiquitous protein modification. Protein O-GlcNAcylation is emerging as a key regulator of critical biological processes including nuclear transport, translation and transcription, signal transduction, cytoskeletal reorganization, proteasomal degradation, and apoptosis [5-9]. There is a complex interplay between phosphorylation and O-GlcNAcylation [10-13]. Increased levels of O-GlcNAcylation are a pathogenic contributor to glucose toxicity and insulin resistance. O-GlcNAcylation contributes to the adverse effects of diabetes on cardiovascular function as well as mediating the response to ischemic injury. Consequently, it is not surprising that O-GlcNAcylation can impair the activity of eNOS [14,15]. Further work is needed to understand these complicated pathways and to identify therapeutic approaches to treat the metabolic syndrome.

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References

1. Tziomalos K, Athyros VG, Karagiannis A, Mikhailidis DP (2010) Endothelial dysfunction in metabolic syndrome: Prevalence, pathogenesis and management. *Nutr Metab Cardiovasc Dis* 20: 140-146.
2. Deanfield JE, Halcox JP, Rabelink TJ (2007) Endothelial function and dysfunction: testing and clinical relevance. *Circulation* 115: 1285-1295.
3. Guterbaum TJ, Braunstein TH, Fossum A, Holstein-Rathlou NH, Torp-Pedersen CT, et al. (2013) Endothelial nitric oxide synthase phosphorylation at Threonine 495 and mitochondrial reactive oxygen species formation in response to a high H₂O₂ concentration. *J Vasc Res* 50: 410-420.
4. Guterbaum JT, Thomas HB, Fossum A, Holstein-Rathlou NH, Torp-Pedersen C (2015) H₂O₂ Treatment of HUVECs Facilitates PKC Mediated Thr495 Phosphorylation on eNOS when Pre-treated with High Glucose Levels. *J Metabolic Synd* 4: 189.
5. Qian J, Fulton D (2013) Post-translational regulation of endothelial nitric oxide synthase in vascular endothelium. *Front Physiol* 4: 347.
6. Laczy B, Hill BG, Wang K, Paterson AJ, White CR, et al. (2009) Protein O-GlcNAcylation: a new signaling paradigm for the cardiovascular system. *Am J Physiol Heart Circ Physiol* 296: H13-28.
7. Butkinaree C, Park K, Hart GW (2010) O-linked [beta]-N-acetylglucosamine (O-GlcNAc): Extensive crosstalk with phosphorylation to regulate signaling and transcription in response to nutrients and stress. *Biochim Biophys Acta* 1800: 96-106.
8. Hu Y, Suarez J, Fricovsky E, Wang H, Scott BT, et al. (2009) Increased enzymatic O-GlcNAcylation of mitochondrial proteins impairs mitochondrial function in cardiac myocytes exposed to high glucose. *J Biol Chem* 284: 547-555.
9. Kreppel LK, Blomberg MA, Hart GW (1997) Dynamic glycosylation of nuclear and cytosolic proteins. Cloning and characterization of a unique O-GlcNAc transferase with multiple tetratricopeptide repeats. *J Biol Chem* 272: 9308-9315.
10. Ngoh GA, Facundo HT, Zafir A, Jones SP (2010) O-GlcNAc signaling in the cardiovascular system. *Circ Res* 107: 171-185.
11. Hart GW (1996) O-GlcNAcylation of key nuclear and cytoskeletal proteins: reciprocity with O-phosphorylation and putative roles in protein multimerization. *Glycobiology* 6: 711-716.
12. Hu P, Shimoji S, Hart GW (2010) Site-specific interplay between O-GlcNAcylation and phosphorylation in cellular regulation. *FEBS Lett* 584: 2526-2538.
13. Zeidan Q, Hart GW (2010) The intersections between O-GlcNAcylation and phosphorylation: implications for multiple signaling pathways. *J Cell Sci* 123: 13-22.
14. Wang Z, Gucek M, Hart GW (2008) Cross-talk between GlcNAcylation and phosphorylation: Site specific phosphorylation dynamics in response to globally elevated O-GlcNAc. *Proceedings of the National Academy of Sciences* 105: 13793-13798
15. Beleznaï T, Bagi Z (2012) Activation of hexosamine pathway impairs nitric oxide (NO)-dependent arteriolar dilations by increased protein O-GlcNAcylation. *Vascular Pharmacology* 56: 115-121.

*Corresponding author: Jorge Suarez, Research Scientist Department of Medicine, University of California, San Diego, California, USA, Tel: (858) 534-9931; Fax: (858) 534-9932; E-mail: jsuarez@ucsd.edu

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