

Understanding the Challenges of Melanogenesis: Key Role of Bioactive Compounds in the Treatment of Hyperpigmentary Disorders

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Abstract

Skin pigmentation is an important human phenotypic trait that gives special aesthetic hue to individuals and also protective covering against solar ultra violet radiations, but excessive pigment production and uneven distribution, appear as serious undesirable hyperpigmentary anomalies. To overcome this, various therapeutic agents and skin-lightening cosmetics are in big demand across Asia, and the quest for fairness has led to identification of many new ingredients. The mechanisms underlying pigmentation has been researched extensively and the knowledge are being updated regularly. To understand the etiology of normal and uneven skin pigmentation, first part of the review serves us basic updated information regarding the normal process of melanogenesis and strict coherent attributes that monitor the skin pigmentation control machinery. While the later part of the review focuses on some abnormal hyperpigmentary anomalies, their mode of action at the cellular level and various approaches that exploit the natural botanicals for their permanent and cost effective treatment.

Keywords: Skin pigmentation; Hyperpigmentation disorders; Melanogenesis; Natural products/botanicals

Introduction

Melanin is a prime contributor of pigmentation patterns in vertebrates including human beings and is synthesized by melanocytes which remain distributed in the basal layer of the skin epidermis. Generally two types of melanin pigment are produced within the melanocytes, i.e. black or brown eu-melanin and red or yellow pheomelanin, which decides the colour of hair, skin or eyes [1,2].

Upon exposure to the UV radiations, melanocytes synthesise melanin in the membrane bound organelle, the melanosomes by multistep, complex process of melanogenesis, and transfer them via their dendritic process to the adjacent keratinocytes where they form perinuclear melanin cap to protect the cells from UV induced damage of nuclear DNA and also by removing the reactive oxygen species (ROS). Moreover in keratinocytes, these melanin pigments give colour to skin in combination with other pigments like carotenoid and haemoglobin derivatives [3].

Melanin synthesis is highly cooperative step carried out by tyrosinase gene family protein i.e. tyrosinase (TYR), tyrosinase related protein-1 (TRP-1) and tyrosinase related protein-2 (TRP-2). Though TYR is the key player of melanogenesis that catalyse the crucial two initial steps of melanin synthesis, TRP-1 and TRP-2 also take important position in the synthesis of melanin [4,5].

Melanogenesis is under a mosaic regulatory control comprising interplay of different factors and signaling pathways. Among them α - melanocyte stimulating hormone (α -MSH), agouti signal protein (ASP) and microphthalmia associated transcriptional factor (MITF) are the predominant regulators of melanin synthetic pathways. Conjointly, one of such regulatory factors or pathways is the discordant

involvement or subsidiary projection of tyrosinase via unorthodox routes resulting into varied skin related abnormalities [6,7].

Hyper reactivity of melanocytes as well as abnormal synthesis and accumulation of melanin leads to the consequences of skin related pigmentation disorders. Looking at the human history, humans have continually tagged and categorized end another on the basis of skin complexion. In most African and Asian populations, fair complexion is labelled/tagged as beauty, grace and high social class whereas dark skin complexion is seen as being of lowest social status. This perception inspires most people of both genders for the recent craze for a fair skin and has given a forceful indulgence of their in skin care merchandise, that improve the skin complexion [8].

The present review is dedicated to the updated knowledge of melanocytes biology including (1) biogenesis of melanosomes and melanin (2) regulation of melanogenesis, (3) hyperpigmentary disorders (4) scientifically validated plants for the treatment of hyperpigmentation, and their mechanism.

Biogenesis of Melanosomes and Melanin

Melanin synthesis starts concurrently with the biosynthesis of specialized intracellular organelles, the melanosomes. Melanosomes are formed by multistep process during which immature, unpigmented melanosomes matured into developed pigmented melanosomes. Based on electron microscopic observations by Seiji, [9] melanosome biogenesis has been categorized into four stages (I-IV), based on the degree of maturation with discrete morphological and biochemical features that reveal discrete routes of the biosynthesis of structural and enzymatic proteins. Hence, melanin production occurs in these particular compartments as the biogenesis of melanosomes proceeds. Early stages (I & II) of melanosome development are referred as premelanosomes which lack pigment but possess other distinct features. Stage I premelanosomes originate from endosomes, are electron lucent and are limited by unit membrane which matures into

stage II premelanosomes. Pmel17 is a transmembrane glycoprotein that localizes in the limiting membrane of stage I premelanosomes and then enters into the matrix of stage II premelanosomes, forms the intraluminal striations here, the formation of intraluminal fibrils being at stage I which get completed in stage II [10].

When the internal striation is formed, the following transport of integral membrane proteins take place: for example enzymes TYR, TRP-1 and TRP-2 begin the production of melanin that conceals the Pmel17-interluminal striation, ultimately melanosomal development proceeds towards stage III melanosomes which is partially pigmented [11]. TYR is the key enzyme of melanin synthesis [12] as it catalyses the initial two crucial steps of melanogenesis, i.e. the hydroxylation of L-tyrosine to 3,4-dihydroxyphenylalanine (L-DOPA), and subsequent oxidation of L-DOPA to DOPA-quinone. DOPA-quinone is the branching point to form either eumelanin or pheomelanin. In the absence of thio compounds like cysteine or glutathione (GSH), DOPA-quinone gets converted into DOPA-chrome. DOPA-chrome either tautomerize into indole 5,6-quinone 2-carboxylic acid (DHICA) the presence of TRP-2 or decarboxylated into 5,6-dihydroxyindole (DHI). TRP-1 is a DHICA oxidase which catalyses the formation of carboxy group containing eumelanin. DHICA and DHI are further oxidized and polymerized into eumelanin [13].

In the presence of thiol compound, the sulfhydryl group of these compound conjugate with DOPA-quinone to form 5-S-cysteinyl-dopa or 5-S-gluthionyl-dopa (in the presence of cysteine or glutathione). Thiol-DOPA gets subsequently oxidized and is polymerised into pheomelanin [14]. Eumelanosomes are elliptical or oval in shape whereas pheomelanosomes are of spherical shape. Fully pigmented stage IV melanosomes are generated after additional synthesis of melanin. These fully melanized stage IV melanosomes are then transported from melanocytic dendrites to the keratinocytes.

Crucial Factors Regulating Melanin Synthesis

Pigmentation factors that regulate biosynthesis of melanin within melanocytes are typically situated within, on, or near the melanosomes. These factors can be divided into three types i.e. protein that regulated melanosomes development, proteins that control melanin synthesis and proteins involved in intracellular trafficking of melanosomal components as well as their transport to the adjacent keratinocytes [15].

Melanosomal structural proteins

PMEL17/GP100 and MART-1 are the important structural proteins that play principal role in biogenesis of melanosomes. PMEL17 is an integral membrane protein that endures transfer from the endoplasmic reticulum to the Golgi apparatus where it is glycosylated and eventually exported to the limiting membrane of stage I premelanosomes [16]. Here it undergoes proteolytic cleavage and forms intraluminal striation of stage II premelanosomes, melanin pigment mask these fibrils and ultimately mature stages III and IV melanosomes formed. MART-1 is considered as melanosome specific marker and is found plentiful in the early melanosomes, is obligatory for the maturation of PMEL17 [17].

Enzymes / Proteins that control melanin biosynthesis

The three important enzymes that take part in melanin synthesis belong to tyrosinase gene protein family. These are tyrosinase (TYR), tyrosinase related protein-1 (TRP-1) and tyrosinase related protein-2

or dopachrome tautomerase (TRP-2 or DCT). All the three enzymes of melanin synthesis are transcriptional products of microphthalmia associated transcriptional factor (MITF) which is also known as “master regulator” of melanogenesis. Under normal conditions, after getting stimulation from UV radiation, MITF ensures the uninterrupted transcription of tyrosinase genes via cAMP signalling etc. [18]. Instead of MITF, many factors including biogenesis of lysosome-related organelles complexes (BLOC-1), ocular albinism type 1 protein (OA1), oculocutaneous albinism II (OCA2 also known as P) and SLC45A2 affect the trafficking and hence the function of these enzymes [10,19].

Receptors and proteins involved in melanosomal transport

After the deposition of melanin granules within the melanosomes, these are being migrated through microtubules (MT) from perinuclear zone to dendritic tips of melanocytes for transfer into the neighbouring keratinocytes. In addition to the MT, proteins like kinesin (prograde) and dynein (retrograde) seem to participate in trafficking of melanosomes [20].

A combination of genetic and cell biological studies have revealed that the activated small GTPase Rab27a together with its effector protein melanophilin, associates with mature melanosomes and recruits the actin motor protein, myosin VA (MYO5A) expressed in melanocytes. These proteins connected to the mature melanosomes in the order of Rab27A-melanophilin-MYO5A, this complex enables migration of melanosomes from microtubules onto the peripheral actin network of melanocytes, enabling successive transport to the adjacent keratinocytes [21-22].

Previous pioneering research work done in our laboratory, related to skin pigmentation has revealed that the major signals involved in pigment transport within the pigment producing cells are dependent on a special class of cell surface receptors called G-protein-coupled receptors (GPCRs). Many of these receptors i.e. adrenergic [23-27], cholinergic or cholinomuscarinic [28-34], histaminergic [35-39], serotonergic [40] have been found on melanophores/melanocytes of lower vertebrate species that regulate the translocation of pigment granules from one place to another. These large numbers of studies have also demonstrated the physiological and pharmacological role of autonomic receptors from a comparative evolutionary aspect of skin pigmentation. Our data categorically point that, at the cellular and molecular level, receptors particularly those of histamine (H1, H2, H3 and H4) and serotonin (5HT1, 5HT2, 5HT3 and 5HT4) play specific role in animal pigmentation including mammalian melanocytes Figure 1.

On the other hand endothelin and melatonin receptors have also found to be present on melanocytes, regulating translocation of pigment granules leading to control of skin pigmentation or phenotypical appearance of skin [41].

Hyperpigmentation of skin

Hyperpigmentation may be defined as increased pigmentation of the skin, which is generally recognized by darkening or increase in the natural color of the skin usually due to an increased deposition of melanin pigment in the epidermis and/or dermis [42]. On the basis of location of over deposition of melanin granules, hyper pigmentation can be classified as epidermal and dermal hyper pigmentation [43]. Epidermal hyper pigmentation is because of melanin pigmentation and has a brownish hue, while, dermal pigmentation is called

'ceruloderma' or 'blue hyper pigmentation' which may either be due to melanin or due to non-melanin pigments [44].

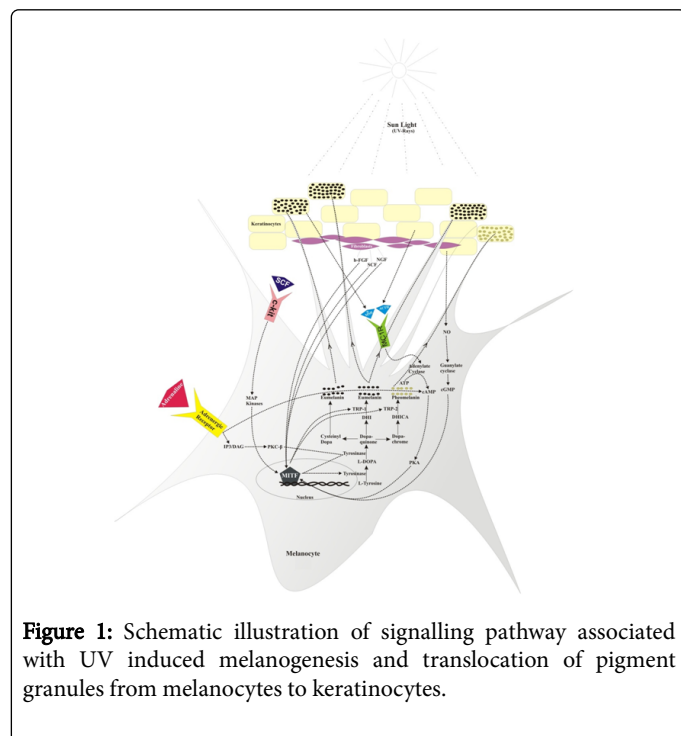


Figure 1: Schematic illustration of signalling pathway associated with UV induced melanogenesis and translocation of pigment granules from melanocytes to keratinocytes.

Hyper pigmentation also results from inflammation or other skin insults i.e. skin diseases such as acne or shingles with may leave darkened spots. Scars from skin injury or surgery also may become hyper pigmented. Cosmetic procedures including laser resurfacing, laser hair removal, chemical peels and dermabrasion also may leave the affected area darker than the normal skin color [45].

Although hyper pigmentation is not harmful, it can cause significant cosmetic disfigurement and become a persistent psychosocial burden for the patient, due to the limited efficacy of the available treatments. Apart from this, disfiguring facial lesions can significantly affect a person's overall emotional well-being and can contribute to decreases in social functioning, productivity at work or school, and self-esteem [46,47].

Mechanism of Hyperpigmentation

Abnormal pigmentation or hyper pigmentation may develop at a number of points in the biochemical cascades that regulate healthy pigmentation [48]. It typically results from increased melanin production by existing melanocytes (melanotic hyper pigmentation), which may occur in the epidermis/dermis or in both or from proliferation of active melanocytes (melanocytotic hyper pigmentation). Hyper pigmentation also may develop when epidermal cells retain too much melanin.

The main causes of skin hyper pigmentation are various: auto immune conditions, sun damage (UV radiation and ionizing radiation), drug reactions (chemicals), hormonal changes (extensive release of α -MSH), genetic factors, medications, hormonal therapy or birth control pills and also rubbing of the skin which can be the triggering factors, resulting in the hyper secretion of melanin from melanocytes causing hyper pigmentation [49-52].

Normally, the melanocytes are located in the basal layer of the epidermis and an increase in their number or activity will cause epidermal hyper pigmentation. However, formed melanin may be transferred to the dermis or, in some cases, dermal melanocytes are also present. A heightened activity or number of melanocytes in these instances will lead to dermal hyper pigmentation. Also, a combination of the above may take place, triggering mixed hyper pigmentation.

Hyperpigmentation Disorders

Hyper pigmentation disorders are characterized by darker skin appearance i.e. brown to gray-brown patches on the face melasma [45,53], dark plaques on skin, (post inflammatory hyper pigmentation) [53-55], light to dark brown spots of 1 to 20 cm ("Café-au-lait" macules)[53], pale brown to dark brown spots on the skin (agespots or solar lentigo)[53], well-circumscribed round to oval or irregular grey patches on the face, neck and trunk (Erythema dyschromicum perstans), netlike pigmentation (Prurigo pigmentosa)[56,57], dark vertical line across the abdomen in pregnant females (linea nigra) [58].

These skin pigmentary abnormalities are seen as aesthetically unfavourable and have led to the development of cosmetic and therapeutic treatment modalities of varying efficacy aimed to modulate skin pigmentation. These depigmenting agents are any ingredient or combination of ingredients that interfere one or more steps of melanogenic pathway, melanosome transfer or post-transfer pigment processing and degradation that result in lowering pigmentation on the surface of the skin [59,60].

Many modalities of treatment for acquired skin hyperpigmentation are available including chemical agents or physical therapies, but none are completely satisfactory. Traditional depigmenting agents, such as hydroquinone, corticosteroids, and kojic acid, although highly effective but can raise several safety concerns (i.e. ochronosis, atrophy, carcinogenesis, and other local or systemic side effects) with long-term exposure [61,62].

To overcome these adverse side effects, benefits of natural and botanical extracts can be used, which provides opportunities to develop new products as novel treatment for various hyperpigmentary anomalies, as medicinal plants are rich sources of bioactive chemicals, free from harmful side effects and are potentially safe and effective skin lightening agents [52,63].

Active compounds isolated from plants, such as arbutin, aloesin, gentisic acid, flavonoids, hesperidin, licorice, niacinamide, yeast derivatives and polyphenols, inhibit melanogenesis without melanocyte toxicity by different mechanisms i.e. acting on one or more steps in the melanogenic pathway, melanosome transfer, or post-transfer pigment processing and degradation.

Approaches for Treatment of Hyperpigmentation by Natural Botanicals

Inhibition of tyrosinase and related enzyme expression

The transcriptional level is the first stage by which the expression of tyrosinase and related melanogenic enzymes may be modulated [64]. To develop a new skin whitening agent for cosmetics and treatment of hyperpigmentation from natural products various phyto molecules have been screened in past few decades which exerting inhibitory effect on formation of tyrosinase, tyrosinase related protein-1 (TRP1)

and tyrosinase related protein-2 (TRP-2), enzymes that catalyses few rate limiting steps of melanogenic pathway [65].

Studies of Cho et al. [66] have demonstrated that isoimperatorin and imperatorin isolated from ethanolic extract of *Angelica daburica* significantly inhibited tyrosinase synthesis in B16 melanoma cells by reducing the level of tyrosinase m-RNA leading to lightening of the skin (Table 1).

Inhibition of tyrosinase activity

Most botanical depigmenting agents act by interfering in the pathways leading to melanin synthesis by inhibiting the activity of tyrosinase [65]. These kinds of tyrosinase inhibitors are widespread in nature and botanical extracts are good source of naturally occurring tyrosinase inhibitors which are commonly used as traditional remedies for pigmentation disorders [67-68].

Regarding this, some earlier studies have suggested that arbutin, isolated from the fresh fruit of the California buckeye and *Aesculus californica*, was used to inhibit the oxidation of L-DOPA catalyzed by mushroom tyrosinase and was effective in the topical treatment of various cutaneous hyperpigmentary disorders characterized by hyperactive melanocyte function [69-71].

Green tea is also reported to be a competitive tyrosinase inhibitor based on *in vitro* studies. The gallic catechin moiety in the major catechin constituents epicatechin gallate, epigallocatechin gallate and gallic catechin gallate is reported to be responsible for this effect [72].

Studies of Picardo and Carrera, [73] have demonstrated that aloein from *Aloe vera* works as a non-competitive inhibitor of tyrosinase, affecting the action of tyrosinase complex in the substratum and reducing the conversion of DOPA into melanin. Kojic acid also inhibits tyrosinase directly, while L-ascorbic acid and its derivatives are believed to act as reducing agents on intermediates in melanin biosynthesis at various points in the oxidation chain reaction from tyrosine/DOPA to melanin. Green tea is also reported to be a competitive tyrosinase inhibitor based on *in vitro* studies.

Tan et al. [74] and Cheng et al. [75] have reported that aloin isolated from leaf extract of *A. vera* acts as a natural skin lightener which can bind not only to the enzyme tyrosinase but also to the enzyme-substrate complex, leading to inactivation of the enzyme resulting lightening of skin.

Glycyrrhiza glabra is a commonly used plant in the skin lightening industry, which works to inhibit the enzyme tyrosinase and limit the amount of pigment produced in melanocyte cells. Previous studies of Nerya et al. [76] and Fu et al. [77] showed that glabridin and isoliquiritigenin, the main ingredients of the hydrophobic fraction of licorice extract have been also shown to inhibit tyrosinase activity in B16 murine melanoma cells. Research also indicated that skin lightening potential of glabridin is found to be greater than that of hydroquinone, the artificial depigmentation agent widely used [76, 78].

Lee et al. [79] have reported that mulberroside F, the active component of dried mulberry (*Morus alba*) leaves also showed inhibitory effects on tyrosinase activity and on melanin formation in melan-a cells. This compound also exhibited superoxide scavenging activity that is involved in the protection against auto-oxidation [80], suggesting a role for *M. alba* as a component of lightening cosmetics.

Yamakoshi et al. [81] have reported the oral administration of a proanthocyanidin rich extract from grape seeds for one year reduced

effectively the hyperpigmentation of women with chloasma. Ellagic acid is a natural polyphenol that is widely found in fruits and vegetables and the main active ingredient of fruit rinds of pomegranate, showed inhibitory activity in the process of melanogenesis by inhibition of the proliferation of melanocytes and melanin synthesis by tyrosinase in melanocytes [82].

Hesperidin, a bioflavonoid existing extensively in the peel and membranes of citrus fruits also shows potent dose-dependent antityrosinase activity of hesperidin in B16 melanoma cells leading to inhibition of melanin synthesis without cytotoxicity [83].

Apart from these, there are several other plant extracts and active ingredients i.e. *Greyia flanaganii* leaf extract [84], ethanolic extracts of *Salvia cryptantha* and *Salvia cyanescens* [85], ethanolic extracts of grape seeds and peels [86], stem bark extract of *Bauhinia rufescens* [87], dried flower extract of *Inula britannica* [88], stems extract of *Liriodendron tulipifera* [89], *Cotoneaster nummularia* extract [90], Cocoa pod extract [91], *Salacia reticulata* extract [92], resveratrol and oxyresveratrol isolated from citrus fruits [93], which have been found to possess strong tyrosinase inhibitory activity as shown in various *in vivo* and *in vitro* pigment cell systems.

Recent studies of Cho et al. [94] have also demonstrated that trans-p-coumaric acid methyl ester and N-(trans-cinnamoyl) tryptamine isolated from the roots of *Oryza sativa* L., works as strong inhibitor of tyrosinase and exerts skin lightening effect on murine B16-F10 melanoma cells. While Han et al. [95] have also shown that there is considerable skin lightening activity in the aerial parts of *Pueraria thunbergiana* extract, which interrupts the maturation of tyrosinase through inhibiting α -glucosidase, consequently reducing the tyrosinase activity in the pigment cells.

Inhibition of Melanin dispersion or translocation

Several studies have focused on the identification of regulatory factors of the melanosome movement in dendrites and of the interaction between keratinocytes and melanocytes during the transfer process [96-97]. In this regard, several molecules are also known to have an effect on the transfer of melanin from melanocytes to keratinocytes, leading to lightening of the skin and can be used as therapeutic agents for hyperpigmentary skin disorders [66].

Niacinamide is a biologically active form of niacin (vitamin B3) found widely in many root vegetables and yeasts, and it is also an important precursor of NADH (nicotinamide adenine dinucleotide) and NADPH (nicotinamide adenine dinucleotide phosphate), inhibits the transfer of melanosomes from melanocytes to keratinocytes in cocultures of human melanocytes and keratinocytes. Some clinical studies using topically applied niacinamide have also demonstrated a reversible reduction in hyperpigmented lesions and increased skin lightness [98].

Recent research work done in our laboratory has demonstrated that plant extracts of *Arachis hypogaea* [24], *Citrus reticulata* [26], *Aloe vera* [25], *Chlorophytum tuberosum* [27] *Ocimum sanctum* [99], *G. glabra* and *Punica granatum* (unpublished data) significantly lighten the skin via stimulation of adrenergic receptors of various subtypes in different melanophore models of animals like fishes, amphibians and reptiles. These findings have provided a strong basis to understand the receptor based mechanism of the skin lightening at the cellular level.

S. No.	Plant Name	Bioactive compound	Experimental Assay / Cell type/ Animal model	Target site/ Mode of action	Reference
1	Angelica daburica	Isoimperatorin and Imperatorin	B16 melanoma cells	Tyrosinase m-RNA level (Tyrosinase Synthesis)	Cho et al. [66]
2	Aesculus californica	Arbutin	Human melanocytes in culture	inhibition of tyrosinase activity at post transcription level, DHICA polymerase activity	Tomita et al. [69] Chakraborty et al. [70] Hori et al. [71]
3	Camellia sinensis (Green tea)	Epicatechin gallate, Epigallocatechin gallate and Gallocatechin gallate	<i>In vitro</i> mushroom Tyrosinase Assay	Inhibition of tyrosinase activity	No et al. [72]
4	Aloe vera	Aloesine and Aloin		Inhibition of tyrosinase activity, Inhibition of pigment translocation	Picardo and Carrera, [73] Ali et al. [25]
5	Glycyrrhiza glabra	glabridin and isoliquiritigenin	B16 murine melanoma cells	Inhibition of tyrosinase activity	Nerya et al. [76] Fu et al. [77] Holloway, [78]
6	Morus alba	mulberroside F	melan-a cells	Inhibition of tyrosinase activity	Lee et al. [79,80]
7	Vitis vinifera	proanthocyanidin	<i>In vivo</i> Human Females	Inhibition of tyrosinase activity	Yamakoshi et al. [81]
8	Punica granatum	Ellagic acid	UV-irradiated guinea pigs	Inhibition of tyrosinase activity	Yoshimura et al. [82]
9	Citrus Fruits	Hesperidin	B16 melanoma cells	Inhibition of tyrosinase activity	Zhang et al. [83]
10	Greyia flanaganii	Ethanolic extract of leaves	Cultured melanocytes cells	Inhibition of tyrosinase activity	Mapunya et al. [84]
11	Salvia cryptantha	Ethanol (EtOH) extracts	<i>In vivo</i> and <i>in vitro</i> models	Tyrosinase activity by ferrous ion-chelating ability, (DPPH) and superoxide radical scavenger activity	Süntar et al. [85]
12	Salvia cyanescens	Ethanol (EtOH) extracts	Rats and mice	Inhibition of tyrosinase activity	Süntar et al. [85]
13	Bauhinia rufescens	a-amyrin acetate	<i>In vitro</i> tyrosinase inhibition assay	Inhibition of tyrosinase activity	Muhammad and Sirat [87]
14	Inula britannica	1-O-acetylbritannilactone, britannilactone	Cultured B16 melanoma cells	Tyrosinase activity, Inhibition of ERK, Akt, cAMP related signalling,	Choo et al. [88]
15	Liriodendron tulipifera	Yangambin, Methyl 4-hydroxy-2-methylbenzoate, Methyl haematommate, 2,6-Dimethoxy-p-quinone	B16F10 cell line	Inhibition of tyrosinase activity	Li et al. [89]
16	Cotoneaster nummularia	ferulic acid, chlorogenic acid, epicatechin, catechin	<i>In vitro</i> Mushroom tyrosinase assay	Inhibition of tyrosinase activity	Zengin et al. [90]
17	Theobroma Cacao (Cocoa pod) extract	flavonoids	<i>In vitro</i> Mushroom tyrosinase assay	Inhibition of tyrosinase activity	Karim et al. [91]
18	Salacia reticulate	root extract	B16 melanoma cells	Inhibition of tyrosinase activity	Suwannalert et al. [92]
19	Arachis hypogaea	Resveratrol, oxyresveratrol	tyrosinase assay	Inhibition of tyrosinase activity, Inhibition of pigment translocation	Park et al. [93] Galgut and Ali, [24]

20	Oryza sativa L	trans-p-coumaric acid methyl ester and N-(trans-cinnamoyl) tryptamine	murine B16-F10 melanoma cells	Inhibition of tyrosinase activity	Cho et al. [94]
21	Pueraria thunbergiana	Aerial part extract	B16F10 melanoma cell line	Downgrading microphthalmia-associated transcription factor, interrupting maturation of tyrosinase	Han et al. [95]
22	Helianthus annuus	Niacinamide	keratinocyte/ melanocyte coculture model and pigmented reconstructed epidermis (PREP) model	Inhibition of pigment translocation from melanocytes to keratinocytes	Hakozaki et al. [98]
23	Chlorophytum tuberosum	Aqueous extract	Fish scale melanophores	Inhibition of pigment translocation	Choudhari et al. [27]
24	Ocimum sanctum	Eugenol	B16F10 Cell line, Amphibian dorsal skin melanophores	Tyrosinase activity, Inhibition of pigment translocation	Ali et al. [99]

Table 1: Summary of scientifically validated active botanicals which can be used as skin lightening agents and their possible mechanism of action.

Control of melanin degradation and its removal

Apart for above two strategies, some other substances that increase the degradation and removal of pigment granules from the skin are also commonly used to remove excessive melanin content within the skin and used as cosmetic and therapeutic agents [66].

Chemical substances have been used as exfoliantes, such as α -hydroxy acids, free fatty acids and retinoic acid, which stimulate cell renewal facilitating the removal of melanised keratinocytes, leading to melanin pigment loss. Earlier studies of Amer and Metwalli, [100] had demonstrated that liquirtin isolated from Licorice root bud extracts, exerted potent skin lightening effects in melasma patients, by removing keratinocytes (desquamation), shortening the cell cycle and facilitating rapid pigment loss.

Conclusion and Future Perspectives

It is a well known fact that melanin is very essential pigment found in melanocytes and provides a defensive mechanism against photo damage of skin cells, but excessive melanin production and uneven distribution leads to undesirable conditions specified as hyperpigmentation. Researchers have determined the chemical changes that occur at every step of the melanogenic pathway, both in healthy individuals and in those with pigmentation disorders and huge advances have been made to understand pigment biology and the processes underlying skin pigmentation in past few decades. More basic understanding of the regulation of melanogenesis could help to develop safe and natural agents for treatment of hyperpigmentation disorders.

There are several conventional chemical treatments used for treating hyperpigmentary dysfunctions but none have been found suitable because of their adverse side effects. On the other hand natural ingredients from plant origin offer safer alternatives in comparison to other hazardous chemical skin lighteners, as well as they also offer additional functionalities as sunscreen boosters, moisturizers, or “anti-aging” ingredients, thereby supporting skin health, and reducing the appearance of wrinkles. On the light of these facts it can be concluded that the natural skin whiteners from plant extracts are more effective, safer, non-toxic and cost effective compared to the chemical skin whiteners with diverse side effects.

At the end of the discussion it is important to note that the previous knowledge and current updates in melanocyte biology and the

processes underlying melanin synthesis have made remarkable progresses over the last few years and have undoubtedly opened new paths in pharmacologic approaches for the treatment of hyperpigmentation. But at the same time, the topic has become more complex and the classification of the bio molecules is continually becoming more complicated to understand. *In vitro* pigmented skin substitutes can be produced by tissue engineering and *in vivo* models which can be exploited as useful tools for understanding these mechanisms and developing appropriated treatments or drugs. Also, as the number of putative depigmenting agents grows there is an increased need for studies to clarify product efficacy, cytotoxicity, topical skin penetration, target oriented drug delivery, stability, safety and efficacy for *in-vivo* pigment cell system.

However it is clear that great progress has been made till now in the filed of normal and uneven skin pigmentation related to its physiology and pharmacology, but it is even more apparent that there is a great deal of work is still left to be done. There are still many questions which are unanswered, concerning the precise mechanism of action and *in-vivo* efficacy of the applied natural molecules. Several other mechanisms involved in pigmentation disorders remain unknown and need to be elucidated upon in order to give affected people a better quality of life. In the coming days we expect inter merging of several disciplines to unravel the mystery of human pigmentation.

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References

1. Brenner M, Hearing VJ (2008) The protective role of melanin against UV damage in human skin. *Photochem Photobiol* 84: 539-549.
2. Ali SA, Naaz I (2015) Current Challenges in Understanding the Story of Skin Pigmentation — Bridging the Morpho-Anatomical and Functional Aspects of Mammalian Melanocytes: Muscle Cell and Tissue. INTECH.
3. Miyamura Y, Coelho SG, Wolber R, Miller SA, Wakamatsu K, et al. (2007) Regulation of human skin pigmentation and responses to ultraviolet radiation. *Pigment Cell Res* 20: 2-13.
4. Lee TH, Seo JO, Baek SH, Kim SY (2014) Inhibitory effects of resveratrol on melanin synthesis in ultraviolet B-induced pigmentation in Guinea pig skin. *Biomol Ther (Seoul)* 22: 35-40.

5. Zaidi KU, Ali AS, Ali SA, Naaz I (2014) Microbial tyrosinases: promising enzymes for pharmaceutical, food bioprocessing, and environmental industry. *Biochem Res Int* 2014: 854687.
6. Slominski A, Tobin DJ, Shibahara S, Wortsman J (2004) Melanin pigmentation in mammalian skin and its hormonal regulation. *Physiol Rev* 84: 1155-1228.
7. Bellei B, Pitisci A, Izzo E, Picardo M (2012) Inhibition of melanogenesis by the pyridinyl imidazole class of compounds: possible involvement of the Wnt/ β -catenin signaling pathway. *PLoS One* 7: e33021.
8. Shankar PR, Subish P (2007) Fair skin in South Asia: an obsession?. *Journal of Pakistan Association of Dermatologists* 17: 100-104.
9. Seiji M, Shimao K, Birbeck MS, Fitzpatrick TB (1963) Subcellular localization of melanin biosynthesis. *Ann N Y Acad Sci* 100: 497-533.
10. Wasmeier C, Hume AN, Bolasco G, Seabra MC (2008) Melanosomes at a glance. *J Cell Sci* 121: 3995-3999.
11. Raposo G, Marks MS (2007) Melanosomes--dark organelles enlighten endosomal membrane transport. *Nat Rev Mol Cell Biol* 8: 786-797.
12. Zaidi KU, Manil A, Ali AS, Ali SA (2013) Evaluation of tyrosinase producing endophytic fungi from *Calotropis gigantea*, *Azadirachta indica*, *Ocimum tenuiflorum* and *Lantana camara*. *Annual Review & Research in Biology* 3: 389-396.
13. Tian-Xiao M, Chao-Feng Z, Miyamoto T, Ishikawa H, Shimizu K et al. (2012) The Melanin Biosynthesis Stimulating Compounds Isolated from the Fruiting Bodies of *Pleurotus citrinopileatus*. *Journal of Cosmetics, Dermatological Sciences and Applications* 2: 151-157.
14. Ito S, Wakamatsu K, d'Ischia M, Napolitano A, Pezzella A (2011) In Melanins and melanosomes: biosynthesis, biogenesis, physiological, and pathological functions. Wiley-Blackwell, USA.
15. Yamaguchi Y, Hearing VJ (2009) Physiological factors that regulate skin pigmentation. *Biofactors* 35: 193-199.
16. Chen Y, Chalouni C, Tan C, Clark R, Venook R, et al. (2012) The melanosomal protein PMEL17 as a target for antibody drug conjugate therapy in melanoma. *J Biol Chem* 287: 24082-24091.
17. Hoashi T, Watabe H, Muller J, Yamaguchi Y, Vieira WD, et al. (2005) MART-1 is required for the function of the melanosomal matrix protein PMEL17/GP100 and the maturation of melanosomes. *J Biol Chem* 280: 14006-14016.
18. Vachtenheim J, Borovanský J (2010) "Transcription physiology" of pigment formation in melanocytes: central role of MITF. *Exp Dermatol* 19: 617-627.
19. Simon JD, Peles D, Wakamatsu K, Ito S (2009) Current challenges in understanding melanogenesis: Bridging chemistry, biological control, morphology, and function. *Pigment Cell Melanoma Res* 22: 563-579.
20. Hara M, Yaar M, Byers HR, Goukassian D, Fine RE, et al. (2000) Kinesin participates in melanosomal movement along melanocyte dendrites. *J Invest Dermatol* 114: 438-443.
21. Hume AN, Collinson LM, Hopkins CR, Strom M, Barral DC, et al. (2002) The leaden gene product is required with Rab27a to recruit myosin Va to melanosomes in melanocytes. *Traffic* 3: 193-202.
22. Hume AN, Wilson MS, Ushakov DS, Ferenczi MA, Seabra MC (2011) Semi-automated analysis of organelle movement and membrane content: understanding rab-motor complex transport function. *Traffic* 12: 1686-1701.
23. Ali SA, Ali AS, Gupta SS (1985) Responses of isolated fish melanophores to adrenergic agonists. *Asian Congress of Pharmacology* 17: 240-241.
24. Galgut JM, Ali SA (2011) Effect and mechanism of action of resveratrol: a novel melanolytic compound from the peanut skin of *Arachis hypogaea*. *J Recept Signal Transduct Res* 31: 374-380.
25. Ali SA, Galgut JM, Choudhary RK (2012) On the novel action of melanolysis by a leaf extract of *Aloe vera* and its active ingredient aloin, potent skin depigmenting agents. *Planta Med* 78: 767-771.
26. Galgut JM, Ali SA (2012) Hesperidin induced melanophore aggregatory responses in tadpole of *Bufo melanostictus* via α -adrenoceptors. *Pharmacologia* 3: 519-524.
27. Chaudhari SA, Peter J, Galgut JM, Ali SA (2012) Melanin inhibitory and melanin stimulatory effects of extracts of *Chlorophytum tuberosum* and *Chlorophytum borivilianum* on isolated fish scale melanophores. *African Journal of Pharmacy and Pharmacology* 6: 919-923.
28. Ali AS, Peter J, Ali SA (1995) Role of cholinergic receptors in melanophore responses of amphibians. *Acta Biol Hung* 46: 61-73.
29. Ali SA, Sultan T, Galgut JM, Sharma R, Meitei KV, et al. (2011) In vitro responses of fish melanophores to lyophilized extracts of *Psoralea corylifolia* seeds and pure psoralen. *Pharmaceutical Biology* 49: 422-427.
30. Sultan T, Ali SA (2011) *Psoralea corylifolia* extracts stimulate cholinergic-like psoralen receptors of tadpole-tail melanophores, leading to skin darkening. *Journal of Receptors and Signal Transduction* 31: 39-44.
31. Sajid M, Ali SA (2011) Mediation of cholino-piperine like receptors by extracts of *Piper nigrum* induces melanin dispersion in *Rana tigerina* tadpole melanophores. *J Recept Signal Transduct Res* 31: 286-290.
32. Ali SA, Meitei KV (2011) *Nigella sativa* seed extract and its bioactive compound thymoquinone: the new melanogens causing hyperpigmentation in the wall lizard melanophores. *J Pharm Pharmacol* 63: 741-746.
33. Ali SA, Meitei KV (2011) On the action and mechanism of withaferin-A from *Withania somnifera*, a novel and potent melanin dispersing agent in frog melanophores. *J Recept Signal Transduct Res* 31: 359-366.
34. Meitei KV, Ali SA (2012) Fig leaf extract and its bioactive compound psoralen induces skin darkening effect in reptilian melanophores via cholinergic receptor stimulation. *In Vitro Cell Dev Biol Anim* 48: 335-339.
35. Ali SA, Ali AS, Ovais M (1993) Effect of histaminergic drugs on tail melanophores of tadpole, *Bufo melanostictus*. *Indian J Exp Biol* 31: 440-442.
36. Ali SA, Peter J, Ali AS (1998) Histamine receptors in the skin melanophores of Indian bullfrog *Rana tigerina*. *Comp Biochem Physiol A Mol Integr Physiol* 121: 229-234.
37. Peter J, Meitei KV, Ali AS, Ali SA (2011) Role of histamine receptors in the pigmentary responses of the wall lizard, *Hemidactylus flaviviridis*. *Current Science* 101: 226-229.
38. Salim S, Ali AS, Ali SA (2011) Insights into the physiomodulatory role of histaminergic receptors in vertebrate skin pigmentation. *J Recept Signal Transduct Res* 31: 121-131.
39. Salim S, Ali AS, Ali SA (2013) 5-HT receptor subtypes as key targets in mediating pigment dispersion within melanophores of teleost, *Oreochromis mossambicus*. *Comparative Biochemistry and Physiology, Part B* 164: 117-123.
40. Ali SA, Salim S, Sahni T, Peter J, Ali AS (2012) 5-HT receptors as novel targets for optimizing pigmentary responses in dorsal skin melanophores of frog, *Hoplobatrachus tigerinus*. *Br J Pharmacol* 165: 1515-1525.
41. Salim S, Ali SA (2011) Vertebrate melanophores as potential model for drug discovery and development: a review. *Cell Mol Biol Lett* 16: 162-200.
42. Konrad K, Wolff K (1973) Hyperpigmentation, melanosome size, and distribution patterns of melanosomes. *Arch Dermatol* 107: 853-860.
43. Thong HY, Jee SH, Sun CC, Boissy RE (2003) The patterns of melanosome distribution in keratinocytes of human skin as one determining factor of skin colour. *Br J Dermatol* 149: 498-505.
44. Lin JY, Fisher DE (2007) Melanocyte biology and skin pigmentation. *Nature* 445: 843-850.
45. Ortonne JP, Bissett DL (2008) Latest insights into skin hyperpigmentation. *J Invest Dermatol Symp Proc* 13: 10-14.
46. Rajagopalan R, Anderson RT (1997) The profile of a patient with contact dermatitis and a suspicion of contact allergy (history, physical characteristics, and dermatology-specific quality of life). *Am J Contact Dermat* 8: 26-31.
47. Chren MM, Lasek RJ, Sahay AP, Sands LP (2001) Measurement properties of Skindex-16: a brief quality-of-life measure for patients with skin diseases. *J Cutan Med Surg* 5: 105-110.
48. Hearing VJ, Tsukamoto K (1991) Enzymatic control of pigmentation in mammals. *FASEB J* 5: 2902-2909.

49. Maeda K and Fukuda M (1991) In vitro effectiveness of several whitening cosmetic components in human melanocytes. *J. Soc. Cosmet. Chem* 42: 361-368.
50. Im S, Kim J, On WY, Kang WH (2002) Increased expression of alpha-melanocyte-stimulating hormone in the lesional skin of melasma. *Br J Dermatol* 146: 165-167.
51. Kang WH, Yoon KH, Lee ES, Kim J, Lee KB, et al. (2002) Melasma: histopathological characteristics in 56 Korean patients. *Br J Dermatol* 146: 228-237.
52. Jennifer C, Stephie CM, Abhishri SB, Shalini BU (2012) A review on skin whitening property of plant extracts. *Int J Pharm Bio Sci* 3: 332-347.
53. Plensdorf S, Martinez J (2009) Common pigmentation disorders. *Am Fam Physician* 79: 109-116.
54. Davis EC, Callender VD (2010) Postinflammatory hyperpigmentation: a review of the epidemiology, clinical features, and treatment options in skin of color. *Journal of Clinical Aesthetic Dermatology* 3: 20-31.
55. Callender VD, St Surin-Lord S, Davis EC, Maclin M (2011) Postinflammatory hyperpigmentation: etiologic and therapeutic considerations. *American Journal of Clinical Dermatology* 12: 87-99.
56. Boer A, Asgari M (2006) Prurigo pigmentosa: an underdiagnosed disease? *Indian J Dermatol Venereol Leprol* 72: 405-409.
57. Böer A, Misago N, Wolter M, Kiryu H, Wang XD, et al. (2003) Prurigo pigmentosa: a distinctive inflammatory disease of the skin. *Am J Dermatopathol* 25: 117-129.
58. Estève E, Saudeau L, Pierre F, Barruet K, Vaillant L, et al. (1994) [Physiological cutaneous signs in normal pregnancy: a study of 60 pregnant women]. *Ann Dermatol Venereol* 121: 227-231.
59. Solano F, Briganti S, Picardo M, Ghanem G (2006) Hypopigmenting agents: an updated review on biological, chemical and clinical aspects. *Pigment Cell Res* 19: 550-571.
60. Yamaguchi Y, Brenner M, Hearing VJ (2007) The regulation of skin pigmentation. *J Biol Chem* 282: 27557-27561.
61. Briganti S, Camera E, Picardo M (2003) Chemical and instrumental approaches to treat hyperpigmentation. *Pigment Cell Res* 16: 101-110.
62. Zhu W, Gao J (2008) The use of botanical extracts as topical skin-lightening agents for the improvement of skin pigmentation disorders. *J Invest Dermatol Symp Proc* 13: 20-24.
63. Parvez S, Kang M, Chung HS, Cho C, Hong MC, et al. (2006) Survey and mechanism of skin depigmenting and lightening agents. *Phytother Res* 20: 921-934.
64. Ebanks JP, Wickett RR, Boissy RE (2009) Mechanisms regulating skin pigmentation: the rise and fall of complexion coloration. *Int J Mol Sci* 10: 4066-4087.
65. Gillbro JM, Olsson MJ (2011) The melanogenesis and mechanisms of skin-lightening agents--existing and new approaches. *Int J Cosmet Sci* 33: 210-221.
66. Cho YH, Kim JH, Park SM, Lee BC, Pyo HB, et al. (2006) New cosmetic agents for skin whitening from *Angelica dahurica*. *J Cosmet Sci* 57: 11-21.
67. Lall N, Kishore N2 (2014) Are plants used for skin care in South Africa fully explored? *J Ethnopharmacol* 153: 61-84.
68. Ribeiro AS, Estanqueiro, Oliveira MB, Lobo JMS (2015) Main Benefits and Applicability of Plant Extracts in Skin Care Products. *Cosmetics* 2: 48-65.
69. Tomita K, Fukuda M, Kawasaki K (1990) Mechanism of arbutin inhibitory effect on melanogenesis and effect on the human skin with cosmetic use. *Fragrance J* 6: 72-77.
70. Chakraborty AK, Funasaka Y, Komoto M, Ichihashi M (1998) Effect of arbutin on melanogenic proteins in human melanocytes. *Pigment Cell Res* 11: 206-212.
71. Hori I, Nihei K, Kubo I (2004) Structural criteria for depigmenting mechanism of arbutin. *Phytother Res* 18: 475-479.
72. No JK, Soung DY, Kim YJ, Shim KH, Jun YS, et al. (1999) Inhibition of tyrosinase by green tea components. *Life Sci* 65: PL241-246.
73. Picardo M, Carrera M (2007) New and experimental treatments of cloasma and other hypermelanoses. *Dermatol Clin* 25: 353-362, ix.
74. Tan C, Zhu W, Lu Y (2002) Aloin, cinnamic acid and sophorcarpidine are potent inhibitors of tyrosinase. *Chin Med J (Engl)* 115: 1859-1862.
75. Nerya O, Vaya J, Musa R, Izrael S, Ben-Arie R, et al. (2003) Glabrene and isoliquiritigenin as tyrosinase inhibitors from licorice roots. *J Agric Food Chem* 51: 1201-1207.
76. Fu B, Li H, Wang X, Lee FS, Cui S (2005) Isolation and identification of flavonoids in licorice and a study of their inhibitory effects on tyrosinase. *J Agric Food Chem* 53: 7408-7414.
77. Holloway VL (2003) Ethnic cosmetic products. *Dermatol Clin* 21: 743-749.
78. Lee SH, Choi SY, Kim H, Hwang JS, Lee BG, et al. (2002) Mulberroside F isolated from the leaves of *Morus alba* inhibits melanin biosynthesis. *Biol Pharm Bull* 25: 1045-1048.
79. Katsube T, Imawaka N, Kawano Y, Yamazakib Y, Shiwakuc K, et al. (2006) Antioxidant flavonol glycosides in mulberry (*Morus alba* L) leaves isolated based on LDL antioxidant activity. *Food Chem* 97: 25-31.
80. Yamakoshi J, Sano A, Tokutake S, Saito M, Kikuchi M, et al. (2004) Oral intake of proanthocyanidin-rich extract from grape seeds improves chloasma. *Phytother Res* 18: 895-899.
81. Yoshimura M, Watanabe Y, Kasai K, Yamakoshi J, Koga T (2005) Inhibitory effect of an ellagic acid-rich pomegranate extract on tyrosinase activity and ultraviolet-induced pigmentation. *Biosci Biotechnol Biochem* 69: 2368-2373.
82. Zhang RZ, Zhu WY, Xie F (2008) Effect of hesperidin on B16 and HaCaT cell lines irradiated by Narrowband-UVB light. *J Clin Dermatol*.
83. Mapunya MB, Hussein AA, Rodriguez B, Lall N (2011) Tyrosinase activity of *Greyia flanaganii* (Bolan) constituents. *Phytomedicine* 18: 1006-1012.
84. Süntar I, Akkol EK, Senol FS, Keles H, Orhan IE (2011) Investigating wound healing, tyrosinase inhibitory and antioxidant activities of the ethanol extracts of *Salvia cryptantha* and *Salvia cyanescens* using in vivo and in vitro experimental models. *J Ethnopharmacol* 135: 71-77.
85. Hsu CK, Chou ST, Huang PJ, Mong MC, Wang CK, et al. (2012) Crude ethanol extracts from grape seeds and peels exhibit anti-tyrosinase activity. *J Cosmet Sci* 63: 225-232.
86. Muhammad A, Sirat HM (2013) Potent microbial and tyrosinase inhibitors from stem bark of *Bauhinia rufescens* (Fabaceae). *Nat Prod Commun* 8: 1435-1437.
87. Choo SJ, Ryoo IJ, Kim KC, Na M, Jang JH, et al. (2014) Hypo-pigmenting effect of sesquiterpenes from *Inula britannica* in B16 melanoma cells. *Arch Pharm Res* 37: 567-574.
88. Li WJ, Lin YC, Wu PF, Wen ZH, Liu PL, et al. (2013) Biofunctional constituents from *Liriodendron tulipifera* with antioxidants and anti-melanogenic properties. *Int J Mol Sci* 14: 1698-1712.
89. Zengin G, Uysal A, Gunes E, Aktumsek A (2014) Survey of phytochemical composition and biological effects of three extracts from a wild plant (*Cotoneaster nummularia* Fisch. et Mey.): a potential source for functional food ingredients and drug formulations. *PLoS One* 9: e113527.
90. Karim AA, Azlan A, Ismail A, Hashim P, Abd Gani SS, et al. (2014) Phenolic composition, antioxidant, anti-wrinkles and tyrosinase inhibitory activities of cocoa pod extract. *BMC Complement Altern Med* 14: 381.
91. Suwannalert P, Kariya R, Suzu I, Okada S (2014) The effects of *Salacia reticulata* on anti-cellular oxidants and melanogenesis inhibition in alpha-MSH-stimulated and UV irradiated B16 melanoma cells. *Nat Prod Commun* 9: 551-554.
92. Park J, Park JH, Suh HJ, Lee IC, Koh J, et al. (2014) Effects of resveratrol, oxyresveratrol, and their acetylated derivatives on cellular melanogenesis. *Arch Dermatol Res* 306: 475-487.
93. Cho JG, Huh J, Jeong RH, Cha BJ, Shrestha S, et al. (2015) Inhibition effect of phenyl compounds from the *Oryza sativa* roots on melanin production in murine B16-F10 melanoma cells. *Nat Prod Res* 29: 1052-1054.

-
94. Han E, Chang B, Kim D, Cho H, Kim S (2015) Melanogenesis inhibitory effect of aerial part of *Pueraria thunbergiana* in vitro and in vivo. *Arch Dermatol Res* 307: 57-72.
 95. Seiberg M (2001) Keratinocyte-melanocyte interactions during melanosome transfer. *Pigment Cell Res* 14: 236-242.
 96. Minwalla L, Zhao Y, Cornelius J, Babcock GF, Wickett RR, et al. (2001) Inhibition of melanosome transfer from melanocytes to keratinocytes by lectins and neoglycoproteins in an in vitro model system. *Pigment Cell Res* 14: 185-194.
 97. Hakozaiki T, Minwalla L, Zhuang J, Chhoa M, Matsubara A, et al. (2002) The effect of niacinamide on reducing cutaneous pigmentation and suppression of melanosome transfer. *Br J Dermatol* 147: 20-31.
 98. Ali SA, Choudhary RK, Naaz I, Khan N, Sajid M, Galgut J, Miraj M, Jakkala L, Ali AS (2015) Comparative characterization and scientific validation of certain plant extracts from their biomedical importance. *Bioscience Biotechnology Research Communications* 8: 78-86.
 99. Amer M, Metwalli M (2000) Topical liquiritin improves melasma. *Int J Dermatol* 39: 299-301.
 100. Young Kang H, Ortonne JP (2009) Melasma update. *Actas Dermosifiliogr* 100 Suppl 2: 110-113.