

## Lysophosphatidic Acid- A Target in Ovarian and Endometrial Cancer Therapy

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### Abstract

Lysophosphatidic acid (LPA), one of the simplest and most potent lysophospholipids exerting many physiological and pathological actions on various cell types, plays also an essential role in tumorigenesis and cancer metastasis. Overexpression of LPA and its receptors is a common phenomenon in metastatic carcinomas that can be used in new diagnostics strategies. Both, in the ovarian and endometrial cancer cells, LPA activates various signal transduction pathways, leading to the increased proliferation and metastatic abilities of the cells. In this review we would like to prove that development of potential treatment strategies by targeting LPA has a great promise in therapeutics.

**Keywords:** Lysophosphatidic acid; Lysophosphatidic acid receptors; Phosphatidic acid; Phospholipids

**Abbreviations** LPA: Lysophosphatidic Acid; Lpars: Lysophosphatidic Acid Receptors; PA: Phosphatidic Acid; Pls: Phospholipids; PLD: Phospholipase D; ATX: Autotoxin; PLA: Phospholipase A; Lpls: Lysophospholipids; Spla2-Secretory Phospholipase A2; PS-PLA1: Phosphatidylserine-specific Phospholipase A1; LCAT: Lecithin-Cholesterol Acyltransferase; FIGO: Federation of Gynecology and Obstetrics; VEGF: Vascular Endothelial Growth Factor; HIF-1α: Hypoxia Inducible Factor-1α; IGF2: Insulin-like Growth Factor-2; hTR: RNA Component; hTERT: Human Telomerase Reverse Transcriptase Activity; HREs: Hypoxia-responsive Elements; MMPs: Matrix Metalloproteinases

### Introduction

The present article focuses particularly on one of the simplest and most potent lysophospholipids-lysophosphatidic acid (LPA) and summarizes recent knowledge on the biological role of LPA signaling via LPA receptors (LPARs) in the pathogenesis of ovarian and endometrial cancer. We would also like to search for the evidence to present LPA as target molecule for the establishment of novel chemoprevention agents in clinical cancer approaches.

### Lysophosphatidic acid production

Lysophosphatidic acid is a simple phospholipid that consists of a phosphate, a glycerol and a fatty acid. Initially the first studies revealed LPA effects on blood pressure, uterine smooth muscle contraction and platelet aggregation [1-3]. Subsequently, different studies revealed the effects of LPA on many other physiological and pathological actions in various cell types, such as: cell proliferation and differentiation [4], cytoskeletal rearrangement [5] or cell-to-cell interactions [6]. Finally, it has been proven that LPA is implicated in the pathogenesis of various diseases in the human including carcinogenic cell invasion and tumorigenesis [7].

In the human body LPA can be found both intra and extracellular. It was detected in many various biological fluids such as serum and plasma [8,9], tears [10], ascites [11], seminal plasma [12] and follicular fluid [13]. Moreover, it can also be produced in various cell types like: endometrial cells [14], ovarian cells [14-16], mast cells [17], erythrocytes [18] and neurons [19].

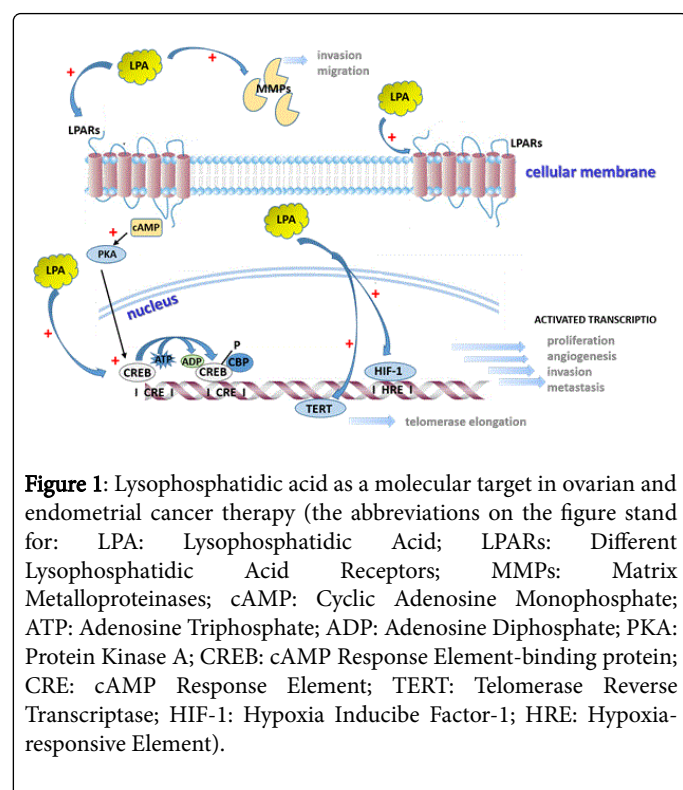
In the human body, two general pathways of LPA production have been demonstrated. In the first pathway phosphatidic acid (PA) is produced from phospholipids (PLs) by phospholipase D (PLD), also called autotoxin (ATX) or from diacylglycerol by diacylglycerol kinase and consequently there is deacylation of PA to LPA by phospholipase (PLA)-type enzymes [20]. In the second pathway, PLs are first converted to lysophospholipids (LPLs) by the action of secretory (sPLA2), PS-PLA1, and lecithin-cholesterol acyltransferase (LCAT), and then the LPL is converted to LPA by ATX [8]. The first pathway is characteristic for intracellular LPA production while the second for serum and plasma. These two ways of LPA synthesis reflect possible levels of regulation-or deregulation in the organism being especially important at such pathological status as cancer [7]. Moreover, LPA-dependent different signaling pathways have clear therapeutic repercussions since pharmaceutical drugs targeting certain enzymes differ from those targeting other LPA biosynthetic pathways [21,22].

### G protein coupled receptor-mediated LPA signaling

In mammals, LPA interacts with G protein-coupled transmembrane receptors. So far, at least six types of LPA receptors (LPAR) have been identified, such as LPAR1/EDG2, LPAR2/EDG4, LPAR3/EDG7, LPAR4/P2Y9/GPR23, LPAR5/GPR92 and LPAR6/P2Y5 as well as newly identified GPR87 [23]. These LPARs are expressed in various organs and cells [20]. LPA signaling via various LPARs leads to a variety of cellular responses such as for example cell growth, migration, differentiation, morphogenesis and protection from apoptosis [24]. In recent studies, it has been demonstrated that LPA receptors are new candidates for therapeutic targets in cancer therapy [25]. There is also much evidence in the literature describing

overexpression of one or more of LPARs in certain types of cancers. However, most cancers overexpress multiple subtypes of LPA receptors (LPARs) [26]. Specifically, LPAR1 has been shown to be a regulator of cancer cell motility and metastasis [27,28]. Ovarian cancers predominantly express LPAR2 which are likely to play an important role their aggressiveness [29]. LPAR3 is the dominant receptor subtype in some human melanomas and might be inhibitory to their growth and viability [30]. On the other hand, LPAR4, LPAR5 and LPAR6 are also expressed in the human ovary but at relatively low levels [31].

The influence of LPA on the reproductive system function of the female has been examined and described for about 30 years. Since the first reports published by Jarvis et al. [32] in women, the involvement of LPA signaling in the female reproductive organ disfunction clearly point at the possible development of the future therapeutic strategies. The present article focuses particularly on LPA as the potential (Figure 1).



## Lysophosphatidic acid as a therapeutic target in ovarian cancer

These days ovarian cancer causes more mortalities than any other gynecological cancer distinctive for female population worldwide. This type of cancer occurs mostly after menopause when the ovaries have limited physiological role therefore impaired ovarian function rarely causes any symptoms. Moreover, deep in the pelvis anatomical location of the ovaries precludes many symptoms of this neoplasm unless the tumor reaches large size or is disseminated towards other organs. Taking above into consideration, there are a lot of difficulties in detection of ovarian cancer in its early stage [33]. Due to this fact, ovarian cancer is usually diagnosed at its advanced stage when the survival rates are poor. Almost 90% of women are diagnosed with metastatic disease in the pelvis or abdomen and for these patients 5-year survival rates are lower than 30%. In contrast, in patients

diagnosed with early stage of ovarian cancer confined only to the ovaries, the 5-year survival rate exceeds even 90% [34]. The symptoms of ovarian cancer are pelvic or abdominal pain, urinary frequency or urgency, increased abdominal size or bloating and difficulty in ingestion [35,36].

Ovarian tumor stage is determined according to the International Federation of Gynecology and Obstetrics (FIGO) staging system. There are four stages of ovarian cancer. In the I stage, the tumor is confined to ovaries or fallopian tubes. In the II stage, the tumor involves one or both ovaries or fallopian tubes with pelvic extension (below pelvic brim) or primary peritoneal cancer. In the III stage, the tumor involves one or both ovaries or fallopian tubes or primary peritoneal cancer with cytologically or histologically confirmed spread to the peritoneum outside the pelvis and/or metastasis to the retroperitoneal lymph nodes. In the most advanced stage IV distant metastases excluding peritoneal metastases occur [37].

Due to the fact, that there are a lot of difficulties in detection of ovarian cancer, it is essential to develop a specific and sensitive method for its early detection and treatment. On the other hand, this type of cancer is the most thoroughly studied with respect to LPA signaling in carcinogenesis. It is known that LPA is produced by ovarian cancer cells and acts as the ovarian cancer activating factor [14,38,39]. It was found that LPA levels in the serum samples from ovarian cancer patients were much higher than in the serum samples from the group of healthy patients [40]. However, there are also reports that there was no difference in LPA levels between malignant and benign ovarian cancer tumors [33]. Increased levels of LPA were also found in ascites of ovarian cancer patients and in the corresponding plasma samples [38,41-43].

Taking into consideration the etiopathology of ovarian cancer, many studies suggested that LPA played very important role in the progression and pathogenesis of ovarian cancer [39,44-49]. In the aspect of cancer progression, Goldsmith et al. [47] documented that LPA stimulated the proliferation of ovarian cancer cells via the  $g_{12}$  proto-oncogene  $G_{12}$ , which is the most potent  $\alpha$  subunit in promoting cell proliferation and neoplastic transformation [47,50]. LPA stimulated the potent activation of CREB via the  $g_{12}$  proto-oncogene by stimulating the phosphorylation of Ser133 of cAMP response element-binding protein (CREB), leading to activation of CREB, which has been implicated in ovarian cancer cell proliferation [51]. Linnerth et al. [51] also demonstrated that LPA activated CREB very rapidly – the activation was observed 3 minutes after LPA treatment. Moreover, the phosphorylation of CREB was stimulated by the expression of the constitutively activated mutation of  $G_{12}$  even in the absence of LPA, whereas silencing  $G_{12}$  abrogated LPA-activated stimulation of CREB. Therefore, LPA-mediated activation of CREB via  $G_{12}$  was through the cAMP-independent mechanism in which Ras-ERK-dependent signalling pathway was involved [51]. Also, the expression of the dominant negative S133A mutant CREB led to the attenuation of the proliferation of ovarian cancer cells stimulated by LPA [51]. It proves that LPA- $G_{12}$  signalling axis is involved in ovarian cancer cell proliferation. There is the unique  $G_{12}$ -dependent mechanism through which LPA signalling converges on CREB to stimulate the proliferation of ovarian cancer cells [49].

There are continuous efforts in the literature to establish whether different cellular effects on cell proliferation, motility and invasion in cancer cells depend on the type of LPA receptor. Many studies documented the overexpression of LPAR2 and LPAR3 in ovarian cancer cell lines in comparison with normal ovarian epithelial cells

[29,39,52]. The elevated expression of LPAR2 and LPAR3 also stimulated the migration and invasion of ovarian cancer cells [53]. What is more, LPA promoted angiogenesis in ovarian tumors via the stimulation of vascular endothelial growth factor (VEGF) expression [54]. Goetzl et al. [29] and Hu et al. [54] demonstrated that LPA induced mRNA and protein expression of VEGF, indicating mainly LPAR2 involvement in this process. Moreover, Fujita et al. [55] documented the correlation between the LPAR2 and LPAR3 expression levels and the induction of VEGF expression in ovarian cancer cells. Moreover, the study of Yu et al. [53] proved that the knockdown of LPAR2 and LPAR3 led to the suppression of the production of VEGF in ovarian cancer cells. Lee et al. [56] reported that LPA induced VEGF expression through the activation of Hypoxia Inducible Factor-1 $\alpha$  (HIF-1 $\alpha$ ), which is known to play the central role in tumor progression and angiogenesis [57-59]. Through binding to the hypoxia responsive elements within the target gene, HIF-1 activates transcription of various hypoxia-inducible genes, such as angiogenic-VEGF or proliferation/survival factors-insulin-like growth factor-2 (IGF2) [60]. Therefore LPA-induced HIF-1 $\alpha$  activation was probably regulated by translational regulation, not by protein stabilisation [56].

Other factor playing the crucial role in tumorigenesis is telomerase. Telomerase is an RNA-dependent DNA polymerase, synthesizing telomeric DNA [61]. It contains two components: the RNA component (hTR) that is the template for telomeric DNA synthesis and the catalytic protein with the human telomerase reverse transcriptase activity (hTERT) that is responsible for the addition of the telomeric repeats onto the end of chromosome [62,63]. It has been well documented that the expression of telomerase is required for the oncogenic transformation of many normal cell types [64,65]. In the case of the ovary, telomerase is absent in normal ovarian surface epithelium and premalignant lesions, but up-regulated in 95% of ovarian carcinomas [66]. Yang et al. [45] in the in vitro studies found that all LPARs were expressed in various ovarian cancer cell lines. These authors also found that LPA up-regulated hTERT mRNA in ovarian cancer cells through the activation of HIF-1 $\alpha$  [45]. Moreover, the mutation of one or two hypoxia-responsive elements (HREs) in the 342 promoter region abolished the induction of hTERT promoter activity by LPA [45]. Taking above facts into consideration, the authors concluded that HIF-1 $\alpha$  was required for the transcription activity of LPA on hTERT [45]. The results of the above findings demonstrated that the expression and activity of telomerase in ovarian cancer cells was regulated by LPA, which in turn suggests that telomerase is an important molecule through which LPA exerts its oncogenic effects pointing at LPA as the potential molecular target in the anti-cancer treatment strategies.

In invasion and metastasis of tumor cells matrix metalloproteinases (MMPs) play the same as important role as telomerase. They are proteolytic enzymes which induce extracellular matrix degradation. Especially MMP-2 and MMP-9 have been demonstrated to contribute to the progression of cancer cells [67,68]. Fishman et al. [69] demonstrated that in ovarian cancer cells LPA promoted cell migration and invasion through the activation of MMP-2, while Jeong et al. [70] proved that this active lysophospholipid stimulated cell invasion through the Ras/Rho/ROCK signalling and subsequent MMP-9 production. Moreover, it was found that in ovarian cancer MMP-7 secretion and activation was regulated by LPA [71].

There are many pathways of the action of LPA which show the significant role of LPA in ovarian tumorigenesis. On one hand, in the therapeutic strategies, elevated LPA levels in the serum samples [33]

might be exploited as the potential biomarkers of ovarian cancer, even in its early stages. On the other hand, the interactions between LPA signalling and gep proto-oncogene G $\alpha_{12}$  and VEGF expressions or MMP and telomerase activities give the possibility for LPA to become the target molecule for novel chemoprevention agent in clinical cancer approaches.

### **Lysophosphatidic acid as a therapeutic target in endometrial cancer**

Endometrial cancer is a major cause of morbidity and mortality for women worldwide. According to epidemiological data of the Polish National Cancer Registry 2009 it was the fourth most common malignancy among women in Poland, causing 7.3% cases with the incidence ratio of 15/100 000. The mortality ratio was 2.4/100 000, which led to the twelfth place in terms of the causes of cancer deaths in Poland [72]. Most women with endometrial cancer are diagnosed at an early stage with uterine-confined tumors, often after generally known characteristic symptoms like atypical menstrual periods, abnormal vaginal or uterine bleeding [73]. Despite the overall favourable prognosis of endometrial cancer, some women have neoplasms with more aggressive histology, and are at substantial risk of recurrence and death. The main prognostic factors include age, race, stage, grade, depth of invasion, tumor size and cell type [74]. The epidemiology of endometrial cancer is multifactorial. The most important risk factors for endometrial cancer associated with the development of endometrial carcinoma are unopposed estrogen exposure and obesity [75]. Endometrial cancers have been broadly classified into two types [76]. Type I neoplasms, including endometrioid adenocarcinomas, are most common, generally arise from atypical endometrial hyperplasia and are estrogen dependent. Type II cancers include more aggressive histological variants such as clear-cell and serous carcinomas and uterine carcinosarcomas. Non-endometrioid tumors are less common than endometrioid tumors but are associated with disproportionately high mortality. Routinely, endometrial cancer is successfully treated with surgery and/or radiotherapy [77]. However, there is always a group of patients with an advanced or recurrent disease, or those who wish to preserve their fertility. Therefore, there is still an increasing demand for introducing more effective, targeted, and less morbid therapies. Moreover the research of the predictive factors of recurrence or death is at least the same as important.

As the potential candidates for targeted anti-cancer therapy, like in the case of ovarian cancer, are also considered the members of LPA family. While the connection between LPA and tumorigenesis in ovaries is fairly well understood, the connection between LPA and endometrial cancer is not well examined. Most of the studies were performed in vitro using the endometrial carcinoma cell line HEC1A. Hope et al. [78] reported that among the four principle LPA receptors (LPAR1, LPAR2, LPAR3, LPAR4), LPAR2 was predominantly expressed by HEC1A cells. It was also documented that the physiological level of LPA stimulated the invasion and proliferation of HEC1A cells [78,79]. Moreover, Wang et al. [79] pointed at LPA as the strong promoter of urokinase plasminogen activator, which elevated levels were correlated with tumor malignancy. What is more, the knockdown of LPAR2 caused the suppression of LPA-induced HEC1A invasion, but there was no significant changes in the level of migration of HEC1A cells [79]. Besides, the knockdown of LPAR2 blocked LPA-induced activation of MMP-7 which plays important regulatory role in cell surface proteolysis and is capable of binding to a variety of cell surface proteins, such as E-cadherin,  $\beta$ -integrin and tumor necrosis



factor- $\alpha$  [79]. In endometrial cancer, like in the ovarian cancer, the overexpression of MMP-7 initiates the activation of MMP-2 which promotes cancer invasion [78]. All of the above data suggest the possibility of LPA-dependent targeted molecular therapy also in endometrial cancer.

## Conclusion

In this review, we provided updated evidence for LPA signaling via LPARs in ovarian and endometrial cancer cells. Moreover, we had an intention to present LPA as a promising molecular target in the diagnosis and therapy of ovarian and endometrial cancer.

So far, in most reproductive organ associated malignancies, targeted therapies have not entered clinical practice. Despite important advances of clinical trials in the therapy of cancer over the past few decades, treatment failure and mortality rates in the majority of malignant diseases, remain unacceptably high. We suspect that cure rates will not improve significantly unless alternative treatments to conventional chemotherapy regimens are developed. At present, there is no cancer treatment that is based on the inhibition of any of the enzymes responsible for LPA synthesis, LPARs, or signaling downstream of these receptors. Therefore, we suppose that improving our knowledge of the physiological and pathological consequences of LPA signaling may lead to the development of therapeutic agents that will enable us to target this signaling cascade. In particular, we predict that such treatments could be used together with immunotherapy that stimulates host's immune response and with other traditional treatments to achieve better clinical prognosis of ovarian and endometrial cancer patients in the near future.

## References

1. Tokumura A, Fukuzawa K, Tsukatani H (1978) Effects of synthetic and natural lysophosphatidic acids on the arterial blood pressure of different animal species. *Lipids* 13: 572-574.
2. Tokumura A, Fukuzawa K, Yamada S, Tsukatani H (1980) Stimulatory effect of lysophosphatidic acids on uterine smooth muscles of non-pregnant rats. *Arch Int Pharmacodyn Ther* 245: 74-83.
3. Schumacher KA, Classen HG, Späth M (1979) Platelet aggregation evoked in vitro and in vivo by phosphatidic acids and lysoderivatives: identity with substances in aged serum (DAS). *Thromb Haemost* 42: 631-640.
4. Pustilnik TB, Estrella V, Wiener JR, Mao M, Eder A, et al. (1999) Lysophosphatidic acid induces urokinase secretion by ovarian cancer cells. *Clin Cancer Res* 5: 3704-3710.
5. Moolenaar WH (1995) Lysophosphatidic acid, a multifunctional phospholipid messenger. *J Biol Chem* 270: 12949-12952.
6. Fukushima N, Weiner JA, Contos JA, Contos JJ, Rehen SK, et al. (2002) Lysophosphatidic acid influences the morphology and motility of young, postmitotic cortical neurons. *Mol Cell Neurosci* 20: 271-282.
7. Kim KS, Sengupta S, Berk M, Kwak YG, Escobar PF, et al. (2006) Hypoxia enhances lysophosphatidic acid responsiveness in ovarian cancer cells and lysophosphatidic acid induces ovarian tumor metastasis in vivo. *Cancer Res* 66: 7983-7990.
8. Aoki J, Taira A, Takanezawa Y, Kishi Y, Hama K, et al. (2002) Serum lysophosphatidic acid is produced through diverse phospholipase pathways. *J Biol Chem* 277: 48737-48744.
9. Sano T, Baker D, Virag T, Wada A, Yatomi Y, et al. (2002) Multiple mechanisms linked to platelet activation result in lysophosphatidic acid and sphingosine 1-phosphate generation in blood. *J Biol Chem* 277: 21197-21206.
10. Liliom K, Guan Z, Tseng JL, Desiderio DM, Tigyi G, et al. (1998) Growth factor-like phospholipids generated after corneal injury. *Am J Physiol* 274: 1065-1074.
11. Tokumura A, Kume T, Fukuzawa K, Tahara M, Tasaka K, et al. (2007) Peritoneal fluids from patients with certain gynecologic tumor contain elevated levels of bioactive lysophospholipase D activity. *Life Sci* 80: 1641-1649.
12. Hama K, Bandoh K, Kakehi Y, Aoki J, Arai H, et al. (2002) Lysophosphatidic acid (LPA) receptors are activated differentially by biological fluids: possible role of LPA-binding proteins in activation of LPA receptors. *FEBS Lett* 523: 187-192.
13. Tokumura A, Miyake M, Nishioka Y, Yamano S, Aono T, et al. (1999) Production of lysophosphatidic acids by lysophospholipase D in human follicular fluids of In vitro fertilization patients. *Biol Reprod* 61: 195-199.
14. Shen Z, Belinson J, Morton RE, Xu Y, Xu Y (1998) Phorbol 12-myristate 13-acetate stimulates lysophosphatidic acid secretion from ovarian and cervical cancer cells but not from breast or leukemia cells. *Gynecol Oncol* 71: 364-368.
15. Eder AM, Sasagawa T, Mao M, Aoki J, Mills GB (2000) Constitutive and lysophosphatidic acid (LPA)-induced LPA production: role of phospholipase D and phospholipase A2. *Clin Cancer Res* 6: 2482-2491.
16. Luquain C, Singh A, Wang L, Natarajan V, Morris AJ (2003) Role of phospholipase D in agonist-stimulated lysophosphatidic acid synthesis by ovarian cancer cells. *J Lipid Res* 44: 1963-1975.
17. Mori K, Kitayama J, Aoki J, Kishi Y, Shida D, et al. (2007) Submucosal connective tissue-type mast cells contribute to the production of lysophosphatidic acid (LPA) in the gastrointestinal tract through the secretion of autotaxin (ATX)/lysophospholipase D (lysoPLD). *Virchows Arch* 451: 47-56.
18. Fourcade O, Simon ME, Viodé C, Rugani N, Lebalte F, et al. (1995) Secretory phospholipase A2 generates the novel lipid mediator lysophosphatidic acid in membrane microvesicles shed from activated cells. *Cell* 80: 919-927.
19. Fukushima N, Weiner JA, Chun J (2000) Lysophosphatidic acid (LPA) is a novel extracellular regulator of cortical neuroblast morphology. *Dev Biol* 228: 6-18.
20. Okudaira S, Yukiura H, Aoki J (2010) Biological roles of lysophosphatidic acid signaling through its production by autotaxin. *Biochimie* 92: 698-706.
21. Willier S, Butt E, Grunewald TG (2013) Lysophosphatidic acid (LPA) signaling in cell migration and cancer invasion: a focused review and analysis of LPA receptor gene expression on the basis of more than 1700 cancer microarrays. *Biol Cell* 105: 317-333.
22. Barbayanni E, Magrioti V, Moutevelis-Minakakis P, Kokotos G (2013) Autotaxin inhibitors: a patent review. *Expert Opin Ther Pat* 23: 1123-1132.
23. Tabata K, Baba K, Shiraishi A, Fujita N (2007) The orphan GPCR GPR87 was orphanized and shown to be a lysophosphatidic acid receptor. *Biochem Biophys Res Commun* 363: 861-866.
24. Ye X, Chun J (2010) Lysophosphatidic acid (LPA) signaling in vertebrate reproduction. *Trends Endocrinol Metab* 21: 17-24.
25. Lin ME, Herr DR, Chun J (2010) Lysophosphatidic acid (LPA) receptors: signaling properties and disease relevance. *Prostaglandins Other Lipid Mediat* 91: 130-138.
26. Muller R, Berliner C, Leptin J, Pörtner D, Bialecki W, et al. (2010) Expression of sphingosine-1-phosphatereceptors and lysophosphatidic acidreceptors on cultured and xenografted human colon, breast, melanoma, and lung tumor cells. *Tumour Biol* 31: 341-349.
27. Boucharaba A, Serre CM, Guglielmi J, Bordet JC, Clézardin P, et al. (2006) The type 1 lysophosphatidic acid receptor is a target for therapy in bone metastases. *Proc Natl Acad Sci U S A* 103: 9643-9648.
28. Shida D, Fang X, Kordula T, Takabe K, Lépine S, et al. (2008) Cross-talk between LPA1 and epidermal growth factor receptors mediates up-regulation of sphingosinekinase 1 to promote gastric cancer cell motility and invasion. *Cancer Res* 68: 6569-6577.

29. Goetzl EJ, Kong Y, Mei B (1999) Lysophosphatidic acid and sphingosine 1-phosphate protection of T cells from apoptosis in association with suppression of Bax. *J Immunol* 162: 2049-2056.
30. Altman MK, Gopal V, Jia W, Yu S, Hall H, et al. (2010) Targeting melanoma growth and viability reveals dualistic functionality of the phosphonothionate analogue of carba cyclic phosphatidic acid. *Mol Cancer* 9: 140.
31. Noguchi K, Ishii S, Shimizu T (2003) Identification of p2y9/GPR23 as a novel G protein-coupled receptor for lysophosphatidic acid, structurally distant from the Edg family. *J Biol Chem* 278: 25600-25606.
32. Jarvis AA, Cain C, Dennis EA (1984) Purification and characterization of a lysophospholipase from human amnionic membranes. *J Biol Chem* 259: 15188-15195.
33. Meleh M, Pozlep B, Mlakar A, Vrtovec HM, Kralj LZ (2007) Determination of serum lysophosphatidic acid as a potential biomarker for ovarian cancer. *J Chromatogr B Analyt Technol Biomed Life Sci* 858: 287-291.
34. Menon U (2004) Ovarian cancer screening. *CMAJ* 171: 323-324.
35. Goff BA, Mandel LS, Drescher CW, Urban N, Gough S, et al. (2007) Development of an ovarian cancer symptom index: possibilities for earlier detection. *Cancer* 109: 221-227.
36. Kisielewski R, Tołwińska A, Mazurek A, Laudanski P (2013) Inflammation and ovarian cancer--current views. *Ginekolog Pol* 84: 293-297.
37. Mutch DG, Prat J (2014) 2014 FIGO staging for ovarian, fallopian tube and peritoneal cancer. *Gynecol Oncol* 133: 401-404.
38. Xu Y, Shen Z, Wiper DW, Wu M, Morton RE, et al. (1998) Lysophosphatidic acid as a potential biomarker for ovarian and other gynecologic cancers. *JAMA* 280: 719-723.
39. Fang X, Gaudette D, Furui T, Mao M, Estrella V, et al. (2000) Lysophospholipid growth factors in the initiation, progression, metastases, and management of ovarian cancer. *Ann N Y Acad Sci* 905: 188-208.
40. Choi JW, Herr DR, Noguchi K, Yung YC, Lee CW, et al. (2010) LPA receptors: subtypes and biological actions. *Annu Rev Pharmacol Toxicol* 50: 157-186.
41. Xiao Y, Chen Y, Kennedy AW, Belinson J, Xu Y (2000) Evaluation of plasma lysophospholipids for diagnostic significance using electrospray ionization mass spectrometry (ESI-MS) analyses. *Ann N Y Acad Sci* 905: 242-259.
42. Xiao YJ, Schwartz B, Washington M, Kennedy A, Webster K, et al. (2001) Electrospray ionization mass spectrometry analysis of lysophospholipids in human ascitic fluids: comparison of the lysophospholipid contents in malignant vs nonmalignant ascitic fluids. *Anal Biochem* 290: 302-313.
43. Yoon HR, Kim H, Cho SH (2003) Quantitative analysis of acyl-lysophosphatidic acid in plasma using negative ionization tandem mass spectrometry. *J Chromatogr B Analyt Technol Biomed Life Sci* 788: 85-92.
44. Mills GB, Moolenaar WH (2003) The emerging role of lysophosphatidic acid in cancer. *Nat Rev Cancer* 3: 582-591.
45. Yang K, Zheng D, Deng X, Bai L, Xu Y, et al. (2008) Lysophosphatidic acid activates telomerase in ovarian cancer cells through hypoxia-inducible factor-1alpha and the PI3K pathway. *J Cell Biochem* 105: 1194-1201.
46. Murph MM, Liu W, Yu S, Lu Y, Hall H, et al. (2009) Lysophosphatidic acid-induced transcriptional profile represents serous epithelial ovarian carcinoma and worsened prognosis. *PLoS One* 4: 5583.
47. Goldsmith ZG, Ha JH, Jayaraman M, Dhanasekaran DN (2011) Lysophosphatidic Acid Stimulates the Proliferation of Ovarian Cancer Cells via the gep Proto-Oncogene  $Gl\pm 12$ . *Genes Cancer* 2: 563-575.
48. Ward JD, Dhanasekaran DN (2012) LPA Stimulates the Phosphorylation of p130Cas via  $Gl\pm 12$  in Ovarian Cancer Cells. *Genes Cancer* 3: 578-591.
49. Ha JH, Ward JD, Varadarajulu L, Kim SG, Dhanasekaran DN (2014) The gep proto-oncogene  $Gl\pm 12$  mediates LPA-stimulated activation of CREB in ovarian cancer cells. *Cell Signal* 26: 122-132.
50. Radhika V, Hee Ha J, Jayaraman M, Tsim ST, Dhanasekaran N (2005) Mitogenic signaling by lysophosphatidic acid (LPA) involves Galpha12. *Oncogene* 24: 4597-4603.
51. Linnerth NM, Greenaway JB, Petrik JJ, Moorehead RA (2008) cAMP response element-binding protein is expressed at high levels in human ovarian adenocarcinoma and regulates ovarian tumor cell proliferation. *Int J Gynecol Cancer* 18: 1248-1257.
52. Furui T, LaPushin R, Mao M, Khan H, Watt SR, et al. (1999) Overexpression of edg-2/vzg-1 induces apoptosis and anoikis in ovarian cancer cells in a lysophosphatidic acid-independent manner. *Clin Cancer Res* 5: 4308-4318.
53. Yu S, Murph MM, Lu Y, Khan H, Watt SR, et al. (2008) Lysophosphatidic acid receptors determine tumorigenicity and aggressiveness of ovarian cancer cells. *J Natl Cancer Inst* 100: 1630-1642.
54. Hu YL, Tee MK, Goetzl EJ, Auersperg N, Mills GB, et al. (2001) Lysophosphatidic acid induction of vascular endothelial growth factor expression in human ovarian cancer cells. *J Natl Cancer Inst* 93: 762-768.
55. Fujita T, Miyamoto S, Onoyama I, Sonoda K, Mekada E, et al. (2003) Expression of lysophosphatidic acid receptors and vascular endothelial growth factor mediating lysophosphatidic acid in the development of human ovarian cancer. *Cancer Lett* 192: 161-169.
56. Lee J, Park SY, Lee EK, Park CG, Chung HC, et al. (2006) Activation of hypoxia-inducible factor-1alpha is necessary for lysophosphatidic acid-induced vascular endothelial growth factor expression. *Clin Cancer Res* 12: 6351-6358.
57. Semenza GL, Agani F, Iyer N, Kotch L, Laughner E, et al. (1999) Regulation of cardiovascular development and physiology by hypoxia-inducible factor 1. *Ann N Y Acad Sci* 874: 262-268.
58. Semenza GL, Agani F, Feldser D, Iyer N, Kotch L, et al. (2000) Hypoxia, HIF-1, and the pathophysiology of common human diseases. *Adv Exp Med Biol* 475: 123-130.
59. Semenza GL (2003) Angiogenesis in ischemic and neoplastic disorders. *Annu Rev Med* 54: 17-28.
60. Lai YM, Wang HS, Lee CL, Lee JD, Huang HY, et al. (1996) Insulin-like growth factor-binding proteins produced by Vero cells, human oviductal cells and human endometrial cells, and the role of insulin-like growth factor-binding protein-3 in mouse embryo co-culture systems. *Hum Reprod* 11: 1281-1286.
61. Wright WE, Shay JW (2005) Telomere-binding factors and general DNA repair. *Nat Genet* 37: 116-118.
62. Collins K (2006) The biogenesis and regulation of telomerase holoenzymes. *Nat Rev Mol Cell Biol* 7: 484-494.
63. Stewart SA, Weinberg RA (2006) Telomeres: cancer to human aging. *Annu Rev Cell Dev Biol* 22: 531-557.
64. Zongaro S, de Stanchina E, Colombo T, D'Incalci M, Giulotto E, et al. (2005) Stepwise neoplastic transformation of a telomerase immortalized fibroblast cell line. *Cancer Res* 65: 11411-11418.
65. Mizumoto Y, Kyo S, Ohno S, Hashimoto M, Nakamura M, et al. (2006) Creation of tumorigenic human endometrial epithelial cells with intact chromosomes by introducing defined genetic elements. *Oncogene* 25: 5673-5682.
66. Shay JW, Bacchetti S (1997) A survey of telomerase activity in human cancer. *Eur J Cancer* 33: 787-791.
67. Min KW, Kim DH, Do SI, Kim K, Lee HJ, et al. (2010) Expression patterns of stromal MMP-2 and tumoural MMP-2 and -9 are significant prognostic factors in invasive ductal carcinoma of the breast. *APMIS* 122: 1196-1206.
68. Park SY, Jeong KJ, Panupinthu N, Yu S, Lee J, et al. (2011) Lysophosphatidic acid augments human hepatocellular carcinoma cell invasion through LPA1 receptor and MMP-9 expression. *Oncogene* 30: 1351-1359.
69. Fishman DA, Liu Y, Ellerbroek SM, Stack MS, et al. (2001) Lysophosphatidic acid promotes matrix metalloproteinase (MMP) activation and MMP-dependent invasion in ovarian cancer cells. *Cancer Res* 61: 3194-3199.

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70. Jeong KJ, Park SY, Cho KH, Sohn JS, Lee J, et al. (2012) The Rho/ROCK pathway for lysophosphatidic acid-induced proteolytic enzyme expression and ovarian cancer cell invasion. *Oncogene* 31: 4279-4289.
  71. Wang FQ, Smicun Y, Calluzzo N, Fishman DA (2006) Inhibition of matrilysin expression by antisense or RNA interference decreases lysophosphatidic acid-induced epithelial ovarian cancer invasion. *Mol Cancer Res* 4: 831-841.
  72. Didkowska J, Wojciechowska U, Tarkowski W, Zatonski W (2011) Cancer in Poland in 2009. Polish National Cancer Registry. Department of Epidemiology and Cancer Prevention. The Maria Skłodowska-Curie Memorial Cancer Center. Warsaw.
  73. Lotocki RJ, Copeland LJ, DePetrillo AD, Muirhead W, et al. (1983) Stage I endometrial adenocarcinoma: treatment results in 835 patients. *Am J Obstet Gynecol* 146: 141-145.
  74. Wright JD, Neugut AI, Wilde ET, Buono DL, Tsai WY, et al. (2012) Use and benefits of laparoscopic hysterectomy for stage I endometrial cancer among medicare beneficiaries. *J Oncol Pract* 8: 89-99.
  75. Schouten LJ, Goldbohm RA, van den Brandt PA (2004) Anthropometry, physical activity, and endometrial cancer risk: results from the Netherlands Cohort Study. *J Natl Cancer Inst* 96: 1635-1638.
  76. Bokhman JV (1983) Two pathogenetic types of endometrial carcinoma. *Gynecol Oncol* 15: 10-17.
  77. Bakkum-Gamez JN, Gonzalez-Bosquet J, Laack NN, Mariani A, Dowdy SC (2008) Current issues in the management of endometrial cancer. *Mayo Clin Proc* 83: 97-112.
  78. Hope JM, Wang FQ, Whyte JS, Ariztia EV, Abdalla W, et al. (2009) LPA receptor 2 mediates LPA-induced endometrial cancer invasion. *Gynecol Oncol* 112: 215-223.
  79. Wang C, Michener CM, Belinson JL, Vaziri S, Ganapathi R, et al. (2010) Role of the 18:1 lysophosphatidic acid-ovarian cancer immunoreactive antigen domain containing 1 (OCIAD1)-integrin axis in generating late-stage ovarian cancer. *Mol Cancer Ther* 9: 1709-1718.