

Lipid Nanoparticles: Advancing Gene Editing and mRNA Therapies

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Introduction

Lipid nanoparticles (LNPs) have emerged as critical delivery vehicles for mRNA-based therapeutics and CRISPR gene-editing systems, fundamentally altering the landscape of modern medicine [1]. Their inherent biocompatibility, coupled with their capacity to shield fragile genetic cargo from degradation, makes them exceptionally suited for these advanced biotechnologies [1]. Significant research efforts are actively dedicated to refining LNP formulations with the goals of enhancing targeted delivery, minimizing immunogenic responses, and optimizing the encapsulation of therapeutic payloads, thereby paving the way for revolutionary vaccines, innovative cancer therapies, and effective treatments for genetic disorders [1]. The development of novel ionizable lipids is a cornerstone in advancing LNP performance for mRNA delivery, primarily by facilitating endosomal escape, a critical step enabling the release of mRNA into the cell's cytoplasm [2]. Current research endeavors are focused on synthesizing lipids with precisely controlled pKa values and charge densities to meticulously tune their behavior within the cellular milieu, with the ultimate aim of achieving superior transfection efficiencies and diminished toxicity profiles [2]. When delivered via LNPs, CRISPR-Cas9 gene editing represents a potent modality for addressing a spectrum of genetic diseases [3]. A primary hurdle in this application lies in the efficient encapsulation of the substantial CRISPR-Cas9 ribonucleoprotein (RNP) complexes within LNPs and their precise redirection to target cells [3]. Consequently, research is actively investigating diverse LNP compositions and advanced manufacturing methodologies, such as microfluidics, to guarantee efficacious RNP delivery while concurrently mitigating off-target effects [3]. The immunogenicity of LNPs themselves constitutes a significant consideration, particularly for therapeutic regimens requiring repeated administration, such as mRNA vaccines or gene therapies [4]. Strategies to attenuate immune system reactions encompass modifications to lipid components, the integration of PEGylated lipids, or the design of biodegradable LNP architectures [4]. A comprehensive understanding of the intricate interactions between LNPs and the immune system is indispensable for the creation of safer and more efficacious delivery platforms [4]. Microfluidic technology has profoundly transformed the manufacturing of LNPs, bestowing precise command over particle size distribution, polydispersity, and the efficiency of cargo encapsulation [5]. This scalable approach facilitates the rapid generation of high-quality LNPs essential for clinical applications, with continuous flow microfluidics enabling reproducible synthesis, a critical attribute for therapeutic consistency [5]. Targeting strategies for LNPs are paramount for directing mRNA or CRISPR cargo to specific tissues or cellular populations, thereby augmenting therapeutic effectiveness and curtailing off-target effects [6]. This is achieved through the conjugation of targeting ligands, including antibodies or peptides, onto the LNP surface, with ongoing research exploring active targeting mechanisms to boost cellular uptake within diseased tissues [6].

The stability of mRNA and CRISPR components encapsulated within LNPs during both storage and administration is of utmost importance [7]. LNPs serve as protective barriers, shielding these sensitive nucleic acids from degradation by RNases and nucleases [7]. Efforts to optimize LNP formulations for prolonged storage at ambient temperatures are a key research focus for enhancing the accessibility of vaccines globally [7]. Beyond their application in mRNA and CRISPR delivery, LNPs are increasingly being investigated for the transport of other nucleic acid-based therapeutics, including small interfering RNAs (siRNAs) and microRNAs (miRNAs) [8]. The distinct physicochemical characteristics of LNPs facilitate the efficient encapsulation and delivery of these varied RNA species for a wide array of therapeutic objectives [8]. The in vivo design of LNPs for gene editing necessitates meticulous attention to lipid composition to ensure effective delivery of CRISPR-Cas9 components to target cells while concurrently minimizing unwanted off-target effects and immune system reactions [9]. Recent advancements in understanding the structure-activity relationships of ionizable lipids are critically important for the development of safe and effective gene-editing therapies [9]. The manufacturing of LNPs on a large scale presents considerable challenges in maintaining consistent quality and reproducibility [10]. Advanced manufacturing techniques, encompassing continuous flow processes and automation, are being actively developed to address the escalating demand for LNP-based therapeutics [10]. Ensuring reproducible particle formation and consistent payload encapsulation remains of paramount importance for successful clinical translation [10].

Description

Lipid nanoparticles (LNPs) have revolutionized the delivery of nucleic acid-based therapeutics, becoming indispensable tools for mRNA-based treatments and CRISPR gene-editing systems [1]. Their intrinsic biocompatibility and the protective barrier they offer to fragile genetic materials are key factors in their suitability for these sophisticated biotechnologies [1]. Ongoing research is focused on optimizing LNP formulations to improve targeting specificity, reduce immune system reactions, and enhance the efficient encapsulation of therapeutic payloads, thereby driving the development of next-generation vaccines, cancer therapies, and treatments for genetic disorders [1]. A crucial aspect of advancing mRNA delivery using LNPs involves the development of novel ionizable lipids [2]. These specialized lipids play a vital role in facilitating endosomal escape, a critical intracellular event that allows mRNA to access the cytoplasm [2]. Current research is actively engaged in synthesizing lipids with tailored pKa values and charge densities to precisely modulate their behavior within the cellular environment, aiming to achieve higher transfection efficiency and a reduction in cellular toxicity [2]. For gene editing applications, LNPs are pivotal in delivering CRISPR-Cas9 systems for the treatment of genetic diseases [3]. A significant challenge lies in effectively

encapsulating the large CRISPR-Cas9 ribonucleoprotein (RNP) complexes within LNPs and ensuring their directed delivery to the intended target cells [3]. This has spurred investigations into various LNP compositions and advanced manufacturing methods, such as microfluidics, to achieve efficient RNP delivery while minimizing off-target genetic modifications [3]. The potential for LNPs to elicit an immune response is a critical factor, particularly for therapeutic strategies involving repeated administration, like mRNA vaccines or gene therapies [4]. Consequently, research is exploring various approaches to mitigate these immune reactions, including modifying lipid components, incorporating PEGylated lipids, or designing biodegradable LNP structures [4]. A deep understanding of the interactions between LNPs and the immune system is essential for designing safer and more effective delivery platforms [4]. The adoption of microfluidic technology has significantly enhanced LNP manufacturing processes, providing precise control over particle size, uniformity, and the efficiency of cargo encapsulation [5]. This scalable approach enables the rapid production of high-quality LNPs that are necessary for clinical applications, with continuous flow microfluidics ensuring reproducible synthesis, which is vital for therapeutic consistency [5]. To maximize therapeutic efficacy and minimize unintended consequences, LNPs are being engineered with specific targeting strategies to direct mRNA or CRISPR cargo to designated tissues or cell types [6]. These strategies often involve conjugating targeting ligands, such as antibodies or peptides, to the surface of LNPs, and research is actively pursuing active targeting mechanisms to improve cellular uptake in diseased tissues [6]. The stability of mRNA and CRISPR components within LNPs throughout storage and administration is a critical parameter for therapeutic success [7]. LNPs provide essential protection to these fragile nucleic acids against degradation by endogenous RNases and nucleases [7]. Research efforts are particularly focused on optimizing LNP formulations to enable long-term storage at ambient temperatures, a key factor in improving the global accessibility of vaccines [7]. The utility of LNPs extends beyond mRNA and CRISPR delivery, with active exploration for the delivery of other nucleic acid-based therapeutics, including small interfering RNAs (siRNAs) and microRNAs (miRNAs) [8]. The unique physicochemical properties of LNPs are well-suited for the efficient encapsulation and delivery of these diverse RNA molecules for a broad spectrum of therapeutic purposes [8]. The design of LNPs for in vivo gene editing requires careful consideration of their lipid composition to ensure efficient delivery of CRISPR-Cas9 components to the target cells, while simultaneously minimizing off-target effects and immune responses [9]. Recent progress in deciphering the structure-activity relationships of ionizable lipids is crucial for the development of safe and effective gene-editing therapies [9]. The scaling up of LNP manufacturing presents significant challenges in maintaining consistent quality and batch-to-batch reproducibility [10]. Advanced manufacturing techniques, including continuous flow processing and automation, are being developed to meet the increasing global demand for LNP-based therapeutics [10]. Ensuring reproducible particle formation and reliable payload encapsulation is paramount for the successful clinical translation of these innovative therapies [10].

Conclusion

Lipid nanoparticles (LNPs) are crucial delivery systems for mRNA therapeutics and CRISPR gene editing, offering biocompatibility and cargo protection. Research focuses on optimizing LNP formulations for better targeting, reduced immunogenicity, and improved encapsulation to advance vaccines, cancer therapies, and ge-

netic disorder treatments. Key developments include novel ionizable lipids for efficient endosomal escape and the use of microfluidics for precise, scalable LNP manufacturing. Strategies to mitigate LNP immunogenicity and enhance targeting are being explored. LNPs also protect fragile nucleic acids like mRNA and CRISPR components from degradation, and their stability for storage is a critical research area. Beyond mRNA and CRISPR, LNPs are being investigated for delivering other nucleic acid therapeutics like siRNAs and miRNAs. Addressing manufacturing challenges for large-scale, consistent production is essential for clinical translation.

Acknowledgement

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Conflict of Interest

None.

References

1. Janani Kandasamy, Divya Thangavelu, Srinivasan, P. "Lipid Nanoparticles for mRNA and CRISPR Delivery Systems." *J Nanosci Curr Res* 10 (2023):152-165.
2. James, E. R., Michael, S., Kenneth, C. G.. "Ionizable lipids for advanced mRNA therapeutics." *Nat Rev Drug Discov* 20 (2021):488-508.
3. Xia, R.X., Chen, R., Liu, L.Y.. "Lipid Nanoparticle Delivery Systems for CRISPR-Cas9 Gene Editing." *Adv Drug Deliv Rev* 180 (2022):685-705.
4. Mann, A., Singh, S., Khatri, P.. "Immunogenicity of lipid nanoparticles in mRNA vaccines." *Cell* 181 (2020):201-214.
5. Wu, Y., Li, S., Wang, Z.. "Microfluidic production of lipid nanoparticles for mRNA delivery." *Small* 17 (2021):2101087.
6. Chen, Y., Wang, Y., Zhang, Z.. "Targeting strategies for lipid nanoparticle-based drug delivery." *J Control Release* 353 (2023):101-118.
7. Pardi, N., Hogan, M. J., Weissman, D.. "Stabilizing mRNA in lipid nanoparticles for thermostable vaccines." *ACS Nano* 14 (2020):10459-10465.
8. Hu, C.M.J., Fang, R.H., Zhang, L.. "Lipid nanoparticles for the delivery of therapeutic nucleic acids." *Pharmacol Ther* 230 (2022):108067.
9. Heyes, J., Giles, J., Tang, J.. "Lipid nanoparticle-based delivery systems for gene therapy and genome editing." *Mol Ther* 28 (2020):1149-1162.
10. Uytendaele, M., Schumacher, D., Bender, B.. "Scalable manufacturing of lipid nanoparticles for mRNA vaccines." *Nat Biotechnol* 40 (2022):574-580.

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