

Research Article

Open Access

Investigation of the Possible Cytopathological Effect of Human Papillomavirus Infection on p-16^{INK4a} Overexpressed Urothelial Carcinomas of the Bladder in the Urine

Seiko Ogura^{1,2}, Toshitetsu Hayashi^{3,4}, Keiko Yano¹, Masami Sakurai⁵, Takaki Sakurai⁶ and Reiji Haba³

¹Department of Diagnostic Pathology, Saiseikai Noe Hospital, Osaka, Japan

²Graduate School of Medicine, Kagawa University, Kagawa, Japan

³Department of Diagnostic Pathology, Faculty of Medicine, Kagawa University, Kagawa, Japan

⁴Department of Diagnostic Pathology, Takamatsu Red Cross Hospital, Japan

⁵Osaka City University, Japan

⁶Department of Diagnostic Pathology, Kyoto University Hospital, Japan

Abstract

Background and objectives: Human papillomavirus (HPV) is well-known as the etiological agent for uterine cervical cancer or carcinoma of the oropharynx, anus and vulva. However, the cytopathologic effect of the HPV infection in urinary bladder tumors has rarely been reported. The present study has been carried out to search for cytopathologic differences between HPV positive and negative cases with urothelial carcinoma.

Methods: We examined 91 specimens from 63 patients who underwent transurethral resection or biopsy for papillary urothelial carcinomas between May 2010 and September 2012. p-16^{INK4a} expression was evaluated by immunohistochemistry. Detection of HPV DNA was carried out by in situ hybridization on formalin-fixed, paraffinembedded tissue sections. Urine smears were compared cytomorphologically between HPV-positive and HPV-negative cases.

Results: p-16^{INIK4a} overexpression was detected in 29 cases (31.9%, 29/91). Of them, HPV DNA was detected in 11 cases. No significant cytopathologic differences were found in tumor cells when HPV-positive and HPV-negative cases were compared.

Conclusion: HPV could be detected in urothelial carcinomas of the urinary bladder. There were no significant differences in cytopathologic features of urine smears between HPV-positive and HPV-negative cases with urothelial carcinomas. These results suggest that HPV infection in urinary tract does not add substantial clinically relevant importance to the carcinogenesis and cytomorphology of urothelial carcinomas.

Keywords: Human papillomavirus; Urinary bladder; p16^{INK4}; In situ hybridization; Urine; Cytology

Introduction

Human papillomavirus (HPV) is well-known as the major etiological agent for uterine cervical cancer. Its possible relationship with cancers of the oropharynx, anus and vulva has also been investigated [1-7]. Over the past few decades, a considerable number of studies have been conducted to determine the role of HPV in urinary bladder tumors [8-17]. However, the most of their studies have focused on the detection of HPV infection, and little attention has been paid to the cytologic morphology correlated with HPV infection in urinary bladder tumor. On the other hand, detection of HPV-DNA and overexpression of p16 INK4a has been reported at a high rate in cervical squamous cell carcinoma, but there are only occasional reports of HPV-DNA and p16 INK4a expression in urothelial carcinoma of the bladder. The present study has been carried out to search for cytopathologic differences between HPV positive and negative cases with urothelial carcinoma and the possible role of p16 expression in the carcinogenesis of urothelial carcinoma or association with the HPV infection.

Materials and Methods

Cases

We examined 63 patients who underwent transurethral resection for papillary urothelial carcinomas, based on the 2004 WHO classification from May 2010 to September 2012 in the Department of Urology, Saiseikai Noe Hospital, Osaka, Japan. The present study included 91 tissue samples and 76 urine specimens (20 voided urines and 56 bladder washings), and was approved by the institutional "Ethical Review Board" of Saiseikai Noe Hospital.

Immunohistochemistry for p16^{INK4a}

Immunohistochemical staining was carried out on 91 formalinfixed, paraffin-embedded blocks. From each block, 4- μ m-thick paraffin sections were cut and mounted on coated slides. The Ventana automated system (Ventana Automated Systems, Inc., Tucson, AZ) was used with an antibody p16^{INK4a} (clone E6H4, 1: 200 dilution, Becton-Dickinson Company, Ltd), and a paraffin-embedded section of uterine cervical intraepithelial neoplasia was included as a positive control for each run. The expression of p16^{INK4a} was evaluated in a semiquantitative fashion as follows: score 0, no staining; score 1, <20%; score

*Corresponding author: Toshitetsu Hayashi, Department of Diagnostic Pathology, Faculty of Medicine, Kagawa University, 1750-1 Ikenobe, Miki-cho, Kita-gun, Kagawa prefecture, Japan, Tel: +81-87-898-5111; Fax: +81-87-891-2371; E-mail: htoshi@med.kagawa-u.ac.jp

Received June 07, 2015; Accepted June 22, 2015; Published June 24, 2015.

Citation: Ogura S, Hayashi T, Yano K, Sakurai M, Sakurai T, et al. (2015) Investigation of the Possible Cytopathological Effect of Human Papillomavirus Infection on p-16INK4a Overexpressed Urothelial Carcinomas of the Bladder in the Urine. J Cytol Histol 6: 335. doi:10.4172/2157-7099.1000335

Copyright: © 2015 Ogura s, et al. This is an open-access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

Citation: Ogura S, Hayashi T, Yano K, Sakurai M, Sakurai T, et al. (2015) Investigation of the Possible Cytopathological Effect of Human Papillomavirus Infection on p-16INK4a Overexpressed Urothelial Carcinomas of the Bladder in the Urine. J Cytol Histol 6: 335. doi:10.4172/2157-7099.1000335

2, 21%–70%; score 3, >71% (Figures 1-3). The samples displaying score 3 immunoreactivity were considered to have overexpression of $p16^{INK4a}$ following the Nakazawa et al. criteria [18].

HPV DNA in situ hybridization

ISH was carried out on sections showing p16^{INK4a} overexpression on formalin-fixed, paraffin embedded tissue sections cut at a thickness of 4 µm. Following deparaffinization, the tissue was digested in protease solution for 7.5 minutes at 37°C. Slides were dehydrated in ascending order of ethanol concentration (70%, 95%, and 100%) for 1 minute each and air-dried. The probe used was Wide Spectrum HPV Biotinylated DNA probe, which detects HPVs 6, 11, 16, 18, 31, 33, 35, 45, 51, and 52 (Y1404, Dako North America Inc., Carpinteria, CA). It was applied to slides and coverslips were immediately placed over the probe solution and sealed with rubber cement. The slides were placed in a dry oven and denatured at 95°C for 5 minutes. Hybridization was carried out at 37°C overnight, and afterwards, the rubber cement and coverslip were removed and the slides immersed in the warmed stringent wash solution to incubate at 48°C for 30 minutes. Primary streptavidin-AP reagent was applied to the slides at room temperature for 20 minutes. BCIP/NBT substrate solution applied to the slides at room temperature for 2 hours. Tissues have been counterstained by Nuclear Fast Red solution. Positive ISH signal patterns were identified and classified by Cooper et al. [18,19] as follows: (1) punctate, when distinct dot-like intra-nuclear signals were stained (indicative of integrated HPV); (2) diffuse, when nuclei were completely stained (indicative of episomal HPV).



Figure 1: (A) Papillary urothelial carcinoma, low grade; (B) Score 0: no staining Expression of p16 is not detected at all. (a. H.E. stain 100X; b. $p16^{NNC4a}$ immunostain, 100X).





Comparison of cytopathological finding

Urine cytologic specimens consisted of 20 voided urines and 56 bladder washings. They were prepared by conventional methods, such as Cytospin (Thermo Shandon, Pittsburgh, PA) or smear preparations following centrifugation, and fixed in 95% ethanol or airdried for Papanicolaou staining or May-Grünwald-Giemsa staining. We compared the cytomorphological difference in urinary cytology between HPV-positive and HPV-negative cases. The cytopathological features of tumor cells were reviewed, including their background, arrangement, and cellular pleomorphism (variation of cell diameter occurring more than twice), and nuclear and cytoplasmic features. Furthermore, the presence of cytomorphological parameters associated with HPV infection considered previously by Bollmann et al. [20], such like koilocytosis, dyskeratocytes, abortive koilocytes, mild dyskeratosis, parakeratosis, mild nuclear hyperchromasia, mild nuclear variations, binucleation or multinucleation, measles cells, keratohyalin and keratohyalin-like granules, macrocytes, and cytoplasmic folding were reviewed.

Statistical analysis

Data were entered into JMP8.0 (SAS Institute, Inc. North Carolina, USA) software and analyzed. Significant differences between groups were found using the Chi-square test and Fisher's exact test. *P*-values<0.05 were considered significant.



Figure 3: (A) Papillary urothelial carcinoma, high grade (H.E. stain 100X); (B) Score3: Over expression of p16 is detected in the nuclei and/or cytoplasm in more than 70%. (p16^{INK4a} immunostain, 100X)

Results

Patient characteristics

Clinicopathological characteristics of our samples are shown in Table 1. The mean age at the time of urothelial carcinoma diagnosis was 75.3 years (range, 46-91 years). The patients included 72 males and 19 females. Fifty-four patients had low grade urothelial carcinomas (LGUC), and 37 patients had high grade urothelial carcinomas (HGUC). Seventy-four patients had non-invasive tumors, and 17 patients had invasive disease. Sixty-one patients had original tumors and 30 patients had recurrent disease.

Immunohistochemistry for p16^{INK4a}

Twenty-nine of 91 (31.9%) patients showed overexpression (Score 3) of $p16^{INK4a}$ (Table 1). With regard to tumor grade, stage, and original or recurrent disease, overexpression of $p16^{INK4a}$ was detected in 14 (25.9%) of 54 LGUC, 15 (40.5%) of 37 HGUC, 22(29.7%) of 74 non-invasive tumors, 7 (41.2%) of 17 invasive disease, 25 (41.0%) of 61 original tumors, and 4 (13.3%) of 30 recurrent disease.

Detection of HPV-DNA

HPV-DNA was detected in 11 (37.9%) out of 29 cases of p16^{INK4A} overexpression (Table 2). As for tumor grade, stage, and original or recurrent, HPV-DNA was detected in 4 (28.6%) of 14 LGUC, 7 (46.7%) of 15 HGUC, 9 (40.9%) of 22 non-invasive tumors, 2 (28.6%) of 7 invasive cancer, 9 (36%) of 25 original tumors, and 1 (25%) of 4 recurrent disease. All of the HPV DNA-positive specimens showed punctate signals in the nuclei (Figure 4).

Comparison of cytologic findings of HPV positive UC and HPV negative UC

Cytopathologic findings could be evaluated in 29 samples (8 voided urine, 21 bladder washings) consisting of 11 HPV-positive cases and 18 HPV-negative cases. The cytomorphological findings are summarized in Table 3. No significant differences were found in the cytology of tumor cells when HPV-positive case and HPV-negative cases were compared (Figure 5 and Table 3 with corresponding p-values). The cytomorphological parameters associated with HPV infection were observed in 2 (18.2%) cases of HPV-positive cases, and in three (16.7%) HPV-negative cases of benign squamous cells. The cytopathologic parameters observed were bi- or multinucleation, abortive koilocytes, and keratohyalin-like granules (Figure 6). No significant differences were present in cytomorphological findings among HPV-positive and HPV-negative cases.

Discussion

In this study, detection of HPV-DNA by ISH was performed only on cases which had p16^{INK4a} protein as a surrogate marker of HPV infection in cervical tissue with overexpression. p16^{INK4a} binds to cyclindependent kinase4 and inhibits its activity [21,22]. In the cervix, HPV infection inactivates RB protein by HPV E7 protein, and then the free transcription factor E2F increases. The cell cycle proceeds from G1 phase to S phase and cell proliferation is enhanced. At this time, since the E2F protein induces the expression of p16^{INK4a}, it is overexpressed. Detection of HPV-DNA and overexpression of p16^{INK4a} has been reported at a high rate in cervical squamous cell carcinoma [23-25], but reports of HPV-DNA and $p16^{{\scriptscriptstyle\rm INK4a}}$ expression of the bladder have been few. Steinestel et al. [26] found the expression of p16^{INK4a} in 25 specimens of 27 (92.6%), but HPV-DNA was not detected. Moreover, Piaton et al. [27,28] showed that in three patients with p16^{INK4a} immunoreactive tumor cells and high risk-HPV in the urine, HPV genotyping and in situ hybridization for high risk-HPV were negative in tissue sections. However, Shigehara et al. [16] examined 106 cases of urothelial carcinoma, 4 cases of squamous cell carcinoma, 6 cases of adenocarcinoma, and one case of another bladder cancer and found HPV-DNA in 18 out of 117samples (15%). Among them, they reported score3 (>50% of the cells were positive) staining of p16 INK4a in10 samples [16]. In this study, we admitted overexpression in 29 of 91 papillary urothelial carcinoma samples. We conducted ISH, and HPV-DNA was detected in 4 of

		p16 ^{INK4a} status				
		Score 0	Score 1	Score 2	Score 3	
n=91		42	12	8	29	
Age (mean ± S.D)	75.3 ± 8.7					
Gender						
Male	72	29	11	6	26	
Female	19	13	1	2	3	
Tumor grade						
Low	54	32	5	3	14	
High	37	10	7	5	15	
Tumor invasion						
noninvasive	74	38	7	7	22	
invasive	17	4	5	1	7	
original	61	24	7	5	25	
recurrence	30	18	5	3	4	

Score 0, no staining; Score1, <20%; Score2, 21%-70%; Score3, >71%

Table 1: Clinicopathological characteristics of the patients and correlation with p16 $^{\mbox{\tiny NK4a}}$ immunophenotype.

Citation: Ogura S, Hayashi T, Yano K, Sakurai M, Sakurai T, et al. (2015) Investigation of the Possible Cytopathological Effect of Human Papillomavirus Infection on p-16INK4a Overexpressed Urothelial Carcinomas of the Bladder in the Urine. J Cytol Histol 6: 335. doi:10.4172/2157-7099.1000335

Page 4 of 6

Case no.	Age	Sex	Tumor grade	Tumor invasion	Original/Recurrence	ISH signal pattern
1	64	М	low	noninvasive	recurrence	punctate
2	81	М	low	noninvasive	original	punctate
3	65	М	low	noninvasive	original	punctate
4	79	М	low	noninvasive	original	punctate
5	79	F	high	noninvasive	original	punctate
6	46	М	high	noninvasive	original	punctate
7	60	М	high	noninvasive	original	punctate
8	68	М	high	noninvasive	original	punctate
9	77	М	high	noninvasive	original	punctate
10	72	М	high	invasive	original	punctate
11	74	М	high	invasive	original	punctate

M: Male; F: Female

 Table 2: Clinicopathological features of the eleven cases of UC with presence of HPV-DNA.

	Voided urine n=5		Bladder washings n=12		
	HPV (+) n=1	HPV (-) n=4	HPV (+) n=4	HPV (-) n=8	p-value
General features					
Background					
clean	0	2 (50%)	1 (25%)	4 (50%)	0.338
Inflammation	0	0	1 (25%)	0	0.294
Bloody	1 (100%)	2 (50%)	3 (75%)	3 (37.5%)	0.294
Tumor diathesis	1 (100%)	0	1 (25%)	1 (12.5%)	0.191
Arrangement					1.000
Mostly clusters	0	2 (50%)	2 (50%)	4 (50%)	
Single cell and clusters	0	1 (25%)	1 (25%)	2 (25%)	
Mostly single cells	1 (100%)	1 (25%)	1 (25%)	2 (25%)	
Cell uniformity					0.117
Monomorphic	0	3 (75%)	2 (50%)	7 (87.5%)	
Pleomorphic	1 (100%)	1 (25%)	2 (50%)	1 (12.5%)	
Nuclear features					
Chromatin pattern					1.000
Fine granular	0	3 (75%)	2 (50%)	2 (25%)	
Granular	1 (100%)	1 (25%)	2 (50%)	6 (75%)	
Conspicuous nucleoli					0.538
Absent	1(100%)	3 (75%)	2 (50%)	7(87.5%)	
Indistinct/small	0	1 (25%)	1 (25%)	1 (12.5%)	
Present/occasional	0	0	1 (25%)	0	
Cytoplasmic features					
Border					0.620
Indistinct	0	1 (25%)	3 (75%)	4 (50%)	
Distinct	1 (100%)	3 (75%)	1 (25%)	4 (50%)	
Texture					0.319
Homogenous	1 (100%)	2 (50%)	0	6 (75%)	
Variable	0	2 (50%)	4 (100%)	2 (25%)	

Table 3: Comparison of cytopathological findings between HPV-positive UC and HPV-negative UC.

14 LGUC cases (28.6%) and 7 of 15 HGUC cases (46.7%). Previous reports showed either a high rate in Grade 3 [10,17] or a higher rate in Grade 1 [16]. In this study, HGUC showed a higher detection rate of HPV-DNA. This controversy is probably related to our selected cases having IHC overexpression of p16^{INK4a}. Criteria of overexpression of p16^{INK4a} in tissue specimens varied in the literature [3,16,25,27,29]. In our study, overexpressed positive findings meant that there was diffused distribution in more than 70% of the nucleus and cytoplasm of tumor cells. In Shigehara's report [16], among 18 HPV-DNA positive cases there were 6 cases of p16^{INK4a} from 20% to 50%. They were 3 cases of urothelial carcinoma, Grade 1>2, and 3 cases of urothelial carcinoma, Grade 1>2, and 3 cases of urothelial carcinoma, Grade 1>2, MPV-DNA detection by LGUC will rise.

Differences in cellular findings caused by the presence or absence of HPV infection was not observed in papillary urothelial carcinoma in this study. Having an HPV infection suggests the presence of atypical squamous cells, koilocytes, dyskeratotic cells and multinucleated cells in cervical smears [20]. These cells appear mainly at the stages of CIN1 and CIN2 but are rare in the stage of invasive cancer, except when associated with CIN1 and CIN2 lesions. HPV infection is not only involved in squamous cell carcinoma, but also adenocarcinoma in the uterine cervix [30,31]. However, the cellular changes of HPV infection in glandular cells are not well established. In urothelial cells also, cellular changes due to HPV infection is not well understood. HPV-DNA detected by ISH method in this study showed the all integrate pattern; no episomal pattern was observed. Because of these results, finding

Citation: Ogura S, Hayashi T, Yano K, Sakurai M, Sakurai T, et al. (2015) Investigation of the Possible Cytopathological Effect of Human Papillomavirus Infection on p-16INK4a Overexpressed Urothelial Carcinomas of the Bladder in the Urine. J Cytol Histol 6: 335. doi:10.4172/2157-7099.1000335

Page 5 of 6

the cytologic differences between HPV-DNA positive and negative UC cases on urine was difficult. Also, in the urine cytology specimens, because HPV-DNA positive cases can focus on non-tumor cells unlike HPV-DNA negative cases, significant differences in the incidence of abortive koilocytes or bi-nucleation were observed. Atypical squamous cells that appeared in urine cytology specimens of HPV-DNA positive cases in this study were of the "Non-classic type" rather than "Classic type" which includes koilocytes and dyskeratotic cells. There are many established reports of the appearance of atypical squamous cells of non-classic type in uterine cervical smears, and there are types of observed



Figure 4: Detection of HPV DNA with in situ hybridization method showing nuclear punctate signals (HPV ISH 1000X).



Figure 5: Cytopathological findings of tumor cells. The difference was not observed in the cell findings of tumor cells with HPV-negative and positive cases. (A) HPV negative of LGUC; (B) HPV positive of HGUC; (C) HPV negative of HGUC, Papanicolaou stain 1000X).



Figure 6: Cytopathological features of squamous cells. "Non-classical type" observed in voided urine cytology with HPV-positive cases (A and B: binucleate cell, papanicolaou stain 400X) and washing fluid cytology with HPV-negative cases (C: keratohyalin-like granules, May-Grunwald-Giemsa stain 400X).

parameters or combination of parameters indicative of HPV infection [20,31]. However, atypical squamous cells that appeared in HPV-DNA positive cases were observed in both cases of the voided urine and it was not possible to limit them from the urinary bladder epithelium.

Cancer generated from uterine cervix, oral cavity, tonsil, pharynx, anus, vulva, vagina and penis are carcinogenic by HPV infection as observed by IARC. Although attempts have been made to study urinary bladder cancer associated with HPV infection, carcinogenic HPV in bladder cancer is still controversial. Our results support the theory that HPV infection in urinary tract does not add clinically relevant importance to the carcinogenesis and cytomorphology of urothelial carcinomas. Further investigations are needed to clarify the role and correlation between bladder cancer and HPV infection.

Acknowledgements

This study was supported by a grant for the Nishida Furuse Memorial Fund from Saiseikai Noe Hospital.

References

- D'Souza G, Kreimer AR, Viscidi R, Pawlita M, Fakhry C, et al. (2007) Casecontrol study of human papillomavirus and oropharyngeal cancer. N Engl J Med 356: 1944-1956.
- Kreimer AR, Clifford GM, Boyle P, Franceschi S (2005) Human papillomavirus types in head and neck squamous cell carcinomas worldwide: a systematic review. Cancer Epidemiol Biomarkers Prev 14: 467-475.
- Evans MF, Matthews A, Kandil D, Adamson CS, Trotman WE, et al. (2011) Discrimination of 'driver' and 'passenger' HPV in tonsillar carcinomas by the polymerase chain reaction, chromogenic in situ hybridization, and p16(INK4a) immunohistochemistry. Head Neck Pathol 5: 344-348.
- De Vuyst H, Clifford GM, Nascimento MC, Madeleine MM, Franceschi S (2009) Prevalence and type distribution of human papillomavirus in carcinoma and intraepithelial neoplasia of the vulva, vagina and anus: a meta-analysis. Int J Cancer 124: 1626-1636.
- Stanley MA, Winder DM, Sterling JC, Goon PK (2012) HPV infection, anal intraepithelial neoplasia (AIN) and anal cancer: current issues. BMC Cancer 12: 398.
- Alibek K, Karatayeva N, Bekniyazov I (2012) The role of infectious agents in urogenital cancers. Infect Agent Cancer 7: 35.
- 7. Zandberg DP, Bhargava R, Badin S, Cullen KJ (2013) The role of human papillomavirus in nongenital cancers. CA Cancer J Clin 63: 57-81.
- Kitamura T, Yogo Y, Ueki T, Murakami S, Aso Y (1988) Presence of human papillomavirus type 16 genome in bladder carcinoma in situ of a patient with mild immunodeficiency. Cancer Res 48: 7207-7211.
- Furihata M, Inoue K, Ohtsuki Y, Hashimoto H, Terao N, et al. (1993) Highrisk human papillomavirus infections and overexpression of p53 protein as prognostic indicators in transitional cell carcinoma of the urinary bladder. Cancer Res 53: 4823-4827.
- LaRue H, Simoneau M, Fradet Y (1995) Human papillomavirus in transitional cell carcinoma of the urinary bladder Clin Cancer Res 1: 435-440.
- Lopez-Beltran A, Escudero AL, Vicioso L, Muñoz E, Carrasco JC (1996) Human papillomavirus DNA as a factor determining the survival of bladder cancer patients. Br J Cancer 73: 124-127.
- Chan KW, Wong KY, Srivastava G (1997) Prevalence of six types of human papillomavirus in inverted papilloma and papillary transitional cell carcinoma of the bladder: an evaluation by polymerase chain reaction. J Clin Pathol 50: 1018-1021.
- Youshya S, Purdie K, Breuer J, Proby C, Sheaf MT, et al. (2005) Does human papillomavirus play a role in the development of bladder transitional cell carcinoma? A comparison of PCR and immunohistochemical analysis. J Clin Pathol 58: 207-210.
- Barghi MR, Hajimohammadmehdiarbab A, Moghaddam SM, Kazemi B (2005) Correlation between human papillomavirus infection and bladder transitional cell carcinoma. BMC Infect Dis 5: 102.
- Gopalkrishna V, Srivastava AN, Hedau S, Sharma JK, Das BC (1995) Detection of human papillomavirus DNA sequences in cancer of the urinary bladder by in situ hybridisation and polymerase chain reaction. Genitourin Med 71: 231-233.
- Shigehara K, Sasagawa T, Kawaguchi S, Nakashima T, Shimamura M, et al. (2011) Etiologic role of human papillomavirus infection in bladder carcinoma. Cancer 117: 2067-2076.
- De Gaetani C, Ferrari G, Righi E, Bettelli S, Migaldi M, et al. (1999) Detection of human papillomavirus DNA in urinary bladder carcinoma by in situ hybridisation. J Clin Pathol 52: 103-106.
- Nakazawa K, Murata S, Yuminamochi T, Ishii Y, Ohno S, et al. (2009) p16(INK4a) expression analysis as an ancillary tool for cytologic diagnosis of urothelial carcinoma. Am J Clin Pathol 132: 776-784.
- Cooper K, Herrington CS, Graham AK, Evans MF, McGee JO (1991) In situ evidence for HPV 16, 18, 33 integration in cervical squamous cell cancer in Britain and South Africa. J Clin Pathol 44: 406-409.

 Bollmann N, Bankfalvi A, Trosic A, Speich N, Schmitt C, et al. (2005) Can we detect cervical human papillomavirus (HPV) infection by cytomorphology alone? Diagnostic value of non-classic cytological signs of HPV effect in minimally abnormal Pap tests. Cytopathology 15: 13-21.

Page 6 of 6

- Takahashi A, Yamakoshi K, Hara E (2005) Role of p16INK4a/Rb pathway in cell cycle control and cellular senescence. Jikken Igaku 23: 195-200.
- Burd EM (2003) Human papillomavirus and cervical cancer. Clin Microbiol Rev 16: 1-17.
- 23. Nishio S, Fujii T, Nishio H, Kameyama K, Saito M, et al. (2013) p16INK4a immunohistochemistry is a promising biomarker to predict the outcome of low grade cervical intraepithelial neoplasia: comparison study with HPV genotyping. J GynecolOncol 24: 215-221.
- 24. Izadi-Mood N, Asadi K, Shojaei H, Sarmadi S, Ahmadi SA, et al. (2012) Potential diagnostic value of P16 expression in premalignant and malignant cervical lesions. J Res Med Sci 17: 428-433.
- 25. Klaes R, Friedrich T, Spitkovsky D, Ridder R, Rudy W, et al. (2001) Overexpression of p16(INK4A) as a specific marker for dysplastic and neoplastic epithelial cells of the cervix uteri. Int J Cancer 92: 276-284.
- 26. Steinestel J, Cronauer MV, Müller J, Al Ghazal A, Skowronek P, et al. (2013) Overexpression of p16(INK4a) in urothelial carcinoma in situ is a marker for MAPK-mediated epithelial-mesenchymal transition but is not related to human papillomavirus infection. PLoS One 8: e65189.
- Piaton E, Casalegno JS, Advenier AS, Decaussin-Petrucci M, Mege-Lechevallier F, et al. (2014) p16(INK4a) overexpression is not linked to oncogenic human papillomaviruses in patients with high-grade urothelial cancer cells. Cancer Cytopathol 122: 760-769.
- Advenier AS, Casalegno JS, Mekki Y, Decaussin-Petrucci M, Mège-Lechevallier F, et al. (2015) Genotyping of high-risk human papillomaviruses in p16/Ki-67positive urothelial carcinoma cells: even a worm will turn. Cytopathology 26: 106-113.
- Agoff SN, Lin P, Morihara J, Mao C, Kiviat NB, et al. (2003) p16(INK4a) expression correlates with degree of cervical neoplasia: a comparison with Ki-67 expression and detection of high-risk HPV types. Mod Pathol 16: 665-673.
- Murphy N, Ring M, Heffron CC, King B, Killalea AG, et al. (2005) p16INK4A, CDC6, and MCM5: predictive biomarkers in cervical preinvasive neoplasia and cervical cancer. J Clin Pathol 58: 525-534.
- Sana DE, Mayrink de Miranda P, Pitol BC, Moran MS, Silva NN, et al. (2013) Morphometric evaluation and nonclassical criteria for the diagnosis of HPV infection and cytological atypia in cervical samples. Diagn Cytopathol 41: 785-792.