

Inhibitory Potential of Phyto-Constituents from Sorghum Stem on Neuro-modulatory Enzymes: A Computational Approach

Damilola A Omoboyowa^{1*}, Toheeb A Balogun¹ and Oluwatosin A Saibu²

¹Department of Biochemistry, Adekunle Ajasin University, Akungba-Akoko, Ondo State, Nigeria

²Department of Environmental Toxicology, University of Duisburg-Essen, North Rhine-Westphalia, Germany

Abstract

Vertebrate cholinesterases and monoamine oxidase (MOA) play key role in the regulation of the cholinergic system. Inhibitions of these enzymes have emerged as promising target for the management of neurological disorders. In the present study, cholinesterase (acetylcholinesterase (Ache) and butyrylcholinesterase (Bche)) and MOA inhibition potential of secondary metabolites from sorghum stem was elucidated using Glide (Schrodinger maestro 2017-1). The results showed that, the phyto-constituents from sorghum stem binds with higher affinity and lower free energy compared to the standard ligand with quercetin, kaempferol and catechin having glide score of -11.667, -10.599 and -10.535 respectively against Ache. Rutin, quercitrin and chlorogenic acid showed glide score of -12.643, -10.124 and -9.043 against Bche while epicatech, quercetin and kaempferol showed glide score of -10.424, -9.830 and -9.804 against monoamine oxidase. Quercetin has lower glide score against the three enzymes compared to prostigmine (standard ligand). These secondary metabolites provide evidence as small molecules in management of neurodegenerative disorders.

Keywords: Cholinesterase • Monoamine oxidase • Phyto-constituents • Sorghum • Quercetin

Introduction

Neurodegenerative disorders are diseases of the central and peripheral nervous system, associated with protein aggregation and inflammation as well as oxidant generation in the central nervous system (CNS) [1]. These disorders includes: Alzheimer disease, brain tumors, Parkinson's disease, epilepsy etc. Various neurological disorders possess common features at cellular and sub-cellular biological events including the synthesis of new structural and functional protein molecules at the cytosol and endoplasmic reticulum [1]. Insufficient synthesis of neurotransmitters and breaking down of chemical messengers in the synaptic cleft resulting from higher activity of neuronal enzymes such as cholinesterase and monoamine oxidases lead to low concentration of this neurotransmitter which has been linked with cognitive abnormalities [2], since high activity of these enzymes has been attributed to be part of the biological mechanisms associated with neurodegenerative disorders [1].

Cholinesterase including acetylcholinesterase (Ache) and butyrylcholinesterase (Bche) are involved in the hydrolysis of acetylcholine in the synaptic region while monoamine oxidases (MAOs) are involved in the oxidative deamination of endogenous monoamines neurotransmitter; this enzyme is involved in the deamination of biogenic amines and is present in the CNS and PNS tissues [3]. Neurotransmission by biogenic monoamines is involved in brain functions. These amines balance utilizes the enzyme in neuronal and glial cells. Inhibition of this enzyme, increases monoamines concentration in the synaptic cleft [4].

Synthetic drugs targeting Ache/Bche and MOA are used for the management of neurodegenerative disorders; they are not without side

effects such as hepatotoxicity, gastrointestinal disorders, dizziness, diarrhea, vomiting, nausea and pharmacokinetics disadvantages [5,6]. Hence, investigation of natural compounds with Ache/Bche and MAO inhibition potentials used in traditional management of neurological disorders has been the means of searching for new drugs that will liberate human race from these life-threatening disorders.

Sorghum (*Sorghum color*) is a genus of flowering plants in the family Poaceae, it is one of the most cultivated plants in the world [7]. Antioxidants and bioactive chemicals are abundant in sorghum grains [8]. The stem of the plant has been involved in herbal medicine for management of anaemia and malaria [9]. The sorghum stem extract has been reported to modulate acetylcholinesterase activities [7]. Oboh et al. [7] reported the phenolic compounds present in sorghum stem which might contribute to the Ache/Bche and MAO inhibitory activities of the plant. However, screening such large number of drug candidates may not be possible due to cost both in term of money and time. Application of bioinformatics tools such as molecular docking may lead to development of neuro-modulatory compounds with Ache/Bche and MAO inhibitory activities. The present study is aimed to determine the molecular interaction of prostigmine and ten (10) phyto-compounds from sorghum stem with Ache/Bche and MAO target enzymes for neurodegenerative drugs. A picture of Sorghum is shown in Figures 1 and 2.

Materials and Methods

Ligands preparation

Ten (10) characterized phyto-compounds from sorghum stem used in this study were obtained from Oboh et al. [7]. The compounds were used in the production of compound library downloaded from NCBI PUBCHEM database (www.ncbi.nlm.nih.gov/pccompound) in 2D format. The phyto-compounds and prostigmine were prepared according to David et al. [10].

Protein preparation

Crystallized 3D structure of the target proteins; acetylcholinesterase, butyrylcholinesterase and monoamine oxidase were downloaded from protein data bank (<http://www.rcsb.org/pdb/home.do>). The 3D structures were viewed with maestro 11.5 interface and prepared with wizard at pH 6 ± 1. Water molecules and other interfering ligands were deleted from the protein.

*Address for Correspondence: Damilola A. Omoboyowa, Department of Biochemistry, Adekunle Ajasin University, Akungba-Akoko, Ondo State, Nigeria, Tel: + 2347032665874, E-mail: damilola.omoboyowa@aaua.edu.ng

Copyright: © 2020 Omoboyowa DA, et al. This is an open-access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

Received 31 August 2020; **Accepted** 24 November 2020; **Published** 01 December 2020



Figure 1. A picture of *Sorghum color*.

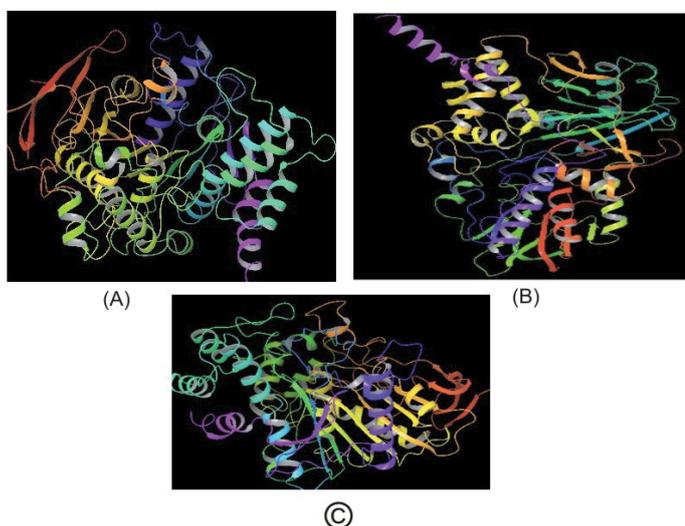


Figure 2. 3D Structures of Proteins (a) Acetylcholinesterase (b) Butyrylcholinesterase (c) Monoamine oxidase.

Receptor grid generation

The area of interaction between the proteins and ligands were generated with receptor grid generation tool in maestro 11.5 which is the area around the binding pocket in term of co-ordinates x, y and z.

Molecular docking using glide

Molecular docking was carried out using glide tool on maestro 11.5 according to Schrodinger Release 2017-1. The Ligands library (Phyto-compounds) were docked into the binding site of the proteins according to standard precision algorithm with the ligand sampling treated as flexible. The ligands interaction tool was used to view the interaction diagram of the ligands with the residues at the binding site of the proteins [11].

ADME toxicity screening

The ligands were subjected to absorption, distribution, metabolism, excretion screening using the Qikprop tool on maestro 11.5 according to Schrodinger Release [12].

Results and Discussion

The interaction of ligands and binding sites of proteins often give credence

Table 1. Docking results of phyto-compounds from sorghum stem against acetylcholinesterase.

S. No	Entry Name	Dock score	Glide score	Glide model
1	Quercetin	-11.656	-11.667	0
2	Kaempferol	-10.587	-10.599	-71.258
3	Catechin	-10.535	-10.535	-64.317
4	Epicatechin	-9.857	-9.857	-70.694
5	Caffeic acid	-7.201	-7.201	0
6	Kaempferol	-7.188	-9.53	-61.405
7	Chlorogenic acid	-6.869	-6.871	-60.644
8	Gallic acid	-6.142	-6.142	-36.276
9	Ferulic acid	-5.913	-5.913	-35.127
10	Prostigmine (control)	-8.933	-8.933	0

Table 2. Docking results of phyto-compounds from sorghum stem against butyrylcholinesterase.

S. No	Entry Name	Dock score	Glide score	Glide model
1	Rutin	-12.632	-12.643	-123.925
2	Quercitrin	-10.113	-10.124	-85.879
3	Chlorogenic acid	-9.041	-9.043	-70.639
4	Quercetin	-8.416	-8.428	-63.86
5	Catechin	-7.324	-7.324	0
6	Epicatechin	-7.064	-7.064	-59.347
7	Gallic acid	-6.8	-6.8	-46.136
8	Kaempferol	-6.284	-6.296	-54.52
9	Caffeic acid	-4.154	-4.154	0
10	Ferulic acid	-3.146	-3.146	0
11	Prostigmine (control)	-3.116	-3.116	-36.78

Table 3. Docking results of phyto-compounds from sorghum stem against monoamine oxidase.

S. No	Entry Name	Dock score	Glide score	Glide model
1	Epicatechin	-10.424	-10.424	-64.623
2	Quercetin	-9.818	-9.83	-58.027
3	Kaempferol	-9.792	-9.804	-62.994
4	Catechin	-9.276	-9.276	-62.78
5	Chlorogenic acid	-8.708	-8.711	0
6	Ferulic acid	-6.932	-6.932	-40.458
7	Caffeic acid	-6.825	-6.825	-40.145
8	Gallic acid	-6.588	-6.588	-37.692
9	Prostigmine (control)	-2.866	-2.866	-48.356

to protein function and drug discovery [13]. The relationship between proteins and ligands can be quantified base on the electrostatic interaction; this interaction is approximated as the docking score of the ligand in relation to active site of the protein [10,14]. The docking results from this study showed the interaction of ten (10) compounds from sorghum stem downloaded from NCBI database against Ache/Bche and MAO. The docking results and ADME toxicity screening of the compounds and standard ligand (prostigmine) were shown in Tables 1-4.

The docking scores of most of the compounds at the binding sites of Bche and MAO revealed that, most of the compounds bind to the active site of the enzymes with lower docking score compared to the standard ligand. Quercetin, kaempferol, Catechin and epicatechin were showed to have lower binding score against Ache compared to progstimine. The results obtained agree with the findings of Srikanth et al. [15] who reported comparative molecular docking study of phytochemicals from *Morinda citrifolia* on Ache and Bche. Inhibition of the enzymes is dependent on the formation of different bonds between amino acids residues at the binding site and the ligands [10].

The amino acid residues at the bind ingregion of Ache that stabilize Ache-quercetin interaction are TYR 70, TYR 121, TRP 54, GLY 117, TYR 130, GLH 199, PHE 330 and HIS 440. The interaction between the Bche and rutin was

Table 4. *In silico* ADME toxicity screening.

S. No	Entry Name	ROF	HOA	M.W	Qlogkhsa	Comp ID	M.F
1	Caffeic acid	0	2 (Medium)	180.16	-0.804	689043	C ₈ H ₈ O ₃
2	Catechin	0	2 (Medium)	290.272	-0.415	9064	C ₁₅ H ₁₄ O ₆
3	Chlorogenic acid	1	1 (Low)	354.313	-0.913	1794427	C ₁₆ H ₁₈ O ₉
4	Epicatechin	0	2 (Medium)	290.272	-0.415	72276	C ₁₅ H ₁₄ O ₆
5	Kaempferol	0	3 (High)	286.24	-0.191	5280863	C ₁₅ H ₁₀ O ₆
6	Quercetin	0	2 (Medium)	302.24	-0.348	5280343	C ₁₅ H ₁₀ O ₇
7	Quercitrin	2	1 (Low)	448.382	-0.639	5280459	C ₂₁ H ₂₀ O ₁₁
8	Rutin	3	1 (Low)	610.524	-1.273	5280805	C ₂₇ H ₃₀ O ₁₆
9	Ferulic acid	0	3 (High)	194.187	-0.612	445858	C ₁₀ H ₁₀ O ₄
10	Gallic acid	0	2 (Medium)	170.121	-0.985	370	C ₇ H ₆ O ₅

ROF Viol: Rule of Five violations. HOA: Human Oral Absorption. M.W: Molecular Weight of compounds. Normal range is between 130.0 and 725.0. M.F: Molecular formula. QlogKhsa: Prediction of binding to human serum albumin. Normal range between -1.5 to 1.5

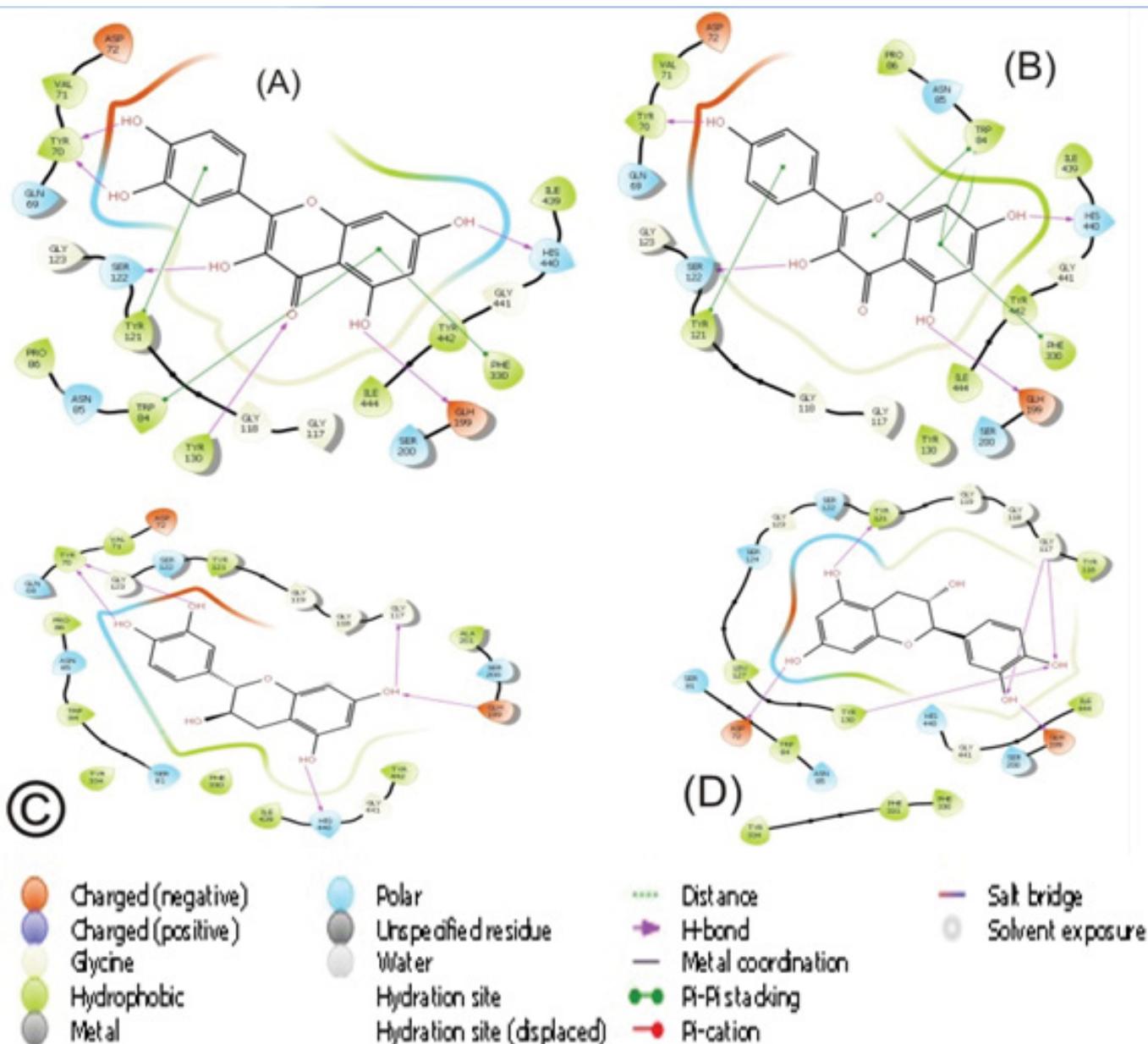


Figure 3. 2D stick diagram of (a) Quercetin; (b) Kaempferol; (c) Catechin; (d) Epicatechin illustrating hydrogen bonds and pi-pi stacking formed with the aminoacid residues at the active site of Acetylcholinesterase.

stabilized by ASP 70, ALA 328, PHE 329, LEU 286, HIP 438, ASN 289 and GLH 197 while MAO-epicatechin interaction was stabilized by LEU 177, PHE 343, GLY 434 and ARG 42. These amino acids interacted with the ligands by forming hydrogen-bond or pi-pi stacking as shown in Figures 3-5. These

bonds may account for the lowest binding energy observed in the interaction since hydrogen-bond plays a crucial role in determining the specificity of ligand binding [16,17].

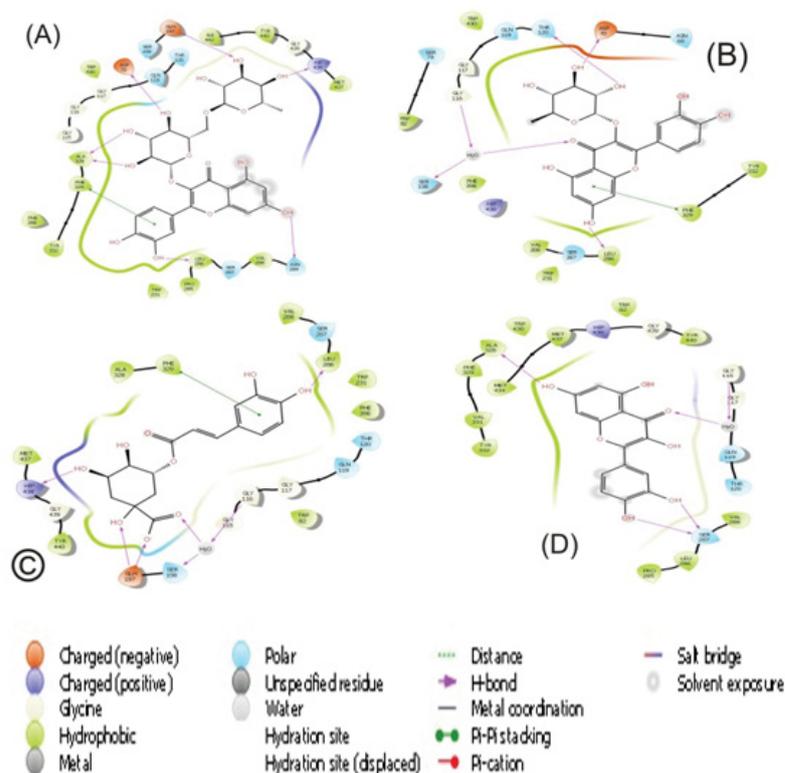


Figure 4. 2D stick diagram of (a) Rutin; (b) Quercitrin; (c) Chlorogenic acid; (d) Quercetin illustrating hydrogen bonds and pi-pi stacking formed with the aminoacid residues at the active site of Butyrylcholinesterase.

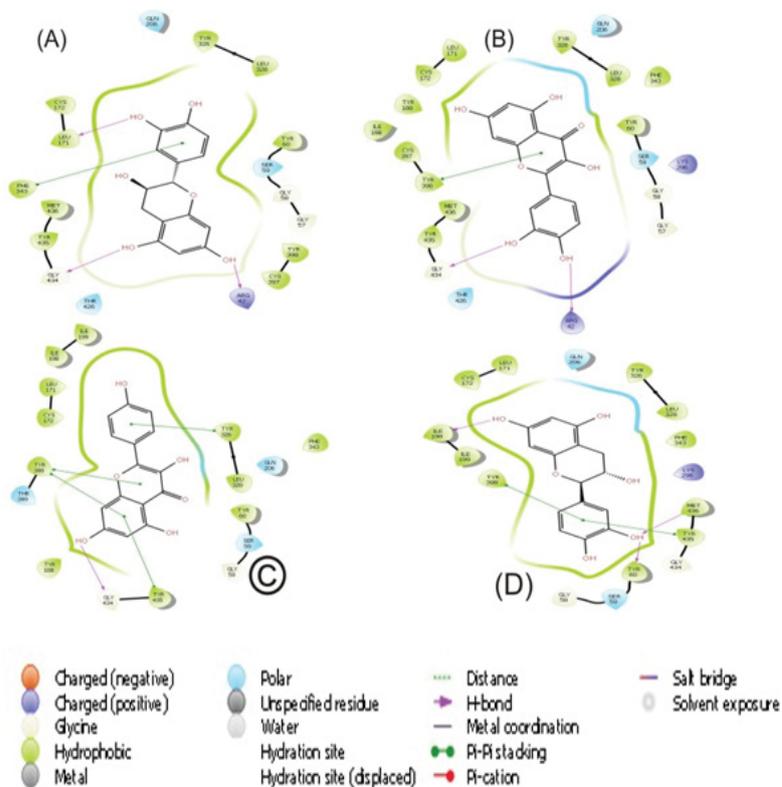


Figure 5. 2D stick diagram of (a) Epicatechin; (b) Quercetin; (c) Kaempferol; (d) Catechin illustrating hydrogen bonds and pi-pi stacking formed with the aminoacid residues at the active site of Monoamine oxidase.

The absorption, distribution, metabolism and elimination (ADME) toxicity screening of compounds depict the efficiency and ability of the ligands to reach its target protein and rate of excretion from the body [10]. The bioinformatics analysis of ADME screening follows the Lipinski's rule of five (ROF) which is required to dictate the drug-like nature of compounds. These criteria include:

molecular weight (< 500 Da), hydrogen bond (≤ 5) and water partition coefficient (\log^p) (<5) [10]. Table 4 revealed that, the ADME profile of compounds from sorghum stem agrees with these rules and majorly with medium human oral absorption (HOA). Therefore, they exhibit drug-like character. The result of the docking analysis and ADME screening revealed that, quercetin, kaempferol, catechin, epicatechin, rutin, quercitrin and chlorogenic acid from

sorghum stem might be of pharmaceutical importance for the management of neurodegenerative disorders.

Conclusion

From the *in silico* study, phyto-compounds from sorghum stem could be potential inhibitor of Ache/Bche and MAO proteins. These phyto-compounds showed better molecular interaction than the standard ligand (prostigmine) via hydrogen bond and pi-pi stacking. These results depicted new insight into the therapeutic use of sorghum for the management of neurodegenerative disorders.

Acknowledgements

The Authors wish to acknowledge Adedokun N.S. of Center for Bioinformatics and Drug Discovery, Adekunle Ajasin University, Akungba-Akoko, Ondo State for providing the software for the analysis.

Conflict of Interest

The authors declare no conflict of interest.

References

- Rasool, Mahmood, Arif Malik, and Muhammad Saeed Qureshi, et al. "Recent updates in the treatment of neurodegenerative disorders using natural compounds." *Evid-Based Compl Alternat Med* (2014).
- Felder, Christian C, Frank P Bymaster, and John Ward, et al. "Therapeutic opportunities for muscarinic receptors in the central nervous system." *J Med Chem* 43 (2000): 4333-4353.
- Delumeau, JC, D Bentue-Ferrer, and JM Gandon, et al. "Monoamine oxidase inhibitors, cognitive functions and neurodegenerative diseases." In *Amine Oxidases: Funct Dysfunct*, Springer, Vienna, (1994): pp: 259-266.
- Riederer, Peter, and Thomas Müller. "Use of monoamine oxidase inhibitors in chronic neurodegeneration." *Exp Opin Drug Metabol Toxicol* 13 (2017): 233-240.
- dos Santos Pisoni, Diego, Jessie Sobieski da Costa, and Douglas Gamba, et al. "Synthesis and AChE inhibitory activity of new chiral tetrahydroacridine analogues from terpenic cyclanones." *Europ J Med Chem* 45 (2010): 526-535.
- Seniya, Chandrabhan, Ghulam Jilani Khan, and Kuldeep Uchadia. "Identification of potential herbal inhibitor of acetylcholinesterase associated Alzheimer's disorders using molecular docking and molecular dynamics simulation." *Biochem Res Intern* (2014).
- Oboh, Ganiyu, Taiwo M Adewuni, and Ayokunle O Ademosun, et al. "Sorghum stem extract modulates Na⁺/K⁺-ATPase, ecto-5'-nucleotidase, and acetylcholinesterase activities." *Comparat Clin Pathol* 25 (2016): 749-756.
- Taylor, John RN, Peter S Belton, and Trust Beta, et al. "Increasing the utilisation of sorghum, millets and pseudocereals: Developments in the science of their phenolic phytochemicals, biofortification and protein functionality." *J Cereal Sci* 59 (2014): 257-275.
- Ilori, Olukemi O, and Olukemi A Odukoya. "Hibiscus sabdarifa and Sorghum bicolor as natural colorants." *Electron J Environ Agric Food Chem* 4 (2005): 858-862.
- David, Temitope I, Niya S Adedokun, and Olaposi I Omotuyi, et al. "Molecular docking analysis of phyto-constituents from Cannabis sativa with pDHFR." *Bioinform 14* (2018): 574.
- Balogun TA, Omoboyowa DA, and Saibu OA. "In silico anti-malaria activity of quinolone compounds against *Plasmodium falciparum* Dihydrofolate Reductase (pDHFR)." *Internat J Biochem Res Rev* 29 (2020): 10-17.
- Schrödinger, L. "Release 2017-1: Lig Prep." LLC, New York, NY: Schrödinger, USA, (2017).
- Joshi CK, Joshi HK. "Molecular docking analysis of acetylcholinesterase with phyto-compounds from medicinal plants." *Multilogic in Science* (2015), pp: 164-168.
- Pagadala, Nataraj S, Khajamohiddin Syed, and Jack Tuszynski. "Software for molecular docking: a review." *Biophys Rev* 9 (2017): 91-102.
- Srikanth JI, Kavimani S, and Uma M. "Comparative molecular docking study of phytoconstituents identified in *Morinda citrifolia* Linn on acetylcholinesterase and butyrylcholinesterase." *J Chem Pharmaceut Res* 9 (2017): 121-125.
- Wilkinson, Anthony J, Alan R Fersht, and David M Blow, et al. "Site-directed mutagenesis as a probe of enzyme structure and catalysis: tyrosyl-tRNA synthetase cysteine-35 to glycine-35 mutation." *Biochem* 22, no. 15 (1983): 3581-3586.
- Cosmas, S, PE Joshua, and OA Durojaye. "In-silico Structure-Activity Relationship and Molecular Docking Studies of Monosubstituted 6-Gingerol against Streptococcus pneumoniae Phosphomevalonate Kinase." *Internat J Sci Eng Res* 9 (2018): 1733-1742.

How to cite this article: Damilola A Omoboyowa, Toheeb A Balogun, Oluwatosin A Saibu. "Inhibitory Potential of Phyto-Constituents from Sorghum Stem on Neuro-modulatory Enzymes: A Computational Approach". *J Comput Sci Syst Biol* 13 (2020) 13:326