Influence of Initial Seeding Density on Gene Expression during Neuronal Priming

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Abstract

Derivation of neurons from pluripotent stem cells has moved from a niche expertise to a widely used technology, prompting a surge to create novel and timely treatments for neurologic disorders such as Parkinson’s disease. However, current methods for derivation of neurons, though widely available, remain inefficient, resulting in sub-optimal yields and purities of potentially therapeutic cells. Optimising cell processing methods to improve yield and purity is necessary and in this work, we assessed the impact of Initial Cell Seeding Density (ICSD) on differentiation of pluripotent cells (human embryonic and human induced pluripotent stem cells) towards a neuronal specification using a recognised laboratory protocol. We found small but significant differences in expression of specific genes related with pluripotency and neuronal lineage commitment associated with ICSD. By reducing ICSD, it may be possible to improve yield from less start material.

Keywords: Neuronal differentiation; Initial cell seeding density; Optimization; Gene expression; Translation; Pluripotent stem cells

Introduction

Regenerative medicine holds promise to treat many currently unmet clinical indications and has seen major change in the last decade, with expectations of the science only increasing with each new discovery and clinical trial undertaken. With these advancements stem cell research has grown from being driven by biological discovery to developing and optimizing processes and protocols to enable their application in the therapeutic arena. As cell therapy process technology develops and researchers look to transplant specific cell types generated using in-vitro differentiation protocols, it will become more and more critical to ensure process efficiencies are optimized in order to achieve the scale of production needed to meet the clinical demand.

In order to successfully address the challenges of cell therapy manufacture at the industry scale, it is important to ensure all stages of the whole bioprocess are optimised first at the bench scale. The very first stage of many expansion or differentiation processes involves plating the cells onto an appropriate substrate for attachment and growth. The substrate is critical, so for example human pluripotent cells are often plated onto Mouse Embryonic Fibroblast (MEF) feeders [1,2] or are specially adapted for survival and growth on acellular material such as matrigel [3]. In addition to substrate, Initial Cell Seeding Density (ICSD) can have a significant impact on cell behaviour and is reported to alter proliferation and differentiation responses of various adult cells during 2D culture [4] and in 3D culture for tissue engineering purposes [5]. In the case of pluripotent cells, passing hESC colonies at lower density of 1:8 resulted in a high degree of spontaneous differentiation versus those passed at high density of 1:3 [6]. Using mouse 46C ESCs, Veraitch et al. demonstrated that lower seeding density specifically enhanced neuroectoderm differentiation, identified by enhanced Sox1 expression [7].

ICSD is important for a second reason too. Scalable expansion of therapeutic cell lines to achieve necessary clinical quantities is still fraught with difficulty [6] with adherent and suspension culture methods currently being investigated [8,9]. This is not just the case for primary somatic cell populations, where expansion is hampered by limited replicative potential and biological changes accompanying cell aging [10], but also for pluripotent cell lines that are prone to karyotypic abnormalities with extended periods in culture [11,12].

Attempts to reduce the start material needed to produce a cellular therapeutic will be of substantial benefit to the industry. Therefore, the aim of this study was to determine the impact of ICSD on expression of key genes during neuronal induction of human pluripotent cell lines. We based our work on an established protocols, which use SMAD signalling inhibition along with dopaminergic factors to induce neuronal differentiation [13,14] and mapped the gene expression profile associated with the shift away from a pluripotent state, during neuronal priming and induction of a specialised phenotype over the first 11 days of differentiation.

Methods

Human pluripotent cell lines

Two cell lines were used to assess the effect of ICSD on neuronal priming. Two cell lines were used to assess the expression of neuronal markers, early neuronal and mature markers as well as specialized dopaminergic markers: the human induced pluripotent stem cell (hiPSC) line, MSUH001 (a kind gift from Dr Cibelli, Cellular Reprogramming Laboratory, Michigan State University); and the human embryonic stem cell (hESC) line, Shef6 obtained from UK Stem Cell Bank. Both hESC and hiPSC pluripotent cell lines (Figure 1A and 1B) were cultured on Mitomycin-C gift from Dr Cibelli, Cellular Reprogramming Laboratory, Michigan State University; and the human embryonic stem cell (hESC) line, Shef6 obtained from UK Stem Cell Bank. Both hESC and hiPSC pluripotent cell lines (Figure 1A and 1B) were cultured on Matrigel [3]. In addition to substrate, Initial Cell Seeding Density (ICSD) can have a significant impact on cell behaviour and is reported to alter proliferation and differentiation responses of various adult cells during 2D culture [4] and in 3D culture for tissue engineering purposes [5]. In the case of pluripotent cells, passing hESC colonies at lower density of 1:8 resulted in a high degree of spontaneous differentiation versus those passed at high density of 1:3 [6]. Using mouse 46C ESCs, Veraitch et al. demonstrated that lower seeding density specifically enhanced neuroectoderm differentiation, identified by enhanced Sox1 expression [7].

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10% KO-Serum replacement, 1% Glutamax, 1% NEAA and 100mM β-mercaptoethanol (All Invitrogen, Paisley, UK) supplemented with basic Fibroblast Growth Factor (bFGF, 4 ng/mL, Prospe-Tany technogene Ltd. East Brunswick, USA). Cells were mechanically passaged twice weekly and maintained at 37°C, 5% (v/v) CO2 prior to initiation of the protocol.

**Neuronal priming**

Priming briefly consisted of the cells being expanded in T25 flasks (Nunc, VWR International Ltd, Leicestershire, UK) in KSRM supplemented with 4 ng/mL bFGF and cultured until colonies compacted. Upon commencement of priming, cells were dissociated into single cell suspension using Tryple Express (Invitrogen) and passed through a cell strainer (40 µm, BD Falcon, Oxford, UK) to ensure a single cell suspension. Cells were resuspended in filter sterilised MEF-conditioned media (CM media, KSRM supplemented with 4 ng/mL bFGF incubated with inactivated MEFs for 3 days, removed and filter sterilised) supplemented with bFGF (10 ng/mL) and ROCK-inhibitor (10 µM, Y27632, Stemolecule, Miltenyi Biotec, Surrey, UK) (complete CM media), then plated onto gelatin-coated 6 well plates (Nunc) and incubated at 37°C for 1 hour. Non-adhered cells were removed and the plates rinsed gently with fresh media to remove any remaining non-attached pluripotent cells. Feeders remained adhered to the gelatin-coated surface. The pluripotent cell suspensions were centrifuged at 1,200 rpm for 3 minutes and resuspended in complete CM media, then counted using a haemocytometer. Human pluripotent cell populations attached pluripotent cells. Feeders remained adhered to the gelatin-coated surface. The pluripotent cell suspensions were centrifuged at 1,200 rpm for 3 minutes and resuspended in complete CM media, then counted using a haemocytometer. Human pluripotent cell populations

**Figure 1:** A Phase contrast image of IPS colony (magnification × 10, scale bar = 400 µm) and ICC for Oct4 pluripotency marker. B Graph representing flow cytometry data for hESC and hiPSC culture expression of pluripotent surface markers SSEA4 (n=3) and TRA-1-60 (n=2). Error bars represent SD ± 1. C Diagram demonstrating the stages of the priming protocol. D Growth profile of the hES cell line over 7 days of culture expansion. E Growth profile of the IPS cell line over 7 days of expansion (Bars indicated Mean ± SEM).

On day 3 of the population expansion when three clear confluence densities were observed across the investigated ICSD, the media was changed to initial differentiation media, KSRM supplemented with noggin (500 ng/mL, R&D Systems, Abington, UK) and TGF-β inhibitor (10 µM, SB431542, Miltenyi Biotec ). Day 1 media was supplemented with Sonic Hedgehog (100 ng/mL SHH) (R&D Systems) and Purmorphamine (2 µM, Miltenyi Biotec). Media was maintained until day 3, when CHIR99021 (3 µM, Miltenyi Biotec) was added. After an additional 2 days, the media was modified to contain 25% N2, neuronal supplement (PAA, Somerset, UK) in addition to factors: Noggin, SHH, Purmor and CHIR, but excluding SB431542. N2 concentration was incrementally increased over the following 6 days (to day 11) through 25%, 50% and 75% of the media constituent.

**Confluence analysis**

Analysis of confluence was performed using a phase contrast microscope (Nikon, Eclipse TI-E) with an inbuilt automated platform. 20 random images (10 × magnifications) were taken of each duplicate well. Images were processed using the PHANTAST software [15]. Briefly, cell regions on phase contrast microscopy images are characterised by high variations in intensity as opposed to image background regions that are mostly homogeneous. These regions of high variability were detected using a local contrast filter. Image confluence was computed as the ratio of the number of cell pixels to the total number of pixels. Images of biological duplicates were processed and averages were taken
of all readings for a given ICSD. Readings were taken over a time course up to 7 days, to ascertain the growth profile of the two cell lines and to determine the appropriate time point to commence priming the cells.

Quantitative Polymerase Chain Reaction

Cells were mechanically harvested from the culture well and processed for total RNA extraction using QIAzol Lysis Reagent (Qiagen, Manchester, UK). Total RNA content and purity was then quantified using a NanoDrop 1000 spectrophotometer (ThermoFisher Scientific, Warrington, UK). Reverse transcription was achieved using QuantiTect Reverse transcription Kit (Qiagen, Manchester, UK) and gene expression analysis was performed using QuantiFast SYBR Green PCR Kit (Qiagen) according to the manufacturer's instructions. The QuantiTech primers (all Qiagen) used were: housekeeper genes ubiquitin C (UBC-1-SG), gapdh & β-actin; pluripotency markers, nanog, sox2 & oct4; epiblast marker, fgf5; neuronal markers, otx2, pax6 & sox1; and dopaminergic markers, TH and Lmx1a. Expression data was normalised against three housekeeping genes (β-Actin, GAPDH and UBC-1-SG) and calculated relative to day 0. All samples were processed in triplicate.

qPCR data was collected using the BioRad CFX Connect Real-Time System (Bio-Rad, Hertfordshire, UK) and was categorised and processed using CFX Manager Data analysis software (Bio-Rad). Samples were categorised using a number of criteria: biological set (biological replicate number); cell type; technical replicate; target gene and sample name. Relative expression (ΔΔCq) was calculated using the BioRad Gene study programme, normalising target gene expression against housekeeping gene levels. Significance was found when p<0.05. Data is shown against a logarithmic scale (log10).

Results

To investigate the effect of ICSD on the expression of target genes through a neuronal priming protocol on pluripotent stem cell populations (Figure 1A-1C) we initially investigated the growth profiles of two human pluripotent cell lines at 3 different ICSDs over a period of 7 days. It was found that both cell lines reached confluence by 6 days at all ICSDs tested (Figure 1D and 1E). In line with expectation, at earlier time points lower ICSD resulted in lower confluence compared with cells seeded at higher ICSD. Therefore, subsequent neuronal differentiation experiments were commenced from day 3 when differences in confluence could be identified.

Expression levels of target genes were assessed at day 0, 5 and 11 following induction of neuronal differentiation. In the case of pluripotency genes, nanog were significantly down-regulated (p<0.01) at day 5 and 11 following induction for all ICSDs in hESCs and hiPSCs (Figure 2A and 2D). In both cell lines sox2 expression increased, though only marginally (Figure 2C and 2F). In terms of differences between ICSDs, only a very marginal increase in nanog down-regulation was seen for 10 k/cm² and 18 k/cm² versus 25 k/cm² ICSD (p<0.05) in hESCs (Figure 2A) and conversely small increases in sox2 up-regulation were seen at 25 k/cm² versus 10 k/cm² for hESCs (p<0.05). Epiblast marker fgf5 underwent moderate time-dependent up-regulation over the 11 day neuronal induction protocol for only 25 k/cm² for both hESCs and hiPSCs (Figure 3A and 3B). Notably, differences in gene expression between different ICSDs were observed, with a consistent trend of increased fgf5 expression at 25 k/cm² versus 10 k/cm² (at day 5 for hiPSCs (p<0.05) and at day 11 for both hESCs (p<0.01) and hiPSCs (p<0.05)).

Next, expression of neuronal genes was assessed to determine whether changes in population-wide gene expression were evident at different ICSDs. There was a positive increase in expression of neuronal markers sox1, pax6 and otx2 at days 5 and 11 versus day 0 (Figure 4A-4F). Very small, but significant increases in expression of sox1 (Figure 4A) and otx2 (Figure 4C) were seen in hESCs at day 11 under the lower seeding density of 10 k/cm² versus 25 k/cm² ICSD (p<0.05). Similar differences were not seen for hiPSCs. Furthermore, a general increase in gene expression was seen from day 5 to day 11 for hESCs (sox1 and pax6 p<0.01, otx2 p<0.05) but this was not the case for hiPSCs. In fact, a significant decrease in expression of otx2 (p<0.01) was observed from day 5 to day 11.
Discussion

Multiple bioprocess parameters can impact on the responses of human cells during processing [7]. ICSD and degree of confluence at the point of initiation of differentiation protocols should also be considered. In this study, three different ICSDs were studied and the time at which neuronal priming was initiated was determined according to the point at which three different levels of in-culture density were seen. At the start of the neuronal priming protocol, SMAD inhibitors were supplemented to the differentiation media [13,14] and were observed to have no effect on population viability.

Over the course of the priming protocol pluripotency markers and markers of the neuronal lineage were assessed. It was found that consistent, significant drops in pluripotency markers OCT4 and NANOG occurred for both hESC and hiPSC cultures (Figure 2). This would indicate that the cultures shifted from expressing genes associated with maintained pluripotency towards a differentiated state. Moreover, it was accompanied by a concurrent increase in expression of FGFR (Figure 3), indicating differentiation to the late epiblast stage and early ectoderm phase [16]. Increased expression of SOX2 (Figure 2) is also associated with acquisition of a neuronal progenitor phenotype [17] and further supports a shift away from pluripotency towards the ectodermal/neuronal lineage. The higher seeding density of 25 k/cm² led to higher population-wide expression of FGFR, which may seemingly suggest that a higher ICSD results in a more efficient differentiation for both hESCs and hiPSCs. However, combined with the increased expression of later neuronal markers at lower ICSD, it is possible that in the higher ICSD investigated, a higher proportion of the population stay in this late epiblast/early ectoderm stage, whereas at 10 k/cm², stagnated expression of FGFR possibly indicates differentiation past the epiblast stage. The suggestion that lower FGFR expression in hESCs at 10 k/cm² indicates a move down the ectoderm lineage is enforced by the significant difference in SOX2 expression observed at day 11 (Figure 4A). This trend was repeated for OTX2 at 10 k/cm² where higher expression was noted compared to the other ICSDs (Figure 4C), possibly indicating more rapid differentiation towards mature neuronal states at lower ICSD. This supports previous observations in mouse ESCs, where increased neurectodermal differentiation was noted at low ICSD [7]. Expression of neuronal genes by hiPSCs was not as prominent as those observed for hESCs, with relative expression of the majority of genes screened never showing greater than 1×10²ΔΔCq increase. This may indicate a reluctance of the cell type to move down the ectoderm lineage. In the case of hiPSCs, the reduction in OTX2 expression over time (Figure 4F) would suggest that differentiation is not toward a mid-neural tube phenotype. This assumption in hiPSCs is backed up by the expression profile of TH and LMX1A (Figure 5C, 5D) with TH showing significantly reduced expression at both 18 and 25 k/cm² and LMX1A expression showing a significant drop across all ICSDs within the hiPSC populations. In the case of hESCs, the trend for TH and LMX1A (Figure 5A, 5B) was similar to that found for the other neuronal genes; the lowest ICSD of 10 k/cm² led to significantly greater expression of TH and LMX1A than the higher ICSDs. The highest ICSDs showed
the only significant drop in lmx1a (Figure 5B) expression observed in hESCs, potentially reiterating that lower ICSD and level of confluence at initiation of differentiation can improve the efficiency of lineage specification needed to drive towards a dopaminergic neuronal cell phenotype.

Conclusion

In the current study, low ICSD resulted in overall improved neuronal differentiation in hESCs, with greater expression of TH and lmx1a indicating a potential move towards a dopaminergic lineage. hESCs had a greater propensity to differentiate towards an ectoderm cell type than MSUH001 hiPSCs, when assessing population-wide gene expression. Even though substantial differences are evident between different pluripotent cell lines, this data still provides evidence that ICSD can affect target gene expression and hence differentiation during neuronal lineage commitment.

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References