

Journal of Molecular Biomarkers & Diagnosis

Review Article

Open Access

Inflammatory Breast Cancer Stem Cells: Contributors to Aggressiveness, Metastatic Spread and Dormancy

Cynthia M. van Golen¹ and Kenneth L.van Golen²⁴

¹Department of Biological Science, Delaware State University, USA

²Department of Biological Science, University of Delaware and The Helen F. Graham Cancer Center, USA

Abstract

Cancer stem cell populations have been identified for several types of cancers and suggest a way for tumor cells to be resistant to therapies. Further, because of the longevity, endurance and replicative potential of cancer cells with stem-like properties, other malignant attributes such as recurrence after long periods of dormancy can also be explained. Inflammatory breast cancer (IBC) is a unique and aggressive form of breast cancer that has a clinical course unlike other forms of breast cancer. The main hallmark of IBC is prolific invasion of the dermal lymphatic vessels by tumor emboli leading to rapid metastasis of the disease. Despite an extremely aggressive treatment approach, the majority of women with IBC present with disease recurrence suggesting the presence of chemo resistant and/or dormant breast cancer cells. Current evidence suggests that IBC tumor emboli contain distinct populations of cells with stem cell-like properties. Thus, specific targeting of these stem cell-like cancer cells may be the key to effectively treating IBC.

Keywords: Inflammatory breast cancer; Breast cancer stem cells; Dormancy; Metastasis

Introduction to Inflammatory Breast Cancer

Inflammatory breast cancer (IBC) is arguably the most aggressive and persistent form of breast cancer known with 5- and 10-year disease-free survival rates of <42% and <20%, respectively [1-4]. IBC is extremely unique in its manifestation, presentation and molecular pathology [5-7]. This form of breast cancer affects younger women with a mean age of ~ 46 years old and is reported in girls as young as 12 years old [8-10].

The term inflammatory breast cancer was first coined by Drs. Lee and Tannenbaum in 1924 to unite what was thought to be a number of individual cancers that affected younger women, under a single term [11]. However, the term "inflammatory" breast cancer is a misnomer, as the tumor lacks the conspicuous presence of immune effector cells [8,12]. The appearance of the disease does resemble that of an infection with a number of primary skin changes that occur including redness, thickening of the skin, erythema, peau d' orange and warmth to the touch [7,8,11,12]. In addition, the breast size increases and can be sore, the nipple can invert and if advanced, the breast can become necrotic [7,8,11,12]. Patients report that the onset of these symptoms can occur within a very short period of time 24-48 hours.

A palpable mass is often missing in IBC. Instead the primary tumor presents as diffuse cords or sheets in the breast [7,8,11,12]. IBC is highly lymph invasive and the main hallmark of the disease is lympho vascular invasion of the dermal lymphatic vessels of the skin overlying the breast [7,13,14]. It is suggested that the presence of the tumor emboli within the dermal lymphatic vessels cause edema and the skin changes described above. However, physicians report a persistence of symptoms after treatment and apparent complete pathological response [15]. Unfortunately, due to the nature of these symptoms coupled with the rapidity of onset, IBC is often misdiagnosed as an infection and inappropriately treated [15,16]. IBC experts estimate that nearly 90% of IBC cases are initially misdiagnosed.

It is also suggested that the presence of tumor emboli in the dermal

lymphatic vessels are responsible for the prolific metastasis associated with IBC. By definition, IBC is a T4d tumor and is always diagnosed as a stage IIIb or IV tumor, with all women having lymph node involvement and nearly 1/3 having gross distant metastases in visceral organ, bone and brain [2,7,8,17]. The progression to advanced disease is extremely rapid, progressing from what appears to be normal breast to advanced disease within 6 months [7].

As stated above, IBC is molecularly distinct from other forms of breast cancer [5]. Our laboratory was the first to identify genes uniquely altered in IBC, particularly RhoC GTPase, which is over expressed in over 90% of IBC patient specimens [19]. We demonstrated that active RhoC GTPase is required to drive the invasive IBC phenotype through reorganization of the actin cytoskeleton [20,21]. Extensive analysis has shown that IBC expresses a unique gene profile compared to cell-type of origin matched, stage matched or receptor status (eg. estrogen receptor/progesterone receptor (ER/PR) +, Her2+ or triple negative) non-IBCs [14,22]. Interestingly, many gene changes that are observed during progression of non-IBC are opposite for IBC [5,6]. For example, E-cadherin and caveolin-1 and -2 are over expressed in IBC compared to non-IBC [13,26,27].

In addition to the unique molecular signature and route of dissemination, IBC tends to be resistant to therapy [3,28]. Recurrences, including chest wall recurrences, are common and often occur in soft tissue and visceral organ [29]. Because of this, many people believe that IBC emboli contain a large number of breast cancer cells with stem cell-like properties. The SUM149 IBC cell line, emboli from the Mary-X

*Corresponding author: Kenneth L. van Golen, 320 Wolf Hall, Department of Biological Science, The University of Delaware, Newark, DE 19716, USA, Tel: 302-831-2669; Fax:302-831-2281; E-mail: klvg@udel.edu

Received May 21, 2012; Accepted June 20, 2012; Published June 25, 2012

Citation: van Golen CM, van Golen KL (2012) Inflammatory Breast Cancer Stem Cells: Contributors to Aggressiveness, Metastatic Spread and Dormancy. J Mol Biomarkers Diagn S8:002. doi:10.4172/2155-9929.S8-002

Copyright: © 2012 van Golen CM, et al. This is an open-access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited

xenograft model and IBC patient emboli are shown to contain a large population of cancer cells with a stem cell signature [30,31].

Cancer Stem Cells in Inflammatory Breast Cancer

Cancer stem cells became a wide spread interest with the identification of putative stem cells in leukemia [32]. Since that time, cancer stem cells have been identified for multiple hematologic and solid tumors, including breast, prostate, brain, skin, liver, and lung tumors [33]. As with all stem cells, tumor stem cells have prolonged self-renewal capacity and the potential to produce progeny that will differentiate into specialized cells of the tissue of origin [34]. The original determination of a cancer stem cell population was based upon the ability of those cells to reform a tumor *in vivo*, usually experimentally determined through serial injection into immune compromised experimentally [34,35]. Based upon these criterion, tumor stem cells are also known as tumor-initiating cells [33]. However, cancer stem cells are also closely linked to more advanced tumor phenotypes, including angiogenesis, chemo resistance, and metastases [33,36,37].

Several markers have been identified in cancer stem cells, including those from breast carcinoma. CD44 is considered a stem cell marker for several tumors, including bladder, breast, colon, and head and neck. CD133 is another common marker, found in stem cells from colon carcinoma, Ewing's sarcoma, and pancreatic cancer. In breast carcinomas, the absence of CD24 is also indicative of a stem cell phenotype [34,38]. In addition to these markers, signal transduction pathways involved in the breast cancer stem cell phenotype have been elucidated. Notch signaling, involved in differentiation and cell fate determination during development, is up-regulated in breast cancer stem cells and correlates with the CD44+/CD24- marker phenotype [39]. Furthermore, inhibition of Notch signaling abrogates cancer stem cell self-renewal [39]. Tumor necrosis factor alpha (TNF), acting through the nuclear factor kappa B (NF-kB) signaling pathway, upregulates SLUG, a mediator of the epithelial to mesenchymal transition in breast cancer cells. SLUG up-regulation in turn promotes a stem cell phenotype within breast cancer cells [40]. Additional pathways implicated in the breast cancer stem cell phenotype include those of aldehyde dehydrogenase [41], bone morphogenic protein (BMP), Wnt [42] and ganglioside GD2 [43].

As previously discussed, IBCs are highly aggressive and metastatic at diagnosis. Given the aggressive nature of this tumor, it is not surprising that experimental evidence supports the involvement of breast cancer stem cells in its progression. Tumor cells isolated from the MARY-X model of human IBC express both embryonal stem cell markers (Nestin, Rex1, and Stellar) and the classic breast cancer stem cell signature (CD44+/CD24-/CD133+/aldehyde dehydrogenase1 (ALDH1)+) [31]. Furthermore, 74% of human IBC samples contain a genetic signature indicative of a high cancer stem cell composition, as opposed to 44% of non-IBC samples [42]. Stem cell-related signaling, particularly through Hedgehog and transforming growth factor beta (TGF β), is also more active in IBC vs. non-IBC samples [42]. Nestin, a protein primarily associated with neural stem cells, is highly expressed in both triple negative and IBC tumors [44]. As with other types of breast tumors, ALDH1 is highly up-regulated in IBC, and its expression correlates with increased metastasis and decreased survival [30]. Finally, IBCs express a unique microRNA (miRNA) profile, and several of these miRNAs are also expressed in tumor stem cells [45]. Therefore, the presence of high stem cell content in IBC may contribute to the aggressive nature of this disease, but may also offer attractive therapeutic targets.

Farnesyl Transferase Inhibitors as Potential Cancer Stem Cell-Targeted IBC Therapeutics

Farnesyl transferase inhibitors (FTIs) were originally developed to target Ras-based tumors [46]. However, their performance in the clinic was disappointing thus, attention turned to non-Ras targets, specifically Rho GTPases [47,48]. In a study of the effects of FTIs on the SUM149 IBC cell line, we observed a significant, but reversible, loss of RhoC GTPase-mediated motility and invasion [49]. These results strongly suggest that FTIs could be used as novel adjuvant therapy for IBC [49].

Interestingly, FTI-treated IBC cells exhibited a unique spread and flattened morphology with pronounced actin stress fibers [49]. This unique morphology is reminiscent of what is described for an *in vitro* model of breast cancer cell dormancy [51]. Interestingly in this model, RhoA GTPase activity was decreased [52,53]. RhoA and RhoC GTPases tended to have opposing actions in tumor cells, exhibiting a balance between inactive and active forms of the two GTPases [54,55].

The mechanisms of tumor cell dormancy are currently not well understood. In a recent study from our laboratory we demonstrated that treatment of MCF-7 non-IBC cells with FTI L-422,831, leads to a phenotype reminiscent of dormancy [56,57]. Further, FTI treatment of the MCF-7 cell line leads to profound changes in Rho GTPase activation [56,57]. Specifically, RhoA GTPase becomes hypoactivated while RhoC GTPase becomes hyperactivated, producing radical changes in the cell cytoskeleton and cellular morphology identical to what was observed in the FTI-treated SUM149 IBC cells [49].

Similar to what is observed for the *in vitro* model [51,52], FTIinduced dormancy is reversible [49,56,58]. Upon FTI withdrawal cells grow normally after exiting from nearly two weeks of dormancy [56]. FTI-treated cells have minimal metabolic activity and undergo autophagy [56]. Autophagy is the process where a cell degrades organelles such as mitochondria to expend less energy avoiding apoptosis [59]. Autophagy is regulated through the extracellular matrix and is suggested to be required for dormancy [60-62]. In addition, activation of the c-jun NH2 terminal kinase (JNK/SAPK) signaling pathway occurs during autophagy [63,64]. Increased RhoC GTPase activation during FTI treatment increases JNK/SAPK signaling leading to breast tumor cell dormancy [56].

Stem cell associated markers such as ALDH1 and CD44 are shown to be expressed by a subpopulation of cancer cells in both tumors and cells lines [65-67]. It is suggested that breast cancer cells with stem cell-like properties are responsible for metastatic spread [68,69], while RhoC GTPase is expressed by highly metastatic cancer cells that exhibit "stemness" properties [30,70]. Cancer stem cells have also been linked to dormancy. It is thought that a metastatic stem cell, arriving in a non-conducive microenvironment undergo prolonged dormancy until an event such as a cytokine storm or extracellular matrix remodeling re-activate cancer cells [68]. Breast cancer cells undergoing post-FTItreatment dormancy, express ALDH1 and CD44, suggesting that breast cancer stem cells are susceptible to FTI-induced dormancy [57]. This phenotype is due to the activation of JNK/SAPK signaling resulting from increased RhoC activation. The activation of JNK/SAPK signaling in turn may lead to induction of autophagy allowing the breast cancer cells to remain inactive. These studies may have profound implications for the use of FTIs in the clinic. Potentially, FTI-induced dormancy could synchronize tumor cells; with FTI withdrawal would allow growth making the cells more susceptible to chemotherapeutics.

Citation: van Golen CM, van Golen KL (2012) Inflammatory Breast Cancer Stem Cells: Contributors to Aggressiveness, Metastatic Spread and Dormancy. J Mol Biomarkers Diagn S8:002. doi:10.4172/2155-9929.S8-002

Conclusions

The presence of cancer stem cells in epithelial tumors, including IBCs, is now widely accepted, and laboratories are now therapeutically targeting several of the proteins associated with stem cell phenotypes. Breast cancer stem cells are highly resistant to traditional radio- and chemotherapy. ALDH1 can metabolize chemotherapeutic drugs, and CD44+/CD24- cells are enriched in remaining breast tumor tissue following chemotherapy [71]. Targeting HER2, a receptor thought to aid in the stem cell phenotype, may decrease this particular population of cells [69,70]. Small hairpin (sh) RNA targeting CD44 induces breast cancer stem cells to differentiate, resulting in a lower tumorigenic potential [70]. This putatively could reintroduce chemo-susceptibility in these breast cancer cells. In melanoma, antibodies targeting CD20 are effective at reducing the stem cell population and causing metastases to regress [71]. Therefore, antibodies or specific inhibitory agents targeting CD44, ALDH1, or other stem cell-like markers in IBC could be beneficial at reducing the aggressive progression of this disease.

References

- Bristol IJ, Woodward WA, Strom EA, Cristofanilli M, Domain D, et al. (2008) Locoregional treatment outcomes after multimodality management of inflammatory breast cancer. Int J Radiat Oncol Biol Phys 72: 474-484.
- Lerebours F, Bieche I, Lidereau R (2005) Update on inflammatory breast cancer. Breast Cancer Res 7: 52-58.
- Harris EE, Schultz D, Bertsch H, Fox K, Glick J, et al. (2003) Ten-year outcome after combined modality therapy for inflammatory breast cancer. Int J Radiat Oncol Biol Phys 55: 1200-1208.
- Gonzalez-Angulo AM, Hennessy BT, Broglio K, Meric-Bernstam F, Cristofanilli M, et al. (2007) Trends for inflammatory breast cancer: is survival improving? Oncologist 12: 904-912.
- Joglekar M, KL van Golen (2012) Molecules That Drive the Invasion and Metastasis of Inflammatory Breast Cancer. Inflammatory Breast Cancer: An Update 161-184.
- Radunsky GS, van Golen KL (2005) The current understanding of the molecular determinants of inflammatory breast cancer metastasis. Clin Exp Metastasis 22: 615-620.
- Kleer CG, van Golen KL, Merajver SD (2000) Molecular biology of breast cancer metastasis. Inflammatory breast cancer: clinical syndrome and molecular determinants. Breast Cancer Res 2: 423-429.
- Jaiyesimi IA, Buzdar AU, Hortobagyi G (1992) Inflammatory breast cancer: a review. J Clin Oncol 10: 1014-1024.
- 9. Tabbane F, el May A, Hachiche M, Bahi J, Jaziri M, et al. (1985) Breast cancer in women under 30 years of age. Breast Cancer Res Treat 6: 137-144.
- Nichini FM, Goldman L, Lapayowker MS, Levy WM, Maier W, et al. (1972) Inflammatory carcinoma of the breast in a 12-year-old girl. Arch Surg 105: 505-508.
- Lee BT (1924) Inflammatory carcinoma of the breast: a report of twenty-eight cases from the breast clinic of Memorial Hospital. Surg Gynecol Obstet 39:580-595.
- Haupt HM, Hood AF, Cohen MH (1984) Inflammatory melanoma. J Am Acad Dermatol 10: 52-55.
- 13. Kleer CG, van Golen KL, Braun T, Merajver SD (2001) Persistent E-cadherin expression in inflammatory breast cancer. Mod Pathol 14: 458-464.
- Tomlinson JS, Alpaugh ML, Barsky SH (2001) An intact overexpressed E-cadherin/alpha,beta-catenin axis characterizes the lymphovascular emboli of inflammatory breast carcinoma. Cancer Res 61: 5231-5241.
- Dawood S, Valero V (2012) Clinical Aspects of Inflammatory Breast Cancer: Diagnosis, Criteria, Controversy. In: Ueno NT, Cristofanilli M, editors. Inlfammatory Breast Cancer: An Update. New York, N.Y Springer 11-20.
- Dawood S, Ueno NT, Valero V, Woodward WA, Buchholz TA, et al. (2012) Identifying factors that impact survival among women with inflammatory breast cancer. Ann Oncol 23: 870-875.

- 17. Woodward WA, Cristofanilli M (2009) Inflammatory breast cancer. Semin Radiat Oncol 19: 256-265.
- 18. van Golen KL, Davies S, Wu ZF, Wang Y, Bucana CD, et al. (1999) A novel putative low-affinity insulin-like growth factor-binding protein, LIBC (lost in inflammatory breast cancer), and RhoC GTPase correlate with the inflammatory breast cancer phenotype. Clin Cancer Res 5: 2511-2519.
- van Golen KL, Bao LW, Pan Q, Miller FR, Wu ZF, et al. (2002) Mitogen activated protein kinase pathway is involved in RhoC GTPase induced motility, invasion and angiogenesis in inflammatory breast cancer. Clin Exp Metastasis 19: 301-311.
- Van Golen KL, Wu ZF, Qiao XT, Bao LW, Merajver SD (2000) RhoC GTPase, a novel transforming oncogene for human mammary epithelial cells that partially recapitulates the inflammatory breast cancer phenotype. Cancer Res 60: 5832-5838.
- 21. Van Laere SU, NT Finetti, P Vermeulen, PB, Lucci, A Birnbaum, D Robertson, FM van Dam, PA Woodward, WA Viens P Dirix, LY; Reuben, JM; Iwamoto, T; Cristofanilli, M; Bertucci, F (2011) An Integrated Analysis of Three Distinct IBC/nIBC Affymetrix Gene Expression Data Sets Further Unveils the Molecular Biology of IBC. San Antonio Breast Cancer Symposium. San Antonio.
- 22. Van Laere S, Van der Auwera I, Van den Eynden G, Van Hummelen P, van Dam P, et al. (2007) Distinct molecular phenotype of inflammatory breast cancer compared to non-inflammatory breast cancer using Affymetrix-based genome-wide gene-expression analysis. Br J Cancer 97: 1165-1174.
- Van Laere S, Van der Auwera I, Van den Eynden GG, Fox SB, Bianchi F, et al. (2005) Distinct molecular signature of inflammatory breast cancer by cDNA microarray analysis. Breast Cancer Res Treat 93: 237-246.
- 24. Van den Eynden GG, Van der Auwera I, Van Laere S, Colpaert CG, van Dam P, et al. (2004) Validation of a tissue microarray to study differential protein expression in inflammatory and non-inflammatory breast cancer. Breast Cancer Res Treat 85: 13-22.
- 25. Van den Eynden GG, Van Laere SJ, Van der Auwera I, Merajver SD, Van Marck EA, et al. (2006) Overexpression of caveolin-1 and -2 in cell lines and in human samples of inflammatory breast cancer. Breast Cancer Res Treat 95: 219-228.
- Gonzalez-Angulo AM, Morales-Vasquez F, Hortobagyi GN (2007) Overview of resistance to systemic therapy in patients with breast cancer. Adv Exp Med Biol 608: 1-22.
- Cristofanilli M, Valero V, Buzdar AU, Kau SW, Broglio KR, et al. (2007) Inflammatory breast cancer (IBC) and patterns of recurrence: understanding the biology of a unique disease. Cancer 110: 1436-1444.
- Charafe-Jauffret E, Ginestier C, Iovino F, Tarpin C, Diebel M, et al. (2010) Aldehyde dehydrogenase 1-positive cancer stem cells mediate metastasis and poor clinical outcome in inflammatory breast cancer. Clin Cancer Res 16: 45-55.
- Xiao Y, Ye Y, Yearsley K, Jones S, Barsky SH (2008) The lymphovascular embolus of inflammatory breast cancer expresses a stem cell-like phenotype. Am J Pathol 173: 561-574.
- Bonnet D, Dick JE (1997) Human acute myeloid leukemia is organized as a hierarchy that originates from a primitive hematopoietic cell. Nat Med 3: 730-737.
- 31. Zhao Y, Bao Q, Renner A, Camaj P, Eichhorn M, et al. (2011) Cancer stem cells and angiogenesis. Int J Dev Biol 55: 477-482.
- Frank NY, Schatton T, Frank MH (2010) The therapeutic promise of the cancer stem cell concept. J Clin Invest 120: 41-50.
- Vermeulen L, Sprick MR, Kemper K, Stassi G, Medema JP (2008) Cancer stem cells--old concepts, new insights. Cell Death Differ 15: 947-958.
- Eyler CE, Rich JN (2008) Survival of the fittest: cancer stem cells in therapeutic resistance and angiogenesis. J Clin Oncol 26: 2839-2845.
- 35. Li F, Tiede B, Massagué J, Kang Y (2007) Beyond tumorigenesis: cancer stem cells in metastasis. Cell Res 17: 3-14.
- Al-Hajj M, Wicha MS, Benito-Hernandez A, Morrison SJ, Clarke MF (2003) Prospective identification of tumorigenic breast cancer cells. Proc Natl Acad Sci U S A 100: 3983-3988.
- Guo S, Liu M, Gonzalez-Perez RR (2011) Role of Notch and its oncogenic signaling crosstalk in breast cancer. Biochim Biophys Acta 1815: 197-213.

Citation: van Golen CM, van Golen KL (2012) Inflammatory Breast Cancer Stem Cells: Contributors to Aggressiveness, Metastatic Spread and Dormancy. J Mol Biomarkers Diagn S8:002. doi:10.4172/2155-9929.S8-002

- 38. Storci G, Sansone P, Mari S, D'Uva G, Tavolari S, et al. (2010) TNFalpha up-regulates SLUG via the NF-kappaB/HIF1alpha axis, which imparts breast cancer cells with a stem cell-like phenotype. J Cell Physiol 225: 682-691.
- 39. Ali HR, Dawson SJ, Blows FM, Provenzano E, Pharoah PD, et al. (2011) Cancer stem cell markers in breast cancer: pathological, clinical and prognostic significance. Breast cancer research 13: R118.
- 40. Van Laere S, Limame R, Van Marck EA, Vermeulen PB, Dirix LY (2010) Is there a role for mammary stem cells in inflammatory breast carcinoma?: a review of evidence from cell line, animal model, and human tissue sample experiments. Cancer 116: 2794-2805.
- Battula VL, Shi Y, Evans KW, Wang RY, Spaeth EL, et al. (2012) Ganglioside GD2 identifies breast cancer stem cells and promotes tumorigenesis. J Clin Invest 122: 2066-2078.
- 42. Piras F, Ionta MT, Lai S, Perra MT, Atzori F, et al. (2011) Nestin expression associates with poor prognosis and triple negative phenotype in locally advanced (T4) breast cancer. Eur J Histochem 55: e39.
- 43. Wu X, Somlo G, Yu Y, Palomares MR, Li AX, et al. (2012) De novo sequencing of circulating miRNAs identifies novel markers predicting clinical outcome of locally advanced breast cancer. J Transl Med 10: 42.
- Gibbs JB, Oliff A, Kohl NE (1994) Farnesyltransferase inhibitors: Ras research yields a potential cancer therapeutic. Cell 77: 175-178.
- 45. Lebowitz PF, Prendergast GC (1998) Non-Ras targets of farnesyltransferase inhibitors: focus on Rho. Oncogene 17: 1439-1445.
- 46. Lebowitz PF, Davide JP, Prendergast GC (1995) Evidence that farnesyltransferase inhibitors suppress Ras transformation by interfering with Rho activity. Mol Cell Biol 15: 6613-6622.
- 47. van Golen KL, Bao L, DiVito MM, Wu Z, Prendergast GC, et al. (2002) Reversion of RhoC GTPase-induced inflammatory breast cancer phenotype by treatment with a farnesyl transferase inhibitor. Mol Cancer Ther 1: 575-583.
- Prendergast GC (2002) Farnesyltransferase inhibitors: potential therapeutic for inflammatory breast cancer? Breast Dis 15: 25-32.
- Barrios J, Wieder R (2009) Dual FGF-2 and intergrin alpha5beta1 signaling mediate GRAF-induced RhoA inactivation in a model of breast cancer dormancy. Cancer Microenviron 2: 33-47.
- 50. Michael EL, Reju K, Monika B, Robert W (2004) Role of RhoA in survival of dormant breast cancer cells. American Association of Cancer Research
- Simpson KJ, Dugan AS, Mercurio AM (2004) Functional analysis of the contribution of RhoA and RhoC GTPases to invasive breast carcinoma. Cancer Res 64: 8694-8701.
- Sequeira L, Dubyk CW, Riesenberger TA, Cooper CR, van Golen KL (2008) Rho GTPases in PC-3 prostate cancer cell morphology, invasion and tumor cell diapedesis. Clin Exp Metastasis 25: 569-579.
- 53. Chatterjee M, van Golen KL (2011) Farnesyl transferase inhibitor treatment of breast cancer cells leads to altered RhoA and RhoC GTPase activity and induces a dormant phenotype. Int J Cancer 129: 61-69.
- 54. Chaterjee M, van Golen KL (2011) Breast cancer stem cells survive periods

of farnesyl-transferase inhibitor-induced dormancy by undergoing autophagy. Bone Marrow Res 2011: 362938.

- 55. Chatterjee M, van Golen KL, RhoA, RhoC (2009) GTPases in dormancy of breast cancer Proc Amer Assoc for Cancer Res Denver, CO.
- 56. Levine B, Kroemer G (2008) Autophagy in the pathogenesis of disease. Cell 132: 27-42.
- 57. Amaravadi RK (2008) Autophagy-induced tumor dormancy in ovarian cancer. J Clin Invest 118: 3837-3840.
- Gewirtz DA (2009) Autophagy, senescence and tumor dormancy in cancer therapy. Autophagy 5: 1232-1234.
- Lock R, Debnath J (2008) Extracellular matrix regulation of autophagy. Curr Opin Cell Biol 20: 583-588.
- Cui Q, Tashiro S, Onodera S, Minami M, Ikejima T (2007) Oridonin induced autophagy in human cervical carcinoma HeLa cells through Ras, JNK, and P38 regulation. J Pharmacol Sci 105: 317-325.
- Wu H, Wang MC, Bohmann D (2009) JNK protects Drosophila from oxidative stress by trancriptionally activating autophagy. Mech Dev 126: 624-637.
- 62. Ginestier C, Hur MH, Charafe-Jauffret E, Monville F, Dutcher J, et al. (2007) ALDH1 is a marker of normal and malignant human mammary stem cells and a predictor of poor clinical outcome. Cell Stem Cell 1: 555-567.
- Huang EH, Hynes MJ, Zhang T, Ginestier C, Dontu G, et al. (2009) Aldehyde dehydrogenase 1 is a marker for normal and malignant human colonic stem cells (SC) and tracks SC overpopulation during colon tumorigenesis. Cancer Res 69: 3382-3389.
- Dontu G, Abdallah WM, Foley JM, Jackson KW, Clarke MF, et al. (2003) In vitro propagation and transcriptional profiling of human mammary stem/progenitor cells. Genes Dev 17: 1253-1270.
- Allan AL, Vantyghem SA, Tuck AB, Chambers AF (2006) Tumor dormancy and cancer stem cells: implications for the biology and treatment of breast cancer metastasis. Breast Dis 26: 87-98.
- 66. Charafe-Jauffret E, Ginestier C, Iovino F, Wicinski J, Cervera N, et al. (2009) Breast cancer cell lines contain functional cancer stem cells with metastatic capacity and a distinct molecular signature. Cancer Res 69: 1302-1313.
- 67. Rosenthal D, Zhang J, Bao L, Zhu L, Merajver S, et al. GTPase influences the breast cancer stem cell phenotype. World Stem Cell Summit. Detroit Michigan
- Kakarala M, Wicha MS (2008) Implications of the cancer stem-cell hypothesis for breast cancer prevention and therapy. J Clin Oncol 26: 2813-2820.
- Cristofanilli M (2010) Novel targeted therapies in inflammatory breast cancer. Cancer 116: 2837-2839.
- Pham PV, Phan NL, Nguyen NT, Truong NH, Duong TT, et al. (2011) Differentiation of breast cancer stem cells by knockdown of CD44: promising differentiation therapy. J Transl Med 9: 209.
- Schlaak M, Schmidt P, Bangard C, Kurschat P, Mauch C, et al. (2012) Regression of metastatic melanoma in a patient by antibody targeting of cancer stem cells. Oncotarget 3: 22-30.

This article was originally published in a special issue, **Potential Biomarkers** and **Therapeutic Targets in Cancer Stem Cells** handled by Editor(s). Dr. Murielle Mimeault, University of Nebraska Medical Center, USA