

Hypothesis: Spirulina may Slow the Growth and Spread of Ovarian Cancer by Interfering with Growth Factor Activity of Lysophosphatidic Acid

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Abstract

Lysophosphatidic acid (LPA) has emerged as a key autocrine growth factor for most ovarian cancers, promoting their proliferation, survival, invasiveness, dissemination within the peritoneal cavity, and angiogenic capacity. Effective LPA signaling requires activation of endosomal NADPH oxidase activity. Free bilirubin is now known to function intracellularly as a potent inhibitor of NADPH oxidase complexes. The cyanobacterial chromophore phycocyanobilin (PhyCB), via intracellular conversion to the bilirubin homolog phycocyanorubin, can likewise inhibit NADPH oxidase activity, and is orally active in this regard. The cell wall polysaccharides of cyanobacteria may also aid cancer control by activating innate immunity and inhibiting angiogenesis. Hence, consumption of edible cyanobacteria such as spirulina may have potential for slowing the growth and spread of ovarian cancer – as it has recently been shown to do with a human pancreatic adenocarcinoma.

Keywords: Lysophosphatidic acid; Ovarian cancer; NADPH oxidase; Spirulina; Phycocyanobilin

Lysophosphatidic Acid – a Key Autocrine Growth Factor for Ovarian Cancer

Lysophosphatidic acid (LPA) is produced by most ovarian cancer cells lines, and acts as an autocrine growth factor via its receptors LPA2 and LPA3 [1-5]. Stimulation of these receptors, linked to several heterotrimeric G proteins, promotes the phosphorylation of Akt1, ERK, EGFR, and STAT3, cox-2 expression, and the transcriptional activity of NF-kappaB and HIF-1alpha [6-11]. These effects stimulate proliferation, inhibit apoptosis, enable release of ovarian cancer cells into the peritoneal fluid by suppressing E-cadherin expression, promote invasion by inducing matrix proteases, and enhance angiogenesis by increasing the production of VEGF [5,12-19]. Studies with human ovarian cancers xenografted in nude mice demonstrate that interference with LPA signaling (via silent RNA knockdown of LPA receptors, or via transfection with an enzyme that cleaves LPA) markedly slows the spread of these cancers [13,20] – implying that LPA is an important growth factor for ovarian cancer in vivo. Levels of LPA in plasma and in peritoneal fluid are markedly higher in ovarian cancer patients than in healthy controls [21-23]. Intraperitoneal levels are highest – in part because peritoneal mesothelium also can make LPA [24] – and likely promote the formation of malignant ascites.

NADPH Oxidase Mediates LPA Signaling

Recent research indicates that stimulation of NADPH oxidase activity within endosomes plays a critical role in transmitting the signals triggered by LPA receptors [25,26]. Thus, the NADPH oxidase inhibitor DPI blocks the ability of LPA to promote phosphorylation of Akt and ERK, to activate NF-kappaB, and to stimulate proliferation [25]. Interaction of LPA with its receptors induces the internalization of these receptors into endosomes, and the activated receptors

stimulate local NADPH oxidase activity that generates hydrogen peroxide within these endosomes; this leads to cysteine sulfenic acid formation in neighboring proteins that may be required for optimal LPA receptor activity [26]. N-acetylcysteine mimics the inhibitory effects of DPI [25], presumably by reversing the oxidation of protein cysteine groups [26-28]. The NADPH oxidase inhibitor apocynin, as well as a cell permeable form of catalase, likewise inhibited LPA-mediated Akt phosphorylation [25]. Other studies with ovarian cancer cells lines have found that DPI promotes apoptosis, and decreases expression of HIF-1alpha and of VEGF [29,30]. Knockdown of the NADPH oxidase component Nox4 – the expression of which is increased in many of these cell lines – exerts similar effects [29].

Phycocyanobilin May Aid Ovarian Cancer Control by Suppressing NADPH Oxidase Activity

These considerations suggest that agents which can safely inhibit NADPH oxidase activity may have important potential for slowing the growth and spread of many ovarian cancers. Free intracellular bilirubin, generated via heme oxygenase activity, functions physiologically as an inhibitor of NADPH oxidase complexes [31-34]. Although bilirubin is too insoluble to be useful as an orally administrable drug, its chemical relative phycocyanobilin (PhyCB), a prominent light-harvesting chromophore in edible cyanobacteria such as spirulina, shares the ability of bilirubin to inhibit NADPH oxidase activity [35,36]. This likely reflects the fact that PhyCB, a metabolite of biliverdin, can be converted intracellularly by biliverdin reductase to the bilirubin homolog phycocyanorubin [35,37]. Moreover, spirulina and PhyCB-enriched spirulina extracts exert marked anti-inflammatory/antioxidant effects in rodent studies, suggesting that PhyCB can inhibit NADPH oxidase after oral administration [38]. Hence, oral consumption of adequate amounts of spirulina, or of PhyCB-enriched spirulina extracts, may have clinical potential for treatment of ovarian cancers. In this regard, dietary spirulina has been reported to slow the growth of a human pancreatic cancer in nude

mice by about 60% [39]; notably, NADPH oxidase activity has been found to be elevated and to exert pro-growth, pro-survival effects in pancreatic cancer cells lines [40-44].

The cell wall polysaccharides of spirulina also may have potential for aiding cancer control, as they can interact with toll receptors to boost innate immunity, while also impeding angiogenesis [45,46].

As a proviso, it should be noted that spirulina probably should not be administered in conjunction with taxane chemotherapy, as the killing mechanism of these drugs appears to be contingent on NADPH oxidase activation [47-50]. On the other hand, in light of evidence that LPA signaling renders ovarian cancer cells relatively resistant to apoptosis induction by platinum drugs and doxorubicin, it is conceivable that NADPH oxidase inhibition could boost responsiveness to these drugs; however, there appears to be no direct evidence for this [51-54]. In rodents, dietary spirulina has been reported to provide protection from the cardiotoxicity of doxorubicin and the nephrotoxicity of cisplatin [55,56].

References

- Fang X, Schummer M, Mao M, Yu S, Tabassam FH, et al. (2002) Lysophosphatidic acid is a bioactive mediator in ovarian cancer. *Biochim Biophys Acta* 1582: 257-264.
- Tanyi JL, Hasegawa Y, Lapushin R, Morris AJ, Wolf JK, et al. (2003) Role of decreased levels of lipid phosphate phosphatase-1 in accumulation of lysophosphatidic acid in ovarian cancer. *Clin Cancer Res* 9: 3534-3545.
- Wang P, Wu X, Chen W, Liu J, Wang X (2007) The lysophosphatidic acid (LPA) receptors their expression and significance in epithelial ovarian neoplasms. *Gynecol Oncol* 104: 714-720.
- Tsujiuchi T, Araki M, Hirane M, Dong Y, Fukushima N (2014) Lysophosphatidic acid receptors in cancer pathobiology. *Histol Histopathol* 29: 313-321.
- Goldsmith ZG, Ha JH, Jayaraman M, Dhanasekaran DN (2011) Lysophosphatidic Acid Stimulates the Proliferation of Ovarian Cancer Cells via the gep Proto-Oncogene Ga (12). *Genes Cancer* 2: 563-575.
- Oyesanya RA, Lee ZP, Wu J, Chen J, Song Y, et al. (2008) Transcriptional and post-transcriptional mechanisms for lysophosphatidic acid-induced cyclooxygenase-2 expression in ovarian cancer cells. *FASEB J* 22: 2639-2651.
- Yang K, Zheng D, Deng X, Bai L, Xu Y, et al. (2008) Lysophosphatidic acid activates telomerase in ovarian cancer cells through hypoxia-inducible factor-1alpha and the PI3K pathway. *J Cell Biochem* 105: 1194-1201.
- Kim EK, Yun SJ, Do KH, Kim MS, Cho M, et al. (2008) Lysophosphatidic acid induces cell migration through the selective activation of Akt1. *Exp Mol Med* 40: 445-452.
- Jeong KJ, Park SY, Seo JH, Lee KB, Choi WS, et al. (2008) Lysophosphatidic acid receptor 2 and Gi/Src pathway mediate cell motility through cyclooxygenase 2 expression in CAOV-3 ovarian cancer cells. *Exp Mol Med* 40: 607-616.
- Seo JH, Jeong KJ, Oh WJ, Sul HJ, Sohn JS, et al. (2010) Lysophosphatidic acid induces STAT3 phosphorylation and ovarian cancer cell motility: their inhibition by curcumin. *Cancer Lett* 288: 50-56.
- Dutta S, Wang FQ, Wu HS, Mukherjee TJ, Fishman DA (2011) The NFkB pathway mediates lysophosphatidic acid (LPA)-induced VEGF signaling and cell invasion in epithelial ovarian cancer (EOC). *Gynecol Oncol* 123: 129-137.
- Hu YL, Tee MK, Goetzl EJ, Auersperg N, Mills GB, et al. (2001) Lysophosphatidic acid induction of vascular endothelial growth factor expression in human ovarian cancer cells. *J Natl Cancer Inst* 93: 762-768.
- Tanyi JL, Morris AJ, Wolf JK, Fang X, Hasegawa Y, et al. (2003) The human lipid phosphate phosphatase-3 decreases the growth, survival, and tumorigenesis of ovarian cancer cells: validation of the lysophosphatidic acid signaling cascade as a target for therapy in ovarian cancer. *Cancer Res* 63: 1073-1082.
- So J, Wang FQ, Navari J, Schreher J, Fishman DA (2005) LPA-induced epithelial ovarian cancer (EOC) in vitro invasion and migration are mediated by VEGF receptor-2 (VEGF-R2). *Gynecol Oncol* 97: 870-878.
- Wang GL, Wen ZQ, Xu WP, Wang ZY, Du XL, et al. (2008) Inhibition of lysophosphatidic acid receptor-2 expression by RNA interference decreases lysophosphatidic acid-induced urokinase plasminogen activator activation, cell invasion, and migration in ovarian cancer SKOV-3 cells. *Croat Med J* 49: 175-181.
- Ogata S, Morishige K, Sawada K, Hashimoto K, Mabuchi S, et al. (2009) Fasudil inhibits lysophosphatidic acid-induced invasiveness of human ovarian cancer cells. *Int J Gynecol Cancer* 19: 1473-1480.
- Wang FQ, Fisher J, Fishman DA (2011) MMP-1-PAR1 axis mediates LPA-induced epithelial ovarian cancer (EOC) invasion. *Gynecol Oncol* 120: 247-255.
- Jeong KJ, Park SY, Cho KH, Sohn JS, Lee J, et al. (2012) The Rho/ROCK pathway for lysophosphatidic acid-induced proteolytic enzyme expression and ovarian cancer cell invasion. *Oncogene* 31: 4279-4289.
- Liu Y, Burkhalter R, Symowicz J, Chaffin K, Ellerbroek S, et al. (2012) Lysophosphatidic Acid disrupts junctional integrity and epithelial cohesion in ovarian cancer cells. *J Oncol* 2012: 501492.
- Yu S, Murph MM, Lu Y, Liu S, Hall HS, et al. (2008) Lysophosphatidic acid receptors determine tumorigenicity and aggressiveness of ovarian cancer cells. *J Natl Cancer Inst* 100: 1630-1642.
- Sedlakova I, Vavrova J, Tošner J, Hanousek L (2011) Lysophosphatidic acid (LPA) (LPA)-a perspective marker in ovarian cancer. *Tumour Biol* 32: 311-316.
- Bese T, Barbaros M, Baykara E, Guralp O, Cengiz S, et al. (2010) Comparison of total plasma lysophosphatidic acid and serum CA-125 as a tumor marker in the diagnosis and follow-up of patients with epithelial ovarian cancer. *J Gynecol Oncol* 21: 248-254.
- Li YY, Zhang WC, Zhang JL, Zheng CJ, Zhu H, et al. (2015) Plasma levels of lysophosphatidic acid in ovarian cancer versus controls: a meta-analysis. *Lipids Health Dis* 14: 72.
- Ren J, Xiao YJ, Singh LS, Zhao X, Zhao Z, et al. (2006) Lysophosphatidic acid is constitutively produced by human peritoneal mesothelial cells and enhances adhesion, migration, and invasion of ovarian cancer cells. *Cancer Res* 66: 3006-3014.
- Saunders JA, Rogers LC, Klomsiri C, Poole LB, Daniel LW (2010) Reactive oxygen species mediate lysophosphatidic acid induced signaling in ovarian cancer cells. *Free Radic Biol Med* 49: 2058-2067.
- Klomsiri C, Rogers LC, Soito L, McCauley AK, King SB, et al. (2014) Endosomal H2O2 production leads to localized cysteine sulfenic acid formation on proteins during lysophosphatidic acid-mediated cell signaling. *Free Radic Biol Med* 71: 49-60.
- Lo Conte M, Carroll KS (2013) The redox biochemistry of protein sulenylation and sulfinylation. *J Biol Chem* 288: 26480-26488.
- Dickinson DA, Forman HJ (2002) Glutathione in defense and signaling: lessons from a small thiol. *Ann N Y Acad Sci* 973: 488-504.
- Xia C, Meng Q, Liu LZ, Rojanasakul Y, Wang XR, et al. (2007) Reactive oxygen species regulate angiogenesis and tumor growth through vascular endothelial growth factor. *Cancer Res* 67: 10823-10830.
- Jiang Z, Fletcher NM, Ali-Fehmi R, Diamond MP, Abu-Soud HM, et al. (2011) Modulation of redox signaling promotes apoptosis in epithelial ovarian cancer cells. *Gynecol Oncol* 122: 418-423.
- Lanone S, Bloc S, Foresti R, Almolki A, Taillé C, et al. (2005) Bilirubin decreases nos2 expression via inhibition of NAD(P)H oxidase: implications for protection against endotoxic shock in rats. *FASEB J* 19: 1890-1892.
- Matsumoto H, Ishikawa K, Itabe H, Maruyama Y (2006) Carbon monoxide and bilirubin from heme oxygenase-1 suppresses reactive oxygen species generation and plasminogen activator inhibitor-1 induction. *Mol Cell Biochem* 291: 21-28.

33. Jiang F, Roberts SJ, Datla Sr, Dusting GJ (2006) NO modulates NADPH oxidase function via heme oxygenase-1 in human endothelial cells. *Hypertension* 48: 950-957.
34. Datla SR, Dusting GJ, Mori TA, Taylor CJ, Croft KD, et al. (2007) Induction of heme oxygenase-1 in vivo suppresses NADPH oxidase derived oxidative stress. *Hypertension* 50: 636-642.
35. McCarty MF (2007) Clinical potential of Spirulina as a source of phycocyanobilin. *J Med Food* 10: 566-570.
36. Zheng J, Inoguchi T, Sasaki S, Maeda Y, McCarty MF, et al. (2012) Phycocyanin and Phycocyanobilin from *Spirulina Platensis* Protect against Diabetic Nephropathy by Inhibiting Oxidative Stress. *Am J Physiol Regul Integr Comp Physiol* 304: R110-120.
37. Terry MJ, Maines MD, Lagarias JC (1993) Inactivation of phytochrome- and phycobiliprotein-chromophore precursors by rat liver biliverdin reductase. *J Biol Chem* 268: 26099-26106.
38. Romay Ch, González R, Ledón N, Ramirez D, Rimbau V (2003) C-phycocyanin: a biliprotein with antioxidant, anti-inflammatory and neuroprotective effects. *Curr Protein Pept Sci* 4: 207-216.
39. **Koníčková R, Vaňková K, Vaníková J, Váňová K, Muchová L, et al. (2014) Anti-cancer effects of blue-green alga *Spirulina platensis*, a natural source of bilirubin-like tetrapyrrolic compounds. *Ann Hepatol* 13: 273-283.**
40. Vaquero EC, Edderkaoui M, Pandol SJ, Gukovsky I, Gukovskaya AS (2004) Reactive oxygen species produced by NAD(P)H oxidase inhibit apoptosis in pancreatic cancer cells. *J Biol Chem* 279: 34643-34654.
41. Mochizuki T, Furuta S, Mitsushita J, Shang WH, Ito M, et al. (2006) Inhibition of NADPH oxidase 4 activates apoptosis via the AKT/apoptosis signal-regulating kinase 1 pathway in pancreatic cancer PANC-1 cells. *Oncogene* 25: 3699-3707.
42. Lee JK, Edderkaoui M, Truong P, Ohno I, Jang KT, et al. (2007) NADPH oxidase promotes pancreatic cancer cell survival via inhibiting JAK2 dephosphorylation by tyrosine phosphatases. *Gastroenterology* 133: 1637-1648.
43. Du J, Liu J, Smith BJ, Tsao MS, Cullen JJ (2011) Role of Rac1-dependent NADPH oxidase in the growth of pancreatic cancer. *Cancer Gene Ther* 18: 135-143.
44. Du J, Nelson ES, Simons AL, Olney KE, Moser JC, et al. (2013) Regulation of pancreatic cancer growth by superoxide. *Mol Carcinog* 52: 555-567.
45. Hirahashi T, Matsumoto M, Hazeki K, Saeki Y, Ui M, et al. (2002) Activation of the human innate immune system by *Spirulina*: augmentation of interferon production and NK cytotoxicity by oral administration of hot water extract of *Spirulina platensis*. *Int Immunopharmacol* 2: 423-434.
46. Kawanishi Y, Tominaga A, Okuyama H, Fukuoka S, Taguchi T, et al. (2013) Regulatory effects of *Spirulina* complex polysaccharides on growth of murine RSV-M glioma cells through Toll-like receptor 4. *Microbiol Immunol* 57: 63-73.
47. Cao DX, Qiao B, Ge ZQ, Yuan YJ (2004) Comparison of burst of reactive oxygen species and activation of caspase-3 in apoptosis of K562 and HL-60 cells induced by docetaxel. *Cancer Lett* 214: 103-113.
48. Cao D, Qiao B, Ge Z, Yuan Y (2005) Amplification loop cascade for increasing caspase activity induced by docetaxel. *J Cell Biochem* 96: 810-820.
49. Alexandre J, Batteux F, Nicco C, Chéreau C, Laurent A, et al. (2006) Accumulation of hydrogen peroxide is an early and crucial step for paclitaxel-induced cancer cell death both in vitro and in vivo. *Int J Cancer* 119: 41-48.
50. Alexandre J, Hu Y, Lu W, Pelicano H, Huang P (2007) Novel action of paclitaxel against cancer cells: bystander effect mediated by reactive oxygen species. *Cancer Res* 67: 3512-3517.
51. Vidot S, Witham J, Agarwal R, Greenhough S, Bamrah HS, et al. (2010) Autotaxin delays apoptosis induced by carboplatin in ovarian cancer cells. *Cell Signal* 22: 926-935.
52. Frankel A, Mills GB (1996) Peptide and lipid growth factors decrease cis-diamminedichloroplatinum-induced cell death in human ovarian cancer cells. *Clin Cancer Res* 2: 1307-1313.
53. Hooks SB, Callihan P, Altman MK, Hurst JH, Ali MW, et al. (2010) Regulators of G-Protein signaling RGS10 and RGS17 regulate chemoresistance in ovarian cancer cells. *Mol Cancer* 9: 289.
54. E S, Lai YJ, Tsukahara R, Chen CS, Fujiwara Y, et al. (2009) Lysophosphatidic acid 2 receptor-mediated supramolecular complex formation regulates its antiapoptotic effect. *J Biol Chem* 284: 14558-14571.
55. Khan M, Shobha JC, Mohan IK, Naidu MU, Sundaram C, et al. (2005) Protective effect of *Spirulina* against doxorubicin-induced cardiotoxicity. *Phytother Res* 19: 1030-1037.
56. Mohan IK, Khan M, Shobha JC, Naidu MU, Prayag A, et al. (2006) Protection against cisplatin-induced nephrotoxicity by *Spirulina* in rats. *Cancer Chemother Pharmacol* 58: 802-808.