

## Short Communication

## Open Access

## Fast Sampling of Adherent Cell Cultures for Optimal Metabolomics Results

Bordag N<sup>1</sup>, Rennefahrt U<sup>1</sup>, Nachtigall J<sup>1</sup>, Maldonado SG<sup>1</sup>, Reszka RC<sup>2\*</sup>, Ramirez-Hernandez T<sup>3</sup>, Kamp H<sup>3</sup>, Fux E<sup>1</sup>, and van Ravenzwaay B<sup>3</sup><sup>1</sup>metanomics GmbH, Berlin, Germany<sup>2</sup>Metanomics Health GmbH, Berlin, Germany<sup>3</sup>BASF SE, Ludwigshafen, Germany

## Abstract

Metabolomics is a valuable tool to gain mechanistic insight into biological processes. It is frequently used to obtain complementary details to other 'omics technologies such as transcriptomics or proteomics. For knowledge generation, reproducible measurements of physiological, intact, and artifact-free metabolite levels are imperative necessitating further standardization of best practices to improve reliability of research outcomes. Here we report a novel cell sample preparation method (MxP® CellCollect) for metabolomics applications using adherent mammalian cells, which reduces the time consumption and physiological stress of conventional methods such as trypsinization or cell scraping. The most common sampling procedures to detach cells from their growth surface, trypsinization and scraping were compared to the MxP® CellCollect method investigating metabolite profiles of two breast cancer cell lines (MDA-MB-231 and MCF7). Metabolite levels as well as direction of metabolite changes differed tremendously revealing issues with trypsinization and scraping risking non-physiological or misleading results in contrast to MxP® CellCollect. Differences in metabolic profiles of cells harvested by trypsinization as compared to MxP® CellCollect or scraping can be directly attributed to prolonged, medium-free incubation time during cell detachment leading to a severely energy-depleted intracellular state. Labile metabolites or metabolites with fast intracellular turnover rates such as glycolysis and TCA cycle intermediates were strongly and significantly decreased by trypsinization. The same was true for amino acids and nucleoside triphosphates. Results obtained with scraping using methanol as solvent were multifaceted. Even mild evaporation of methanol prior metabolite extraction led to temperature- and/or light-dependent degradation of labile metabolites such as nucleoside triphosphates into di- and monophosphates liberating pyrophosphate. Furthermore, lipid metabolites, in particular cell membrane lipids, were found to have significantly lower levels than measured by trypsinization or MxP® CellCollect, indicating that lipid metabolites are insufficiently detached and/or unspecifically adsorb to the hydrophobic dish and scraping tool.

**Keywords:** Metabolomics; *In vitro* sampling procedures; Adherent eukaryotic cells; Scraping; Trypsinization; MxP® CellCollect

## Abbreviations

ANOVA: Analysis of Variance; CHCl<sub>3</sub>: Chloroform; DCM: Dichloromethane; EtOH: Ethanol; FBS: Fetal Bovine Serum; GC: Gas Chromatography; HPLC: High Performance LC; ISTD: Internal Standard; LC: Liquid Chromatography; MeOH: Methanol; MRM: Multiple Reaction Monitoring; MS: Mass Spectrometry; PBS: Phosphate Buffered Saline; PCA: Principal Component Analysis; RPMI: Roswell Park Memorial Institute; TCA: Tricarboxylic Acid Cycle; TNBC: Triple Negative Breast Cancer; s: Second(s)

## Introduction

Metabolomics is a valuable tool generating biological insight from the most diverse sample types ranging from plants, to plasma, tissue or cell culture. The use of metabolomics to obtain insight into the toxicological properties and mode of action of compounds when tested in rats, has been demonstrated in the MetaMap®Tox Project [1,2] and a successful step towards using this technology for *in vitro* systems was shown by [3]. With adherent cells metabolomics is now becoming an invaluable tool to study disease mechanisms and mode of action of compounds, to discover drug targets and off-target effects, to observe resistance strategies, and to investigate toxicities [4-12]. A suitable sampling method is a key feature for routine use of high quality metabolomics with adherent cells. The method must be sufficiently robust, reproducible, easily performable, and most importantly, able to capture the physiological, non-perturbed intracellular metabolism [7]. The purpose of this investigation was to compare the most commonly used sampling procedures such as trypsinization and scraping, with a newly developed method called MxP® CellCollect and to suggest better standards in metabolomics sampling practices.

## Materials and Method

## Cell culture and harvesting

Six technical replicates per cell line and harvest condition were cultivated. MDA-MB-231 cells were freshly obtained from ATCC (HTB-26, Lot. 59922846) using passage 8 and MCF7 house-kept cells equivalent to ATCC (HTB-22) using passage 11 for experiments. Cells harvested by trypsinization or scraping were grown in standard uncoated 8.70 cm<sup>2</sup> 6-well polystyrene culture dishes (83.3920.300, Sarstedt AG&Co.). Cells harvested by MxP® CellCollect were grown in uncoated 6.4 cm<sup>2</sup> lumox® dishes 35 (94.6077.331, Sarstedt AG&Co.). Per dish 0.25·10<sup>6</sup> MDA-MB-231 and 0.3·10<sup>6</sup> MCF7 cells were seeded and grown in RPMI-1640 medium (+10% FBS+2 mM glutamine+100 µg/ml Penicillin/Streptomycin) under standard growth conditions (37°C, 5% CO<sub>2</sub>) reaching 80% confluence after 48 h.

For trypsinization, as described in [13], cells were quickly washed once with PBS (HPLC grade) and 0.2 mL 0.05% trypsin in 0.02% EDTA (R-001-100, Gibco®) were added for 5 min at 37°C. Detached cells were gently suspended in 0.4 mL fresh medium (37°C), transferred and spun down with 900 rpm for 3 min at 4°C. To mimic cell counting an

**\*Corresponding author:** Reszka RC, Metanomics Health GmbH, Berlin, Germany, Tel: +49 30 34807 407; Fax: +49 30 34807 401; E-mail: [regina.reszka@metanomics-health.de](mailto:regina.reszka@metanomics-health.de)

**Received** January 06, 2016; **Accepted** January 23, 2016; **Published** January 25, 2016

**Citation:** Bordag N, Rennefahrt U, Nachtigall J, Maldonado SG, Reszka RC, et al. (2016) Fast Sampling of Adherent Cell Cultures for Optimal Metabolomics Results. Metabolomics 6: 164. doi:[10.4172/2153-0769.1000164](https://doi.org/10.4172/2153-0769.1000164)

**Copyright:** © 2016 Bordag N, et al. This is an open-access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

artificial break of 5 min was introduced. Cells were thereafter washed twice with PBS (4°C).

For scraping, as described in [12], cell monolayers were quickly washed twice with ice-cold 5% maltose solution and residual fluid was removed with vacuum. Immediately 1 mL MeOH (HPLC grade) (-20°C) was added and cells were scraped (2-Posit. Blade 25, thermoplastic elastomers, Sarstedt AG&Co).

For MxP® CellCollect (Figure 1) the lumox® membrane (with the adherent cells on) was cut out with a scalpel, falling into a dish with 37°C warm washing buffer (isotonic NaCl for MxP® Broad Profiling or 4.5 g/L glucose in isotonic NaCl for MxP® Energy) [14]. The membrane was quickly picked up with tweezers and dip-washed twice in two further dishes with fresh washing buffer. The membrane was placed on top of a 2 mL Eppendorf tube and pushed inside with a pipette tip adding 600 µL DCM/EtOH (9:11) for MxP® Broad Profiling or 900 µL (2:1) DCM/EtOH for MxP® Energy (-80°C). The procedure from cutting the membrane until freezing should not exceed 30 s for each sample to prevent metabolic changes during sampling. All harvested cells were immediately snap frozen in liquid nitrogen and stored until extraction at -80°C. Directly before extraction, the MeOH of scraped cells was removed by mild evaporation over a few minutes at 40°C and DCM/EtOH (mixtures as above) were added. Likewise, DCM/EtOH mixtures were added to trypsinized cell pellets after thawing before routine extraction and measurement (described in supplementary information).

## Statistical analysis

Metabolite values were log10-transformed for all statistical analysis in order to better approximate a normal distribution. For univariate analysis a linear ANOVA model was set up in the statistical software R [15] with the categorical fixed factors METHOD and CELL\_LINE (readouts: ratios, p-values, t-values, Benjamini-Hochberg corrected q-values in supplementary Table 1). For multivariate analysis, data was further centered and scaled to unit variance and multivariate statistics were performed by using Simca P+ software (version 13, Umetrics AB, Umeå, Sweden).

## Results and Discussion

A quick sampling is a key prerequisite in order to capture physiological intracellular metabolite levels and to avoid loss of metabolites with high turnover rates (e.g. glycolysis intermediates). Since metabolite concentrations are very sensitive to any variation in the cell environment an ideal harvest and quenching method for metabolomics analysis would immediately stop all intracellular enzymatic activities and subsequently all intracellular changes in metabolite concentrations, without first affecting the cell environment. Furthermore, such a method should be rapid, highly efficient, and reproducible to allow an unbiased measurement of cellular metabolite

concentrations, and to enable direct comparison of a large number of analytical samples. At the same time all cells with all of their metabolites must be collected and contaminations from spent media have to be avoided [16]. The necessary washing must leave metabolism unperturbed and should not induce any leakage or metabolite loss. Although great advances have been achieved by introduction of direct quenching and scraping instead of trypsinization [11,16-19] still a major drawback remains, namely the need for manual scraping and collection of cells. These steps might increase the variability of biological replicates due to differences in efficiency of cell transfer from the cell culture dish or cover slight into the extraction vial. Furthermore, these steps are time-consuming and therefore, might reduce overall attractiveness of this method to be used in high throughput cell culture experiments including several hundred samples in parallel.

## New sampling method – MxP® CellCollect

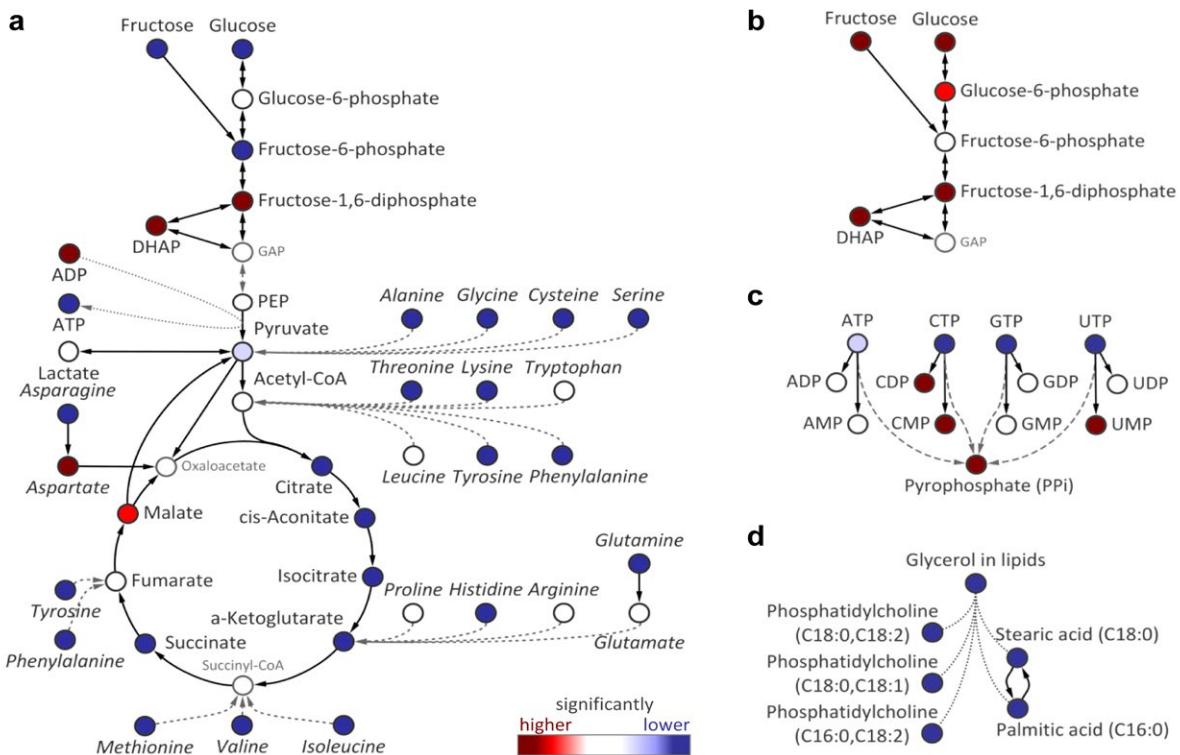
The cells are grown on lumox® dishes, which foremost offer excellent gas diffusion through the thin, gas-permeable, membrane-like bottom on which the cells grow. More importantly, these dishes allow cutting out the membrane and thereby to collect all cells quickly at once (Figure 1). The cells on the membrane are gently and thoroughly washed by immediate dipping into two consecutive dishes. The metabolism is then rapidly quenched by pushing the membrane into a tube with a pipette tip, immediately adding -80°C precooled solvent and further snap freezing in liquid nitrogen. The time from cells being without medium until quenching is typically around a few seconds and the whole procedure takes less than 30 s. Standard coatings such as fibronectin were well compatible and cell-free blanks incubated with medium as additional controls showed no impact on results.

## Comparison of trypsinization and scraping

The two most conventional cell harvesting protocols, trypsinization [13], and scraping in cold MeOH [12], were directly compared to the developed method for two different cell lines while all other parameters (extraction, measurement, normalization) were kept constant. Both multivariate (PCA, supplementary information Figure S1) and univariate analysis (ANOVA, Figure 2 and supplementary information Table 1) revealed obvious differences between all three methods. Differences in the metabolic profiles of cells harvested by trypsinization as compared to MxP® CellCollect or scraping can be directly attributed to the prolonged, medium-free incubation time during cell detachment leading to a severely energy-depleted intracellular state (Figure 2a). Glycolysis and TCA cycle intermediates as well as almost all amino acids and nucleoside triphosphates were strongly and statistically significantly decreased. Adenylate energy charge (AEC=ATP+0.5·ADP/(ATP+ADP+AMP)) was with a value of 0.70 ( $\pm$  0.05) far below AEC levels found with scraping or MxP® CellCollect (with values of 0.82 ( $\pm$  0.07) and 0.84 ( $\pm$  0.03), respectively). Although trypsinization is a valuable method for cell detachment from their growth surface (e.g. for cell passaging), our results and literature data [4,20] emphasize that trypsinization is not a suitable choice for physiological metabolomics since trypsin severely alters the physiological state of cells due to its interaction with membrane proteins resulting in sustained membrane damage in combination with metabolite leakage, and decreased cell viability, all of which significantly alter metabolite levels and cellular metabolite profiles [11]. As a consequence a considerable and highly variable portion of metabolites will be released into the trypsin solution and/or washing buffer depending on the length of trypsinization and washing/centrifugation steps. Furthermore, metabolic data varies for extremely labile metabolites or for metabolites with fast turnover



**Figure 1:** Sampling procedure. MxP® CellCollect sampling consists of cutting the membrane, dip washing the cells on the membrane, and quenching the metabolism with -80°C cold solvent. The transparent membrane was colored red for better visibility.



**Figure 2:** ANOVA comparison of different sampling methods across both cell lines. For a given comparison between two sampling methods red metabolite nodes signify higher metabolite levels while blue nodes signify lower levels with a significance of  $q$ -values  $<0.1$  for least intensive colors to  $q$ -values  $<0.01$  for most intensive colors. Grey metabolites were not measured and grey dashed arrows implicate several enzymatic reaction steps which were omitted for the sake of clarity. a) Comparison of trypsinization versus MxP® CellCollect reveals differences in glycolysis, TCA-cycle and proteinogenic amino acids. b), c), d) Comparison of scraping versus MxP® CellCollect with b) focusing on first steps of glycolysis, c) summarizing results for nucleoside tri-, di- and monophosphates and d) showing results of selected lipid metabolites and their fatty acid precursors. The glycerol in lipids describes the sum of all glycerol moieties that is released from the lipid fraction of a biological sample after hydrolysis of complex lipids and subsequent derivatization prior to gas chromatographic analysis. Several phosphatidylcholines are shown as examples together with two saturated fatty acids which represent the sum of its occurrence both in free and in all lipid-bound forms (dotted lines indicate that fatty acids and phosphatidylcholines contribute to the total sum of conjugated glycerol in complex lipids).

rates in response to changes in cell environment and the numerous time-consuming steps (e.g. centrifugation) required before enzymatic activities are finally inhibited by cell quenching [4,11]. During this time, a considerable portion of intracellular metabolites is lost from cells due to their small molecular size, the intracellular fast turnover rates [11,21], and the different osmotic strength of the applied solutions [22,23]. Nevertheless, trypsinization was and is still used frequently [13,24] and data obtained from trypsinized cells should be treated with utmost care, particularly if related to energy metabolism. The main reason for the use of trypsinization is the possibility to determine cell numbers for normalization purposes of the metabolic data. However, other normalization methods are available such as normalizing to cellular DNA after extraction [25]. In contrast, we have applied the median of all metabolites in a sample for normalization, which works well if the number of metabolites is sufficiently high ( $>50$ ) and data per sample is sufficiently comprehensive ( $<30\%$  missing values). Median normalization also saves considerable time and resources because additional measurements such as DNA or additional biological replicates for cell counting can be omitted. Additionally, harsh extraction protocols with mechanical cell disruption (e.g. bead milling) are necessary for good overall metabolite recoveries, in particular for recovery of lipid metabolites [17], but also strongly fragmented cellular DNA rendering DNA normalization impossible.

Results obtained with scraping were multifaceted and three major

artificial influences were identified. First, the chosen protocol from Urakami et al. included a washing step with 5% maltose which led, despite the very short incubation time, to a considerable increase in early glycolysis intermediates (Figure 2b). This emphasizes the importance of choosing adequate washing solutions. Preferably non-cold, isotonic, phosphate-free solutions should be used containing the same carbon sources as the culture medium tested. These carbon sources can also be fully isotopically labeled, such as all  $^{13}\text{C}$ , thereby slowing down  $^{12}\text{C}$  metabolism during washing and stabilizing early glycolysis intermediates (Bordag et al. in preparation). The second observed effect was that even mild evaporation of the scraping solvent MeOH led to temperature- and/or light-dependent degradation of labile metabolites such as nucleoside triphosphates into di- and monophosphates liberating pyrophosphate (Figure 2c). For MeOH-based extractions solvent removal is not necessary. However, the MeOH poses problems for other extractions and needs to be removed which would require validation of even milder solvent removal (e.g. by lowering temperature or reducing exposure to light or evaporation time). Third, levels of lipids especially from cell membrane lipids were found to have significantly lower levels than measured by trypsinization or MxP® CellCollect (Figure 2d). These findings indicate that lipid metabolites are insufficiently detached from the dish bottom and/or lipidic metabolites unspecifically adsorb to the hydrophobic dish and scraping spatula (area offered for adsorption is very large in comparison to the low amounts of lipidic metabolites). In order to achieve complete

recovery of lipidic metabolites the use of harsher solvents for scraping is necessary which in turn might risk the dissolving of the cell culture dish polystyrene plastic. In contrast, MxP® CellCollect is compatible with many solvents and harsh mechanical cell disruption which is a prerequisite for high recovery of polar and apolar metabolites [17] from the complete sample.

## Conclusion

We developed a user-friendly, fast, efficient, reproducible, and robust sampling method for adherent cells. In summary, MxP® CellCollect proves to deliver more reliable results for metabolic profiling of adherent cells than the widely used scraping and trypsinization procedures. The presented manual sampling is extremely flexible and easily adoptable. The method was developed so that it is also suitable for possible automatization and translation into 96-well format in the future. More than 150 different cell lines have been analyzed by applying MxP® CellCollect so far, indicating overall that this new method is compatible with any adherent cell type and any common extraction or measurement protocol (MS- or NMR-based), easily adoptable without the need for sophisticated equipment. Here a high-throughput, routine MS-based protocol was applied which is also commonly used for sample types such as suspension cells, plasma, blood, or urine, thereby leading to increasing comparability.

## Acknowledgement

The authors sincerely thank all of their team members and colleagues for their constant support and fruitful discussions.

## References

- van Ravenwaay B, Cunha GCP, Leibold E, Loos R, Mellert W, et al. (2007) The use of metabolomics for the discovery of new biomarkers of effect. *Toxicol Lett* 172: 21-28.
- van Ravenwaay B, Coelho-Palermo Cunha G, Fabian E, Herold M, Kamp H, et al. (2010) The use of metabolomics in cancer research. An Omics Perspective of Cancer. Springer.
- Balcke G, Kolle S, Kamp H, Bethan B, Loos R, et al. (2011) Linking energy metabolism to dysfunctions in mitochondrial respiration - A metabolomics in vitro approach. *Toxicology Letters* 203: 200-209.
- Halama A (2014) Metabolomics in cell culture--a strategy to study crucial metabolic pathways in cancer development and the response to treatment. *Arch Biochem Biophys* 564: 100-109.
- Kamp H, Fabian E, Groeters S, Herold M, Krennrich G, et al. (2012) Application of *in vivo* metabolomics to preclinical/toxicological studies: case study on phenytoin-induced systemic toxicity. *Bioanalysis* 4: 2291-2301.
- Kentner D, Martano G, Callon M, Chiquet P, Brodmann M, et al. (2014) Shigella reroutes host cell central metabolism to obtain high-flux nutrient supply for vigorous intracellular growth. *Proc Natl Acad Sci USA* 111: 9929-9934.
- Leon Z, Garcia-Canaveras JC, Donato MT, Lahoz A (2013) Mammalian cell metabolomics: experimental design and sample preparation. *Electrophoresis* 34: 2762-2775.
- Mattes W, Davis K, Fabian E, Greenhaw J, Herold M, et al. (2014) Detection of hepatotoxicity potential with metabolite profiling (metabolomics) of rat plasma. *Toxicol Lett* 230: 467-478.
- Mattes WB, Kamp HG, Fabian E, Herold M, Krennrich G, et al. (2013) Prediction of clinically relevant safety signals of nephrotoxicity through plasma metabolite profiling. *Biomed Res Int*.
- Ser Z, Liu X, Tang NN, Locasale JW (2015) Extraction parameters for metabolomics from cultured cells. *Anal Biochem* 475: 22-28.
- Teng Q, Huang WL, Collette TW, Ekman DR, Tan C (2009) A direct cell quenching method for cell-culture based metabolomics. *Metabolomics* 5: 199-208.
- Uraakami K, Zangiacomi V, Yamaguchi K, Kusuvara M (2013) Impact of 2-deoxy-D-glucose on the target metabolome profile of a human endometrial cancer cell line. *Biomed Res* 34: 221-229.
- Reitman ZJ, Jin G, Karoly ED, Spasojevic I, Yang J, et al. (2011) Profiling the effects of isocitrate dehydrogenase 1 and 2 mutations on the cellular metabolome. *Proc Natl Acad Sci USA* 108: 3270-3275.
- Daemen A, Peterson D, Sahu N, McCord R, Du X, et al. (2015) Metabolite profiling stratifies pancreatic ductal adenocarcinomas into subtypes with distinct sensitivities to metabolic inhibitors. *Proc Natl Acad Sci USA* 112: E4410-E4417.
- R Development Core Team R2R (2014) A language and environment for statistical computing.
- Martano G, Delmotte N, Kiefer P, Christen P, Kentner D, et al. (2015) Fast sampling method for mammalian cell metabolic analyses using liquid chromatography-mass spectrometry. *Nat Protoc* 10: 1-11.
- Danielsson AP, Moritz T, Mulder H, Spegel P (2010) Development and optimization of a metabolomic method for analysis of adherent cell cultures. *Anal Biochem* 404: 30-39.
- Lorenz MA, Burant CF, Kennedy RT (2011) Reducing time and increasing sensitivity in sample preparation for adherent mammalian cell metabolomics. *Anal Chem* 83: 3406-3414.
- Ritter JB, Genzel Y, Reichl U (2008) Simultaneous extraction of several metabolites of energy metabolism and related substances in mammalian cells: optimization using experimental design. *Anal Biochem* 373: 349-369.
- Deitmer K, Numberger N, Kaspar H, Gruber MA, Almstetter MF, et al. (2011) Metabolite extraction from adherently growing mammalian cells for metabolomics studies: optimization of harvesting and extraction protocols. *Anal Bioanal Chem* 399: 1127-1139.
- Villas-Boas SG, Hojer-Pedersen J, Akesson M, Smedsgaard J, Nielsen J (2005) Global metabolite analysis of yeast: evaluation of sample preparation methods. *Yeast* 22: 1155-1169.
- Britten RJ, McClure FT (1962) Amino Acid Pool in Escherichia-Coli. *Bacteriol Rev* 26: 292-335.
- Smeaton JR, Elliott WH (1967) Selective release of ribonuclease-inhibitor from *Bacillus subtilis* cells by cold shock treatment. *Biochem Biophys Res Commun* 26: 75-81.
- Ghosh JC, Siegelin MD, Vaira V, Favarsani A, Tavecchio M, et al. (2015) Adaptive mitochondrial reprogramming and resistance to PI3K therapy. *J Natl Cancer Inst* 107.
- Silva LP, Lorenzi PL, Purwaha P, Yong V, Hawke DH, et al. (2013) Measurement of DNA concentration as a normalization strategy for metabolomic data from adherent cell lines. *Anal Chem* 85: 9536-9542.

## Supplementary Information

### MxP® Broad Profiling

For MxP® Broad Profiling the samples were prepared and subjected to LC-MS/MS, GC-MS analysis as described below. After addition of water the samples were extracted using a ball mill (Retsch, Germany), filtered through a centrifuge filter (Millipore, mesh size 0.2 mm) and fractionated into an aqueous, polar phase and an organic, lipophilic phase. For the transmethanolysis of the lipid extracts (lipophilic phase) a mixture of 140 µL of chloroform, 37 µL of hydrochloric acid (37% by weight HCl in water), 320 µL of methanol and 20 µL of toluene was added to the evaporated extract. The vessel was sealed tightly and heated for 2 hours at 100°C, with shaking. The solution was subsequently evaporated to dryness. The residue was dried completely. The methoximation of the carbonyl groups was carried out by reaction with methoxyamine hydrochloride (20 mg/mL in pyridine, 100 µL for 1.5 hours at 60°C) in a tightly sealed vessel. 20 µL of a solution of odd-numbered, straight-chain fatty acids (solution of each 0.3 mg/mL of fatty acids from 7 to 25 carbon atoms and each 0.6 mg/mL of fatty acids with 27, 29, and 31 carbon atoms in 3/7 (v/v) pyridine/toluene) were added as time standards. Finally, the derivatization with 100 µL of N-methyl-N-(trimethylsilyl)-2,2,2-trifluoroacetamide (MSTFA) was carried out for 30 minutes at 60°C, again in the tightly sealed vessel. The final volume before injection into the GC was 200 µL. For the dried polar phase the derivatization was performed in the following way: the methoximation of the carbonyl groups was carried out by reaction with methoxyamine hydrochloride (20 mg/mL in pyridine, 50 µL for 1.5 hours at 60°C) in a tightly sealed vessel. 10 µL of a solution of odd-numbered, straight-chain fatty acids (solution of each 0.3 mg/mL of fatty acids from 7 to 25 carbon atoms and each 0.6 mg/mL of fatty acids with 27, 29, and 31 carbon atoms in 3/7 (v/v) pyridine/toluene) were added as time standards. Finally, the derivatization with 50 µL of N-methyl-N-(trimethylsilyl)-2,2,2-trifluoroacetamide (MSTFA) was carried out for 30 minutes at 60°C, again in the tightly sealed vessel. The final volume before injection into the GC was 100 µL.

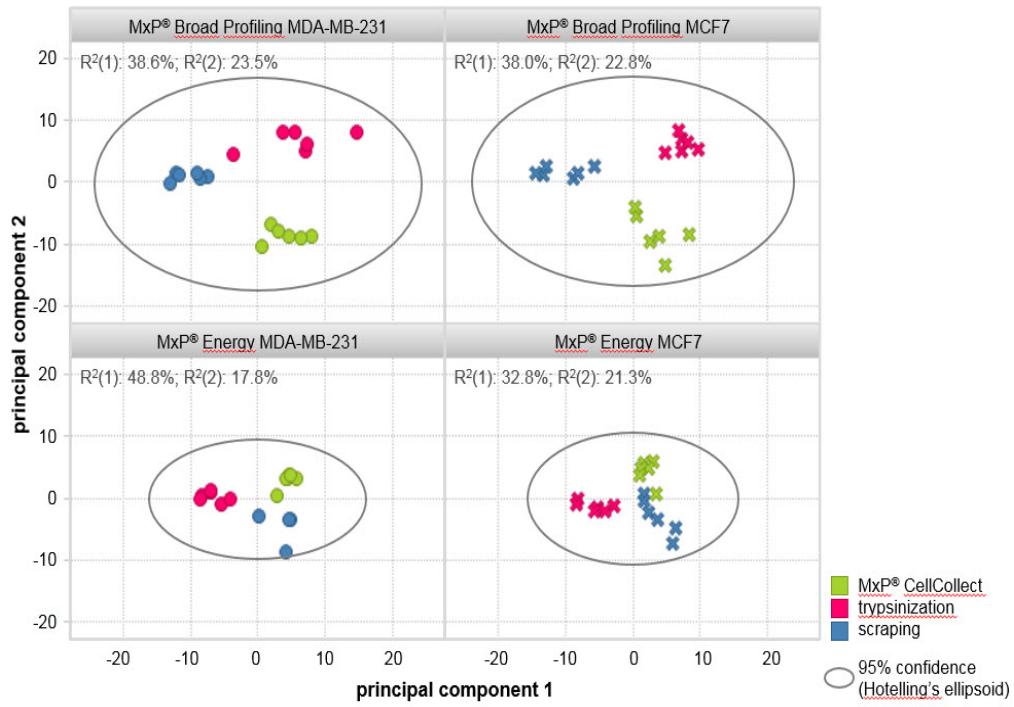
The GC-MS system consists of an Agilent 6890 GC coupled to an Agilent 5973 MSD (Agilent, Waldbronn, Germany), autosamplers were CompiPal or GCPal from CTC (CTC, Zwingen, Switzerland). In LC-MS analysis, both fractions were reconstituted in appropriate solvent mixtures. HPLC was performed by gradient elution on reversed phase separation columns. Mass spectrometric detection, which allows target and high sensitivity MRM (Multiple Reaction Monitoring) profiling in parallel to a full screen analysis, was applied as described in WO2003073464. The HPLC instruments were Agilent 1100 (Agilent, Waldbronn, Germany), the MS instruments were API4000 from SCIEX (AB SCIEX, Darmstadt, Germany). Data of 113 known and 74 unknown metabolites were normalized to the median of reference samples which were derived from a pool formed from additional biological aliquots of all sampling groups (→ polar reference material and lipid reference material) to account for inter- and intra-instrumental variation. In a second step metabolite profiling data was normalized to the sample-specific metabolite median to correct for variations in cell numbers.

### **MxP® Energy**

For MxP® Energy UPLC-MS/MS, an Aquity (Waters) coupled to the API5500 a negative mode ESI-MS/MS-System (Applied Biosystems), were used as described in [3]. In short, samples were kept at all times at 5°C or below. For extraction 1.5 M ammonium acetate and ISTD (<sup>13</sup>C-labeled yeast extract) were added to the cells in the DCM/EtOH quenching solution and the mixture was homogenized for 30 s in a bead mill FastPrep24 (MP Biomedicals) at -20°C. Phase separation was achieved by 2 min centrifugation with 14000 rpm at 4°C. Polar phases were spin-filtrated (Ultrafree<sup>®</sup>-MC 5.0 µm, Millipore) in 5 min. Samples were extracted and spin-filtrated a second time with additional 1.5 M ammonium acetate. Spin filters were finally washed with water and all three filtrates were combined and lyophilized to remove the ammonium acetate. Samples were resuspended in deionized water and the separation was performed by gradient elution at 45°C using 10 mM tributylammonium acetate (pH 6.2) in A) water and B) 50% ACN with a gradient from 5-90% on a 1.5 µm RP UPLC column (VisionHT-HL, 2.1 mm ×10 cm, Alltech-Grom). All 65 known metabolites were measured in MRM mode and each metabolite was normalized against the corresponding <sup>13</sup>C-analyte. In a second step metabolite profiling data was normalized to the median of all metabolites from one replicate dish. For production of <sup>13</sup>C-labeled, yeast extract as ISTD, a yeast culture of *Candida utilis* (DSMZ sp. 2361) was grown in shake flasks on 10 g/L [ $U^{13}C_6$ ] D-glucose in yeast nitrogen base amino acid-free medium (Y1250 Sigma-Aldrich). Cells were cultured under aerobic conditions at 28°C at 180 rpm on an orbital shaker and harvested at an optical density of 4 (600 nm, 1 cm cuvette) by short centrifugation at 4°C. Cells were washed twice with 10 g/L [ $U^{13}C_6$ ] D-glucose in 0.15 M ammonium acetate by centrifugation. The cell pellet was quenched with DCM/EtOH (2/1) and extracted as described for MxP® Broad Profiling. The polar phase containing the labelled metabolites was stored at -80°C until further use as ISTD.

### **Multivariate statistical analysis**

For multivariate analysis, data was further centered and scaled to unit variance and multivariate statistics were performed by using Simca P+ software (version 13, Umetrics AB, Umea, Sweden).



**Figure S1:** Principal component analysis (PCA) of cell line-specific metabolite profiles harvested either by trypsinization, scraping or MxP® CellCollect. The scores plots of all four models for the first two principal components reveal very large differences resulting from sampling within each of the two cell lines from two different measurement platforms.

ANOVA model, fixed categorical factors: ~ 1   METHOD + CELL_LINE		comparison of		Trypsin		Scraping		Scraping		MCF7		Trypsin		Scraping		Scraping		MCF7		Trypsin		Scraping		Scraping		MCF7		Trypsin		Scraping		Scraping		MCF7								
Normalization	ONTOLOGY1_NAME	ONTOLOGY2_NAME	METABOLITE_NAME	TR vs CC Ratio	SC vs CC Ratio	SC vs TR Ratio	MCF7 vs MDA-MB-231	TR vs CellCollect	SC vs CC Ratio	SC vs TR Ratio	MCF7 vs MDA-MB-231	TR vs CellCollect	SC vs CC Ratio	SC vs TR Ratio	MCF7 vs MDA-MB-231	TR vs CellCollect	SC vs CC Ratio	SC vs TR Ratio	MCF7 vs MDA-MB-231	TR vs CellCollect	SC vs CC Ratio	SC vs TR Ratio	MCF7 vs MDA-MB-231	TR vs CellCollect	SC vs CC Ratio	SC vs TR Ratio	MCF7 vs MDA-MB-231	TR vs CellCollect	SC vs CC Ratio	SC vs TR Ratio	MCF7 vs MDA-MB-231	TR vs CellCollect	SC vs CC Ratio	SC vs TR Ratio	MCF7 vs MDA-MB-231							
pool, median	Amino acids	Amino acids, acidic	Aspartate	1.92	1.50	0.78	4.40	2.63E-10	0.0007	7.99E-21	1.24E-05	0.0014	9.55E-20	9.89E-08	6.0298	-3.8312	27.6909	0.2823	1.760	-0.1063	0.6438	0.0285	0.0292	0.0277	0.0232	26	26	26	26	26	26	26	26	26	26	26	26	26	26	26		
pool, median	Amino acids	Amino acids, acidic	Glutamate	1.09	0.91	0.84	0.78	0.7151	0.0475	0.2278	0.7907	0.8018	0.5099	0.2798	0.3694	-0.3557	-0.7283	-1.2377	0.0377	-0.0393	-0.0773	-0.1077	0.1022	0.1106	0.1058	0.0870	24	24	24	24	24	24	24	24	24	24	24	24	24	24	24	
pool, median	Amino acids	Amino acids, aromatic	Phenylalanine	0.77	0.94	1.22	2.93	0.0012	0.3943	0.0094	2.98E-16	0.0037	0.4973	0.0140	1.70E-15	-3.6153	-0.8663	2.8052	18.0090	-0.1144	-0.0280	0.0863	0.4666	0.0316	0.0324	0.0308	0.0258	26	26	26	26	26	26	26	26	26	26	26	26	26	26	26
pool, median	Amino acids	Amino acids, aromatic	Tryptophan	1.03	0.68	0.66	2.12	0.7199	0.0002	6.77E-05	2.40E-10	0.7926	0.0008	0.0002	7.52E-10	0.3626	-4.2915	-4.7697	10.1466	0.0143	-0.1689	-0.1832	0.3253	0.0393	0.0394	0.0384	0.0321	25	25	25	25	25	25	25	25	25	25	25	25	25	25	25
pool, median	Amino acids	Amino acids, aromatic	Tyrosine	0.59	0.85	1.45	2.95	2.47E-07	0.0056	3.59E-05	8.44E-16	1.94E-06	0.0868	0.0001	4.61E-15	-6.9082	-0.2025	4.9752	17.3261	-0.2298	-0.0688	0.1610	0.4699	0.0333	0.0340	0.0324	0.0271	26	26	26	26	26	26	26	26	26	26	26	26	26	26	26
13C-ISTD, median	Amino acids	Amino acids, basic	Arginine	0.45	0.90	2.01	1.28	1.13E-05	0.0001	0.6511	6.60E-05	0.6207	0.0004	0.0893	5.2560	-0.6482	4.3470	1.9149	-0.3493	-0.0453	0.3039	1.0175	0.0665	0.0699	0.0699	0.0562	30	30	30	30	30	30	30	30	30	30	30	30	30	30	30	
pool, median	Amino acids	Amino acids, basic	Asparagine	0.49	1.46	3.01	2.97	1.31E-09	0.0004	1.80E-13	3.70E-15	2.05E-08	0.0004	7.55E-12	1.86E-14	-9.3178	4.6548	14.1996	16.8478	-0.3143	0.1647	0.4790	0.4721	0.0337	0.0354	0.0337	0.0280	25	25	25	25	25	25	25	25	25	25	25	25	25	25	25
pool, median	Amino acids	Amino acids, basic	Glutamine	0.58	0.77	1.33	4.21	0.0023	0.1291	0.0798	3.53E-11	0.0062	1.9294	0.1005	1.16E-10	-3.3776	-0.1567	1.8233	10.8848	-0.2378	-0.1129	0.1249	0.6247	0.0704	0.0270	0.0685	0.0574	26	26	26	26	26	26	26	26	26	26	26	26	26	26	26
pool, median	Amino acids	Amino acids, basic	Histidine	0.48	1.01	2.13	2.80	3.95E-08	0.0055	1.77E-08	9.0553	1.44E-07	0.1935	2.57E-13	-7.6602	0.1199	8.0007	13.0182	-0.3223	0.0052	0.3275	0.4465	0.0421	0.0431	0.0409	0.0343	26	26	26	26	26	26	26	26	26	26	26	26	26	26	26	
pool, median	Amino acids	Amino acids, basic	Lysine	0.58	0.81	1.39	1.33	4.79E-05	0.0054	0.0052	0.0041	0.0008	0.0068	4.8656	-1.8519	3.0539	3.1451	-0.2367	-0.0922	0.1445	0.1247	0.0486	0.0473	0.0397	0.0397	26	26	26	26	26	26	26	26	26	26	26	26	26	26	26		
pool, median	Amino acids	Amino acids, branched chain	Isoleucine	0.51	0.93	3.10	0.0014	0.7022	0.0030	9.76E-08	0.0040	0.7904	0.0051	2.53E-07	-3.5762	-0.3866	3.2696	7.2855	-0.2960	-0.0327	0.2632	0.4915	0.0828	0.0847	0.0805	0.0675	26	26	26	26	26	26	26	26	26	26	26	26	26	26	26	
pool, median	Amino acids	Amino acids, branched chain	Leucine	0.79	1.15	1.46	2.69	0.1206	0.0139	1.10E-08	0.1740	0.4651	0.0201	3.03E-08	-1.6047	0.9402	2.6385	8.2044	-0.1030	0.0617	0.1647	0.4299	0.0642	0.0567	0.0624	0.0523	26	26	26	26	26	26	26	26	26	26	26	26	26	26	26	
pool, median	Amino acids	Amino acids, branched chain	Valine	0.57	1.04	1.84	2.68	3.52E-05	0.0170	8.92E-06	6.25E-11	0.0002	0.7986	3.36E-05	2.04E-10	-4.9824	0.3637	5.5044	10.5969	-0.2467	0.1884	0.2652	0.4278	0.0495	0.0507	0.0482	0.0404	26	26	26	26	26	26	26	26	26	26	26	26	26	26	26
pool, median	Amino acids	Amino acids, neutral	Alanine	0.66	0.92	1.39	2.54	8.37E-05	0.0007	8.68E-13	0.0004	0.4802	0.0015	3.25E-12	-4.6544	-0.9138	3.8235	12.8747	-0.1793	-0.0360	0.1433	0.4044	0.0394	0.0375	0.0314	0.026	26	26	26	26	26	26	26	26	26	26	26	26	26	26	26	
pool, median	Amino acids	Amino acids, neutral	Glycine	0.73	0.97	1.34	1.71	7.24E-08	0.0049	2.08E-07	1.47E-14	0.0266	0.6408	1.19E-06	6.59E-14	-7.4087	-0.6073	6.9772	15.3637	-0.1380	-0.0116	0.1264	0.2333	0.0186	0.0191	0.0181	0.0152	26	26	26	26	26	26	26	26	26	26	26	26	26	26	26
pool, median	Amino acids	Amino acids, neutral	Proline	0.80	0.84	1.06	1.28	0.0164	0.0661	5.25E-05	1.91E-15	0.0023	0.0008	0.0068	-2.5560	-																										

<sup>13</sup> C-ISTD, median	Energy metabolism and relate	Glycolysis/Gluconeogenesis	Fructose-2,6-diphosphate	0.71	3.19	4.50	1.14	0.4812	0.0305	0.0061	0.7438	0.5697	0.0538	0.0098	0.8013	-0.7132	2.2714	2.9493	0.3299	-0.1502	0.5033	0.6535	0.0587	0.2106	0.2216	0.2216	0.1780	30	30	30	30
<sup>13</sup> C-ISTD, median	Energy metabolism and relate	Glycolysis/Gluconeogenesis	Fructose-6-phosphate	0.18	1.34	7.48	1.71	7.17E-06	0.3877	1.30E-06	0.0547	4.39E-05	0.4939	6.42E-06	0.0758	-5.4175	0.8766	6.0253	1.9997	-0.7468	0.1272	0.8740	0.2330	0.1379	0.1451	0.1451	0.1165	30	30	30	30
pool, median	Energy metabolism and relate	Glycolysis/Gluconeogenesis	Glucose-6-phosphate	1.14	1.69	1.48	0.67	0.5150	0.0151	0.0433	0.0190	0.6066	0.0293	0.0571	0.0284	0.6605	2.6100	2.1290	-2.5085	0.0566	0.2280	0.1714	-0.1718	0.0856	0.0874	0.0805	0.0685	25	25	25	25
<sup>13</sup> C-ISTD, median	Energy metabolism and relate	Glycolysis/Gluconeogenesis	Glycerate	0.75	1.86	2.48	0.97	0.1090	0.0019	2.43E-05	0.8353	0.1610	0.0046	7.67E-05	0.8735	-1.6517	3.4147	4.9844	-0.2098	-0.1242	0.2702	0.3944	-0.0133	0.0752	0.0791	0.0636	30	30	30	30	
<sup>13</sup> C-ISTD, median	Energy metabolism and relate	Glycolysis/Gluconeogenesis	Lactate	0.52	1.20	2.29	0.69	9.03E-06	0.2341	5.78E-06	0.0041	0.0004	0.3264	2.31E-05	0.0067	-4.5425	1.2152	5.5232	-3.1208	-0.2828	0.0778	0.3666	-0.1631	0.0623	0.0640	0.0652	0.0523	29	29	29	29
<sup>13</sup> C-ISTD, median	Energy metabolism and relate	Glycolysis/Gluconeogenesis	Phosphoenopyruvate (PEP)	5.08	1.68	0.33	2.45	0.0001	0.1903	0.0081	0.0074	0.0005	0.2778	0.0124	0.0113	4.3945	1.3399	2.8365	0.2728	0.7055	0.2263	-0.4791	0.3894	0.1605	0.1689	0.1357	30	30	30	30	
pool, median	Energy metabolism and relate	Glycolysis/Gluconeogenesis	Pyruvate (additional: Phosphoenopyruvate (PEP))	0.43	0.77	1.79	2.30	0.0193	0.4509	0.0903	0.0058	0.0384	0.5575	0.1110	0.0098	-2.4945	-0.7655	1.7591	3.0069	-0.3682	-0.1156	0.2526	0.3618	0.1476	0.1510	0.1436	0.1203	26	26	26	26
pool, median	Energy metabolism and relate	Ketone bodies	3-Hydroxybutyrate (additional: 2-Hydroxy-3-oxo)	1.38	1.65	1.20	0.96	2.39E-05	7.71E-09	0.0083	0.0409	0.0001	8.07E-08	0.0126	0.4612	4.9366	7.7516	2.8150	-0.8532	0.1384	0.2181	0.0792	-0.0192	0.0281	0.0281	0.0230	32	32	32	32	
<sup>13</sup> C-ISTD, median	Energy metabolism and relate	Pentose phosphate pathway	6-Phosphogluconic acid	0.40	0.65	1.62	0.92	1.89E-10	0.0002	4.59E-05	0.3027	4.32E-08	0.0004	0.0001	0.3567	-4.902	-4.1738	4.7599	-1.0488	-0.3943	-0.1842	0.2101	-0.0372	0.0419	0.0441	0.0355	30	30	30	30	
pool, median	Energy metabolism and relate	Pentose phosphate pathway	Ribose-5-phosphate	1.32	2.50	1.90	1.33	0.0031	7.29E-11	3.46E-08	0.0004	1.41E-09	2.72E-07	0.0007	3.2588	10.5207	7.7162	4.1027	0.1206	0.3985	0.2779	0.1238	0.0370	0.0379	0.0360	0.0302	26	26	26	26	
<sup>13</sup> C-ISTD, median	Energy metabolism and relate	Pentose phosphate pathway	Ribulose-5-phosphate (additional: Xylulose-5-ph)	1.07	1.03	0.96	1.16	0.5413	0.7934	0.1250	0.6261	0.8545	0.7641	0.1626	0.6180	0.2643	-0.3230	1.5782	0.0291	0.0131	-0.0160	0.0629	0.0472	0.0496	0.0399	30	30	30	30		
<sup>13</sup> C-ISTD, median	Energy metabolism and relate	Pentose phosphate pathway	Sedoheptulose-7-phosphate	0.72	0.32	0.44	0.76	0.4559	0.0174	0.0818	0.4510	0.5529	0.0328	0.1021	0.4987	-0.7553	-2.5184	-1.806	-0.7637	-0.1411	-0.4951	-0.3540	-0.1206	0.1869	0.1966	0.1579	30	30	30	30	
<sup>13</sup> C-ISTD, median	Hormones, signal substances	Other hormones, signal substances	Adenosine monophosphate, cyclic (cAMP)	0.86	2.55	2.97	0.58	0.6571	0.0120	0.0042	0.0634	0.7396	0.0242	0.0069	0.0874	-0.4483	2.6733	0.3094	-1.9277	-0.0649	0.4072	0.4721	-0.2387	0.1448	0.1523	0.1223	30	30	30	30	
pool, median	Hormones, signal substances	Other hormones, signal substances	gamma-Aminobutyrate (GABA)	0.65	0.75	1.17	0.88	1.47E-05	0.0023	0.0666	0.0781	8.39E-06	0.0056	0.0860	0.1055	-5.3139	-3.3729	1.9148	-1.8339	-0.1893	-0.1229	0.0663	-0.0532	0.0356	0.0364	0.0290	26	26	26	26	
pool, median	Miscellaneous	Miscellaneous	Pyrophosphate (PPi) (additional: Phosphate (inorganic))	1.51	2.47	1.63	1.43	0.0007	1.02E-08	7.72E-05	0.0004	0.0221	3.8705	0.2367	0.6850	0.0907	0.1809	0.3918	0.2119	0.1558	0.0465	0.0476	0.0379	0.026	26	26	26	26			
pool, median	Miscellaneous	Miscellaneous	Serine, lipid fraction	1.25	1.05	0.84	0.84	0.0052	0.5281	0.0243	0.0124	0.6251	0.0331	0.0097	0.0207	3.2021	-2.9267	0.0976	-0.077	0.0325	0.0325	0.0265	32	32	32	32					
pool, median	Miscellaneous	Polyamines	Putrescine (additional: Agmatine)	1.19	0.88	0.74	1.36	0.0070	0.0488	2.07E-06	0.93E-07	0.0004	1.41E-09	2.72E-07	0.0007	3.2588	10.5207	7.7162	4.1027	0.1206	0.3985	0.2779	0.1238	0.0370	0.0379	0.0360	0.0302	26	26	26	26
pool, median	Miscellaneous	Polyamines	Spermidine	1.13	0.80	0.71	2.12	0.0292	0.0003	3.70E-07	5.39E-16	0.0543	0.0010	2.07E-06	3.01E-15	2.3085	-4.1577	6.7457	16.7520	0.0255	0.0968	0.1493	0.3273	0.0227	0.0233	0.0211	0.0185	26	26	26	26
<sup>13</sup> C-ISTD, median	Nucleobases and related	Nucleotides	Adenosine diphosphate (ADP)	2.91	0.79	0.27	0.72	5.52E-08	0.1454	2.79E-09	0.0130	5.33E-07	0.2134	2.60E-08	0.0199	7.1760	-1.4950	-3.3149	-0.6239	0.4644	-0.1018	-0.5662	-0.1443	0.0647	0.0681	0.0547	30	30	30	30	
<sup>13</sup> C-ISTD, median	Nucleobases and related	Nucleotides	Adenosine monophosphate (AMP)	0.82	1.00	1.21	0.98	0.1403	0.9723	0.1704	0.8438	0.1957	0.9782	0.1970	0.8788	-1.5147	-0.3550	1.4046	-0.1987	-0.0840	-0.0200	0.0819	-0.0093	0.0554	0.0583	0.0469	30	30	30	30	
<sup>13</sup> C-ISTD, median	Nucleobases and related	Nucleotides	Adenosine triphosphate (ATP)	0.63	0.81	1.26	0.75	3.38E-05	0.0426	0.0177	0.0002	0.0727	0.0250	0.0020	-4.8684	-2.1179	2.5089	-3.6097	-0.2001	-0.0916	0.1085	-0.1254	0.0411	0.0432	0.0347	30	30	30	30		
<sup>13</sup> C-ISTD, median	Nucleobases and related	Nucleotides	Cytidine diphosphate (CDP)	3.35	1.67	0.50	0.66	2.83E-08	0.0056	0.0003	3.09E-07	0.0121	0.0007	0.0079	7.4250	0.2450	0.4742	-0.3045	-0.0365	0.5251	0.2219	-0.3018	0.1871	0.0707	0.0744	0.0598	30	30	30	30	
<sup>13</sup> C-ISTD, median	Nucleobases and related	Nucleotides	Cytidine monophosphate (CMP)	0.69</																											

pool, median	Unknown	Unknown polar	Unknown polar (59970918)	0.26	0.98	3.70	0.89	1.51E-12	0.8686	2.21E-12	0.2285	6.30E-11	0.9199	5.79E-11	0.2798	-11.1428	-0.1668	10.9759	-1.2277	-0.5774	-0.0086	0.5687	-0.0519	0.0518	0.0518	0.0423	32	32	32	32		
pool, median	Unknown	Unknown polar	Unknown polar (59970919)	0.40	0.86	2.14	0.83	1.48E-07	0.2882	3.34E-06	0.0990	1.24E-06	0.3889	1.38E-05	0.1329	-6.6929	-1.0801	5.6129	-1.6990	-0.3939	-0.0636	0.3303	-0.0816	0.0588	0.0588	0.0480	32	32	32	32		
<sup>13</sup> C-ISTD, median	Vitamins, cofactors and related	Acyl-carriers and related	Coenzyme A	0.88	1.87	2.12	1.34	0.3567	0.0001	1.04E-05	0.015	0.4477	0.0005	3.76E-05	0.0233	-0.9360	4.3946	5.2842	2.5661	-0.0549	0.2709	0.3258	0.1274	0.0586	0.0616	0.0616	0.0495	30	30	30	30	
<sup>13</sup> C-ISTD, median	Vitamins, cofactors and related	Acyl-carriers and related	Pantothenic acid	0.59	0.79	1.32	0.73	4.47E-09	0.0012	0.0003	2.50E-06	5.35E-08	0.0033	0.0006	5.56E-06	-8.1306	-3.5803	4.1471	-5.7913	-0.2267	-0.1051	0.1217	-0.1368	0.0279	0.0293	0.0293	0.0236	30	30	30	30	
pool, median	Vitamins, cofactors and related	Amino-carriers and related	Pyridoxine	0.33	1.22	3.67	1.12	5.86E-08	0.2250	1.96E-09	0.4028	5.44E-07	0.3209	1.97E-08	0.4555	-7.0213	1.2372	8.2585	0.8478	-0.4800	0.0846	0.5645	0.0473	0.0684	0.0684	0.0558	32	32	32	32		
pool, median	Vitamins, cofactors and related	Cl-carriers and related	Biotin	0.50	1.44	2.91	1.95	1.39E-06	0.0033	1.05E-09	1.35E-07	9.68E-06	0.0073	1.15E-08	3.40E-07	-6.1028	3.2143	8.9516	6.9845	-0.3034	0.1598	0.4632	0.2894	0.0497	0.0497	0.0518	0.0414	28	28	28	28	
pool, median	Vitamins, cofactors and related	Cl-carriers and related	Folic acid	0.23	0.99	4.29	0.68	5.34E-07	0.9793	5.76E-07	0.0533	3.94E-06	0.9793	3.09E-06	0.0743	-6.2458	-0.0262	6.2197	-2.0065	-0.6350	-0.0027	0.6323	-0.1666	0.1017	0.1017	0.0830	32	32	32	32		
pool, median	Vitamins, cofactors and related	Carotenoids	beta-Carotene	1.26	0.77	0.61	0.37	0.0118	0.0051	2.78E-06	1.90E-15	0.0248	0.0111	1.19E-05	9.75E-15	2.6721	-3.0098	5.6759	-14.2972	0.1000	-0.1124	-0.2124	-0.4369	0.0374	0.0374	0.0306	32	32	32	32		
pool, median	Vitamins, cofactors and related	Redox-carrier and related	Coenzyme Q10	1.21	0.76	0.63	2.52	0.0022	3.90E-05	3.05E-09	1.27E-19	0.0061	0.0002	2.75E-08	1.32E-18	3.3258	-4.2765	8.0934	19.9955	0.0820	-0.1176	-0.1997	0.4020	0.0247	0.0247	0.0201	32	32	32	32		
pool, median	Vitamins, cofactors and related	Redox-carrier and related	Flavine adenine dinucleotide (FAD)	1.74	0.95	0.55	1.83	4.28E-09	0.5118	7.29E-10	5.88E-12	5.35E-08	0.6185	8.75E-09	2.11E-11	7.9681	-0.6635	8.6316	10.5600	0.2416	-0.0201	0.2617	0.2614	0.0303	0.0303	0.0248	32	32	32	32		
pool, median	Vitamins, cofactors and related	Glutathione (GSH)	1.85	1.45	0.79	2.48	0.0110	0.1108	0.2967	2.74E-05	0.0233	0.1696	0.3308	5.78E-05	2.7007	1.6398	-1.0609	4.8895	0.2674	0.1624	-0.1050	0.3953	0.0990	0.0990	0.0990	0.0808	32	32	32	32		
<sup>13</sup> C-ISTD, median	Vitamins, cofactors and related	Redox-carrier and related	Glutathione disulfide (GSSG)	1.17	1.32	1.13	2.03	0.2781	0.2275	0.5870	0.0005	0.5687	0.3217	0.6163	0.0010	0.7184	1.2319	0.5492	3.8726	0.0675	0.1217	0.0543	0.3074	0.0939	0.0988	0.0988	0.0794	30	30	30	30	
pool, median	Vitamins, cofactors and related	Redox-carrier and related	Nicotinamide	3.65	0.94	0.26	1.25	3.04E-12	0.6004	8.99E-13	0.0270	1.09E-10	0.6944	3.24E-11	0.0395	10.8394	-0.5291	-11.3683	2.3174	0.5621	-0.0274	-0.5896	0.0981	0.0519	0.0519	0.0423	32	32	32	32		
pool, median	Vitamins, cofactors and related	Redox-carrier and related	Nicotinamide adenine dinucleotide (NAD)	1.02	1.36	1.34	1.19	0.7505	1.14E-05	2.88E-05	0.0001	0.8121	5.59E-05	8.86E-05	0.0017	0.3208	5.1925	4.8718	3.6374	0.0081	0.1348	0.1264	0.0771	0.0260	0.0260	0.0212	32	32	32	32		
<sup>13</sup> C-ISTD, median	Vitamins, cofactors and related	Redox-carrier and related	Nicotinamide adenine dinucleotide phosphate (NADP)	2.44	1.04	0.43	0.93	7.30E-10	0.7150	5.64E-09	0.4221	1.31E-08	0.7976	4.74E-08	0.4709	8.8481	0.3686	-8.0405	-0.8139	0.3874	0.0170	-0.3700	-0.0301	0.0437	0.0460	0.0460	0.0370	30	30	30	30	
<sup>13</sup> C-ISTD, median	Vitamins, cofactors and related	Redox-carrier and related	Nicotinamide adenine dinucleotide phosphate,	0.95	0.73	0.78	0.73	0.5172	0.0016	0.0078	0.0001	0.6066	0.0041	0.0121	0.0003	-0.6555	-3.4741	-2.8512	-4.3913	-0.0241	-0.1346	-0.1105	-0.1367	0.0368	0.0388	0.0388	0.0311	30	30	30	30	
<sup>13</sup> C-ISTD, median	Vitamins, cofactors and related	Redox-carrier and related	Nicotinamide adenine dinucleotide, reduced (N)	1.66	1.04	0.63	0.49	0.0442	0.8815	0.0748	0.0015	0.0748	0.9275	0.0952	0.0026	2.1004	0.1504	1.8458	-3.4919	0.2203	0.0166	-0.2037	-0.3096	0.1049	0.1104	0.1104	0.0886	30	30	30	30	
pool, median	Vitamins, cofactors and related	Tocopherols and related	alpha-Tocopherol	1.24	1.53	1.07	1.13	0.0926	0.0275	0.5684	0.1426	0.0489	0.5993	0.3000	1.7338	2.3102	0.5764	1.1722	0.0941	0.1254	0.0313	0.0519	0.0543	0.0543	0.0443	32	32	32	32			

Statistical quantities	Explanation
Ratio	ANOVA estimate of effect on ratio scale (transformed Estimate: $10^{\text{Est.log}}$ )
p.value	p-value of ANOVA t-statistics
t.value	ANOVA t-statistics
Est.log	ANOVA contrast estimate (on log10-scale)
Std.Err	Standard error of ANOVA contrast estimate (on log10-scale)
DF	Degrees of freedom for contrast estimate
q.value	Benjamini-Hochberg multiple comparisons q-value (= False-discovery rate/FDR)

Coloring of ratios:
ratio > 1 & p-value < 0.01
ratio > 1 & $0.01 \leq p\text{-value} < 0.05$
ratio > 1 & $0.05 \leq p\text{-value} < 0.10$
ratio < 1 & $0.05 \leq p\text{-value} < 0.10$
ratio < 1 & $0.01 \leq p\text{-value} < 0.05$
ratio < 1 & p-value < 0.01

Font style of ratios:
<b>bold:</b> ( ratio < 0.5 ) or ( ratio > 2 )
normal: ( ratio > 0.5 ) or ( ratio < 2 )

Coloring of p-values:
$0.05 \leq p < 0.10$
$0.01 \leq p < 0.05$
$p < 0.01$

METABOLITE\_NAME contains an "Additional": Quantitation can be disturbed by metabolites exhibiting identical analytical characteristics with respect to quantitation method.

Literature data and/or comparison with alternative methods (e.g. LC-MS/MS, GC-MS) suggest that disturbing metabolite is present at minor levels only.