

Case Report

False Negative Pleural Cytology in Patient with Malignant Pleurisies: Is Pleural EpCAM-Positive Microparticles a Complementary Tool for Diagnosis?

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Abstract

The diagnosis of neoplastic pleurisies remains difficult because of frequent false negative cytological analysis. Thoracoscopy remains the gold standard to distinguish benign from malignant pleural effusion, but it is invasive and not suitable for all patients.

Microparticles (MPs) are promising potential biomarkers and could represent a new approach to identify patients with malignant pleural effusions.

We have recently reported the presence of EpCAM-positive-MPs (EpCAM+MPs) in malignant pleurisies that could be routinely used as a complementary tool with cytology for the diagnosis of pleural malignancy.

Here we describe three cases of pleural cancer patients negative for cancer cells but positive for EpCAM+MPs in the pleural fluid.

This study confirmed the possibility to apply in clinical practice recent data about EpCAM+MPs, detected in pleural fluid, as a non-invasive pleural biomarker useful to increase sensitivity of cytology. In particular, we suggest in patients with a strong suspicion of pleural cancer to search pleural EpCAM+-MPs as a complementary diagnostic tool in case of negative cytological analysis of pleural fluid.

Keywords: Malignant pleural effusion; Thoracoscopy; Microparticles; EpCAM

Introduction

Non-invasive biomarkers to differentiate benign from malignant pleural effusion are needed [1,2]. If thoracentesis is usually the first step to achieve cytological analysis in patients with pleural effusion thoracoscopy remains the gold standard to obtain tissue for final diagnosis [3].

However, cytology can produce false negative results and thoracoscopy is an invasive approach not suitable for all patients.

Microparticles (MPs) are extracellular vesicles released by all cell types and are considered promising potential biomarkers for diagnosis, prognosis, and disease monitoring.

These extracellular vesicles with a size of 0.1-1 micron, originate from blebbing of cell membranes after cell activation or apoptosis. They generally express the anionic phospholipid phosphatidylserine, and membrane antigens derive from their parental cells [4]. Microparticles are also released by tumor cells [5-9]. However, very few data have reported the presence of these MPs in pleural fluid [10-12].

We have recently reported the presence of EpCAM-positive microparticles in pleural fluid that could represent a new approach to non-invasively identify patients with malignant pleural effusions [13].

Here we describe three cases of cancer patients negative for cancer cells but positive for EpCAM-MPs in the pleural fluid after thoracentesis, illustrating a potential interest of this analysis to improve the diagnosis of patients suffering from malignant pleurisy.

Case Reports

Case 1

A 66-year-old man presented with dyspnea and a pleural effusion on the left side at chest radiograph. His past medical history was significant for head and neck cancer. Citation: Roca E, Laroumagne S, Lacroix R, Dutau H, Judicone C, et al. (2017) False Negative Pleural Cytology in Patient with Malignant Pleurisies: Is Pleural EpCAM-Positive Microparticles a Complementary Tool for Diagnosis?. J Mol Biomark Diagn S2: 028. doi: 10.4172/2155-9929.S2-028

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Physical examination revealed a good performance status (PS 1, ECOG classification), no weight loss, and, at the lower part of the left hemithorax, decreased breath sounds, and dullness to percussion. A CT scan showed a left-sided pleural effusion, and no pleural or parenchymal abnormalities were noted. The patient underwent thoracentesis and the cytological analysis did not detect the presence of cancer cells.

metastases and about 2300 ml of hematic pleural fluid were drained (Figure 1a). We searched the presence of EpCAM positive MPs in pleural fluids using highly sensitive flow cytometry. As illustrated in Figure 2a, compared to control, a huge number of MPs positive for EpCAM was detectable in the pleural fluid of this patient (Figure 2 and Case 1).

Given the high suspicion for relapse of cancer disease, thoracoscopy was performed It revealed diffuse pleural nodules suggesting Although the cytology was negative, the histological report of pleural biopsy was consistent with the diagnosis of metastasis from head and neck cancer.



Figure 1: Thoracoscopic features of the three clinical cases.

(Lung: , Parietal Pleura: , Diaphragm:)

a: Case 1- Metastatic pleural cancer with huge nodules and neoplastic lymphangitis.

b: Case 2- Pleural metastasis of prostatic adenocarcinoma.

c: Case 3- Costo-diaphragmatic gutter in patient with breast cancer, multiple nodules are disseminated on the parietal pleural () and on the diaphragm ().

Case 2

A 67-year-old patient visited the outpatient clinic for a right pleural effusion documented at chest radiography. He had never smoked. Physical examination revealed a good performance status (PS-0, ECOG classification), and it was unremarkable. The thoracentesis performed to improve its symptoms showed after biological analysis an exudate without malignant cells. A chest CT scan confirmed right pleurisy without pulmonary or pleural lesions. The patient underwent thoracoscopy for diagnostic and medical purposes. Pleural biopsies were taken and a pleurodesis using dedicated talc was carried out due to the strong macroscopic suspicion of malignant disease (Figure 1b). Compared to control, EpCAM positive MPs were also detected in pleural liquid by cytometry (Figure 2 and Case 2).

The histological diagnosis of pleural biopsy revealed metastasis from adenocarcinoma of probable prostatic origin. A total body CT scan showed a prostate cancer with bone metastasis.

Representative flow cytometry graphs of EpCAM labeling on MPs from pleural fluid. The control experiments with appropriate isotype antibodies are displayed below.



Figure 2: EpCaM-positive microparticles in pleural effusion.

Case 3

A 37-year-old woman with a significant smoking history presented with dyspnea, fatigue, and weight loss. The physical examination revealed dullness to percussion with decreased breath sounds at the lower part of the left hemithorax.

A left-sided pleural effusion was identified on chest X-ray and confirmed by the chest CT without lung or mediastinal abnormalities. Thoracentesis yielded pleural fluid consistent with an inflammatory pleurisy without malignant cells.

Compared to control, EpCAM positive MPs were detected in pleural liquid by cytometry (Figure 2 and Case 3), at a level comparable to patient 2.

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By thoracoscopy 500 ml of citrine pleural liquid were removed and pleural samples were taken (Figure 1c).

Pleural biopsies during the procedure using an optical biopsy forceps confirmed the diagnosis of pleural metastasis from breast cancer. Mammography identifies a nodular lesion at the level of the left-upper-outer lobe strongly suggestive of breast cancer.

Discussion

Cytological analysis has variable diagnostic yields depending on multiple factors and situations. However, according to the recommendations, thoracentesis for sampling the fluid still remains the first step leading to cytological analysis for the diagnosis of pleural cancer. Indeed, in pleurisy, it is largely used to distinguish benign from malignant pleural effusion. Nevertheless, this technique presents some limitations. In fact, the cell poorness, or the difficulties to distinguish tumor cells from reactive or inflammatory mesothelial cells can cause false negative results [14].

At contrary, thoracoscopy allowing pleural biopsies usually achieves the diagnosis between benign and malignant effusion. For this reason, it is the gold standard technique in case of pleural abnormalities (nodules, thickenings...) and pleural effusion after imaging procedures. However, thoracoscopy has some limitations and cannot be applied for all patient. In fact, advanced age, poor performance status, and comorbidities can limit this invasive procedure.

The presence of specific microparticles in the pleural fluid could be considered as promising non-invasive biomarkers beside the cytological analysis of the fluid and can increase the diagnostic yield.

Microparticles can easily be detected from different body fluids, including peripheral blood, urine, cerebrospinal fluid, saliva, or synovial and vitreous fluids [15].

Moreover, MPs contain many antigens also present in the cell of origin [16,17]. For this reason, they could have considered useful as biomarkers for the screening and the diagnosis of a cancer at an early stage [18-23].

In a recent study, we documented the presence of high amounts of normal cells-derived and tumor-derived MPs in pleural fluids [24]. In this study, we found EpCAM+ MPs deriving from cancer cells, which identified patients suffering from carcinoma although the pleural cells were no longer detectable. Therefore, EpCAM+MPs could be considered as promising biomarker complementary to cytology to distinguish benign and malignant pleurisy (Figure 2).

For the first time, we reported here three clinical cases of cancer patients negative for cancer cells at pleural cytology but positive for EpCAM-MPs, illustrating the potential of MPs detection to improve the diagnosis of patients suffering from malignant pleurisy.

These clinical cases show the possibility to apply a non-invasive method, based on the detection of pleural EpCAM-positive MPs, which is complementary to pleural cytological analysis.

Therefore, even if the cytological analysis is negative, the detection of EpCAM-positive MPs could be added to cytology to better discriminate benign from malignant pleural effusions. The positivity of MPs in patient with negative cytology could be explained by the diffusion properties of the vesicles in the pleural fluid. In addition, the fact those MPs, deriving from apoptotic cancer cells, remain present even if parental cells are no longer detectable because of the process of apoptosis. Moreover, in contrast to cytology it is important to remember that flow cytometry is an operator-independent technique allowing counting MPs.

The MPs also contain protein nucleic acids and therefore carry a genetic signature and could be implicated in carcinogenesis. In particular, the MPs have a role in the progression of the disease, in cellular function and in genetic regulation. Further molecular biology techniques applied on MPs of neoplastic pleurisies could be interesting to study their genetic signature and their carcinogenic potential. Therefore, this present report opens new perspectives about the utility of MPs from pleural fluid. Moreover, it offers the prospect of designing new therapeutic approaches.

In particular, since anti-EpCAM target therapy are available [25], it could be of interest to identify patients with EpCAM-positive MPs to define more personalized therapeutic approaches [26].

Conclusion

We have recently reported that EpCAM+MPs detected in the pleural fluid are correlated with the presence of neoplasia and in particular, with adenocarcinoma type. These reported cases illustrate how this marker can complete the usual cytological analysis of the pleural fluid.

Consequently, pleural EpCAM+MPs combined with pleural cytological analysis could be relevant for poor performance status patient for whom more invasive procedure is not available in case of suspicion of pleural cancer.

Moreover, since cancerous MPs have also several activities and genetic signatures related to malignant responses, EpCAM+ MPs could be involved in carcinogenesis of the pleural microenvironment. Finally, EpCAM+MP detectable in pleural fluid could act as a target for specific treatment as already reported in others cancers [27,28].

References

- 1. Rodriguez-Panadero F, Borderas Naranjo F, Lopez Mejias J (1989) Pleural metastatic tumours and effusions. Frequency and pathogenic mechanisms in a post-mortem series. Eur Respir J 2 366-369.
- 2. Light RW (2011) Pleural effusions. Med Clin North Am 95: 1055-1070.
- 3. Skalski JH, Astoul PJ, Maldonado F (2014) Medical thoracoscopy. Semin Respir Crit Care Med 35: 732-743.
- Burnier L, Fontana P, Kwak BR, Angelillo-Scherrer A (2009) Cell-derived microparticles in haemostasis and vascular medicine. Thromb Haemost 101: 439-451.
- De Broe ME, Wieme RJ, Logghe GN, Roels F (1977) Spontaneous shedding of plasma membrane fragments by human cells in vivo and in vitro. Clin Chim Acta 81: 237-245.
- Berckmans RJ, Nieuwland R, Tak PP, Boing AN, Romijn FP, et al. (2002) Cell-derived microparticles in synovial fluid from inflamed arthritic joints support coagulation exclusively via a factor VII-dependent mechanism. Arthritis Rheum 46: 2857-2866.
- Van Blitterswijk WJ, Emmelot P, Hilkmann HA, Hilgers J, Feltkamp CA (1979) Rigid plasma-membrane-derived vesicles, enriched in tumourassociated surface antigens (MLr), occurring in the ascites fluid of a murine leukaemia (GRSL). Int J Cancer 23: 62-70.
- Rupp AK, Rupp C, Keller S, Brase JC, Ehehalt R, et al. (2011) Loss of EpCAM expression in breast cancer derived serum exosomes: role of proteolytic cleavage. Gynecol Oncol 122: 437-446.
- 9. Freyssinet JM (2003) Cellular microparticles: what are they bad or good for? J Thromb Haemost. 1: 1655-1662.

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- Bard MP, Hegmans JP, Hemmes A, Luider TM, Willemsen R, et al. (2004) Proteomic analysis of exosomes isolated from human malignant pleural effusions. Am J Respir Cell Mol Biol 31: 114-121.
- 11. Park JO, Choi DY, Choi DS, Kim HJ, Kang JW, et al. (2013) Identification and characterization of proteins isolated from microvesicles derived from human lung cancer pleural effusions. Proteomics 13: 2125-2134.
- 12. D'Souza-Schorey C, Clancy JW (2012) Tumor-derived microvesicles: shedding light on novel microenvironment modulators and prospective cancer biomarkers. Genes Dev 26: 1287-1299.
- 13. Roca E, Lacroix R, Judicone C, Laroumagne S, Robert S, et al. (2016) Detection of EpCAM-positive microparticles in pleural fluid: A new approach to mini-invasively identify patients with malignant pleural effusions. Oncotarget 19: 3357-3366.
- Bedrossian CW (1998) Diagnostic problems in serous effusions. Diagn Cytopathol 19: 131-137.
- 15. Piccin A, Murphy WG, Smith OP (2007) Circulating microparticles: Pathophysiology and clinical implications. Blood Rev 21: 157-171.
- Taylor DD, Gercel-Taylor C (2008) MicroRNA signatures of tumorderived exosomes as diagnostic biomarkers of ovarian cancer. Gynecol Oncol 110: 13-21.
- 17. Zocco D, Ferruzzi P, Cappello F, Kuo WP, Fais S (2014) Extracellular vesicles as shuttles of tumor biomarkers and anti-tumor drugs. Front Oncol 4: 267.
- Logozzi M, De Milito A, Lugini L, Borghi M, Calabro L, et al. (2009) High levels of exosomes expressing CD63 and caveolin-1 in plasma of melanoma patients. PLoS One 4: e5219.
- Mizutani K, Terazawa R, Kameyama K, Kato T, Horie K, et al. (2014) Isolation of prostate cancer-related exosomes. Anticancer Res 34: 3419-3423.

- Mitchell PJ, Welton J, Staffurth J, Court J, Mason MD, et al. (2009) Can urinary exosomes act as treatment response markers in prostate cancer?. J Transl Med 7: 4.
- Khan S, Jutzy JM, Valenzuela MM, Turay D, Aspe JR, et al. (2012) Plasma-derived exosomal survivin, a plausible biomarker for early detection of prostate cancer. PLoS One 7: e46737.
- 22. Skog J, Wurdinger T, Van Rijn S, Meijer DH, Gainche L, et al. (2008) Glioblastoma microvesicles transport RNA and proteins that promote tumour growth and provide diagnostic biomarkers. Nat Cell Biol 10: 1470-1476.
- Shao H, Chung J, Balaj L, Charest A, Bigner DD, et al. (2012) Protein typing of circulating microvesicles allows real-time monitoring of glioblastoma therapy. Nat Med 18: 1835-1840.
- 24. Roca E, Lacroix R, Judicone C, Laroumagne S, Robert S, et al. (2016) Detection of EpCAM-positive microparticles in pleural fluid: A new approach to mini-invasively identify patients with malignant pleural effusions. Oncotarget 19: 3357-3366.
- 25. Berek JS, Edwards RP, Parker LP, DeMars LR, Herzog TJ, et al. (2014) Catumaxomab for the treatment of malignant ascites in patients with chemotherapy-refractory ovarian cancer: a phase II study. Int J Gynecol Cancer 24: 1583-1589.
- 26. Bokemeyer C (2010) Catumaxomab-trifunctional anti-EpCAM antibody used to treat malignant ascites. Expert Opin Biol Ther 10:1259-1269.
- 27. Sebastian M (2010) Review of catumaxomab in the treatment of malignant ascites. Cancer Manag Res 2: 283-286.
- Eskander RN, Tewari KS (2013) Epithelial cell-adhesion moleculedirected trifunctional antibody immunotherapy for symptom management of advanced ovarian cancer. Clin Pharmacol: 55-61.

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