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EGFR Primary T790M and L858R Double Mutation Confers Clinical Benefit to Erlotinib and Resistance to Osimertinib in One Lung Adenocarcinoma Patient: A Case Report

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Case Report

Abstract

Introduction: EGFR TKI has been widely studied in both research and clinic. However, only few limited studies focus on EGFR primary resistance rather than acquired mutation. Usually, EGFR T790M mutation is resistant to first generation TKI (erlotinib, gefitinib) and sensitive to third generation TKI Osimertinib. Here, we report an EGFR primary T790M and L858R double mutation patient confers clinical benefit to Erlotinib and resistance to Osimertinib.

Case presentation: Here we present a 70 year-old woman with lung adenocarcinoma harboredprimary T790M and L858R EGFR double mutation and underwent multiple lines of treatments. She benefited from the first-generation of EGFR TKI Erlotinib, and later quickly developed resistance to the third-generation TKI Osimertinib after disease progression, then followed a few rounds of chemotherapies. Multiple resistant EGFR mutations were detected, indicating frequent complex tumor heterogeneity in later stage patients caused by subclone evolution. Colonies with distinct Osimertinib resistance mutations included well known EGFR mutations L792V, L718V/R, G796S/A, and novel EGFR G729V and D1014V mutations, which are predicted to be acquired osimertinib resistance mutations by 3D-structural remodeling.

Conclusion: Our study revealed that EGFR primary T790M accompany with L858R mutation could benefit from erlotinib, and has stable disease for 7 months. The patient also benefited from chemotherapy for three months after resistance to osimertinib. ctDNA–based assay is a valuable tool for late stage cancer diagnosis, monitoring and intervention.

Keywords: EGFR; Erlotinib; Osimertinib; Lung adenocarcinoma; Circulating tumor DNA assay

Case Presentation

Abbreviations: EGFR: Epidermal Growth Factor Receptor; TKI: Tyrosine Kinase Inhibitors; NSCLC: Non-Small Cell Lung Cancer; ARMS-PCR: Amplification Refractory Mutation System Polymerase Chain Reaction; ddPCR: Droplet Digital Polymerase Chain Reaction; CT: Computerized Tomography; PET-CT: Positron Emission Tomography–Computed Tomography; ctDNA: Circulating tumor DNA; PFS: Progression-Free Survival; CEA: Carcinoembryonic Antigen

Introduction

The epidermal growth factor receptor (EGFR)-tyrosine kinase inhibitors (TKIs) include the previously widely used first-generation gefitinib and erlotinib, the second-generation afatinib and nowadays the third-generation osimertinib with remarkable activity in nonsmall cell lung cancer (NSCLC) patients [1,2]. The sensitive biomarkers, primary and acquired resistant mechasnism were summarized in Table 1 from literature [3-8]. Minor clones with the T790M mutation in treatment-naive tumors that contain classic sensitizing mutations were under estimated by conventional PCR and Sanger sequencing methods. Studies using more sensitive techniques have reported T790M mutations in 35% and 79% of EGFR-mutant, NSCLC pretreatment specimens and the T790M mutation in some cases were associated with longer progression-free survival to erlotinib [9,10]. Here we describe one patient who had primary L858R and T790M double mutation was sensitive to erlotinib and resistant to osimertinib, finally benefited from chemotherapy for three months after resistance to osimertinib.

A 63-year-old famale, with ECOG performance status of 1 was diagnosed in June 2010 to be lung adenocarcinoma stage IB(pT2N0M0). The patient had no smoking history, and no family history of malignant tumor. ARMs PCR detected both L858R and T790M mutations in the tumor sample. Droplet digital PCR(ddPCR) for the same FFPE slides confirmed the initial ARMs PCR results with allele frequency 0.254 for L858R mutation and 0.0021 for T790M mutation (Supplementary Table 1). Adjuvant chemotherapy with 4 cycles of gemcitabine and Cisplatin were administrated after surgical resection (left pneumonectomy). No recurrence was seen until September, 2014.

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Generation	Drug	Sensitive biomarkers	Prin	nary resistance mechanism	Ac	cquired resistance mechanism
1 st	Gefitinib	EGFR L858R			1.	Secondary EGFR mutations, major in EGFR T790M.
	Erlotinib	EGFR ex19del	1. 2. 3. 4. t	EGFR Exon 20 insertions or duplications. EGFR Exon 20 T790M mutation. EGFR germline BIM deletion. EGFR germline T790M mutation (rare case report).	2. 3. 4.	Gene copy alterations of alternative pathways including
2 nd	Afatinib	EGFR L858R				MET or ERBBZ or MAPK amplification. Mutations in downstream effector molecules of EGER
		EGFR ex19del				including KRAS, NRAS, BRAF V600E, PIK3CA
		EGFR wildtype (Relative potent compared to 1 st generation TKI)				mutation. Other mechanism including EMT, SCLC transformation, AXL or NF-kb activation et al.
3 rd	Osimertinib	EGFR L858R	- 1. 2. 3.	EGFR Exon 20 insertions or duplications. EGFR germline BIM deletion. SCLC transformation.	1. 2. 3. 4.	Secondary EGFR mutations, major in EGFR C797S/R,
		EGFR ex19del				L798I, G796S/R, L792F/H.
		EGFR L858R+T790M				MET or ERBB2 or MAPK amplification.
		EGFR ex19del+T790M				Mutations in downstream effector molecules of EGFR including KRAS, NRAS, BRAF V600E, PIK3CA mutation. Other mechanism including EMT, SCLC transformation, Src-AKT, Src-YAP1 activation et al.

Table 1: Sensitive biomarkers, primary and acquired resistance mechanism for EGFR three generation TKIs.



RECIST standard was used for assessment of response.

In September 2014, computed tomography (CT) scan revealed multiple lung nodules. No metastasis was seen by abdominal ultrasound examination and magnetic resonance imaging (MRI). In Feburary 2016, positron emission tomography (PET)-computed tomography (CT) discovered the left clavicular upper and mediastinal multiple lymph node metastasis. She was treated with Erlotinib treatment since then, and stable disease was confirmed according to RECIST assessment. The patient had 7-month progression-free survival (PFS) (Figure 1). In September 2016, a nodular pattern in the left lung was emerged, and AZD9291 (trial agent at that time) was taken. In November 2016, the disease was progressed. From November 2016, the patient had then been treated for six cycles of combination of Pemetrexed 0.8g dwith Cis-Paclitaxel 40 mg d1-3,q3w. The patient had partial response after two cycles of Pemetrexed/Cisplatin (PP) treatment and stable disease after four cycles. The disease control time is about 3 months. In April 2017, the cancer metastasized into brain and later liver. A plasma-based circulating tumor DNA (ctDNA) assay was performed in June 2017. The multiple heterogeneous EGFR mutations, such as L858R and T790M were detected and these mutations were also observed by ARMs PCR in the initial test. In addition, more EGFR mutations, such as L792V, G796A/S and L718V/R were also detected and those are well

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known mutations to resist 3rd TKI. At the same time some unknown mutations such as G729V, P936S/A/T, D1014V, R334H, S1081N, R677H were discovered. L792V and G796A/S are in trans. However

all mutations are in cis with T790M (Figure 2). Molecular modeling predicted G729V and D1014V could be acquired osimertinib resistance mutations (Figure 3).

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Discussion

De novo T790M mutation was considered as a well-known primary resistant mechanism to EGFR first generation TKI [3,5,11] and is about 1% to 8% of NSCLC patients with an activating EGFR mutation [12]. One meta-analysis result shows that primary (pretreatment) T790M mutation was associated with worse PFS and OS in patients with advanced NSCLC treated with 1st EGFR-TKIs [13]. However, Roseall et al. reported that Low BRCA1 levels neutralized the negative effect of the T790M mutation and were associated with longer progressionfree survival to erlotinib [9]. In the Iressa Pan-Asia Study (IPASS), 11 patients were identified to have primary T790M mutations using an ARMS PCR. Of 11 patients, 7 had concurrent activating EGFR mutations, and 4 of these received gefitinib. Three demonstrated a partial response, while one had stable disease, suggesting that patients with double mutations were sensitive to first generation TKI [14]. In this case, the patient with T790M and L858R mutations was initially treated with erlotinib and had 7-month PFS. T790M mutation clone was very low compared to sensitive L858R mutation in the primary tissue, while T790M clones were expanded after progression showed by Arms PCR.

Then the patient become quickly resistant to the third-generation TKI osimertinib. We extrapolated that low osimertinib dosage might be one of the possible reasons for osimertinib primary resistance, especially in this case EGFR amplification was detected in patient ctDNA samples. In the AURA study, sixty treatment-naive patients with locally advanced or metastatic EGFRm NSCLC received osimertinib 80 or 160 mg once daily (30 patients per cohort). We can see that Overall ORR was 67% (95%CI, 47% to 83%) in the 80 mg group, 87% (95% CI, 69% to 96%) in the 160 mg group, suggest that low osimertinib dosage has lower ORR [15]. The well known variants to resist to EGFR 3rd TKI are C797S, L798I, L692V, E709K mutation and ERBB2, MET amplification [16-18]. Several case reports also showed some new EGFR resistant mutations like G796D [19], G724S [20], L718Q [21]. Besides these major resistant mutations, multi-clone

heterogeneous resistance was also reported [22,23]. In this patient, ctDNA assay detected several known osimertinib resistant mutations such as EGFR G796A/S, L792V. Molecular modeling demonstrated that G796A/S directly clashes (red lines) into the methoxybenzene ring of osimertinib while L792V clashes into the V742 of the VAIK motif and destabilizes the ATP binding pocket (Figure 3). Thus the green ones (T790M, G796A/S, L792V) within the pocket could be the major drivers for the drug resistance [19,22]; while two yellow ones (G729V and D1014V) are line on the boundary, could be the secondary drivers for the drug resistance. As for L718V/R, these mutations have been reported as a resistance mutations identified in patients who progressed on third-generation EGFR TKI [21]. Transformation to SCLC has also been described recently as a resitant mechanism during osimertinib treatment in at lease 7 clincial cases [4,24]. TP53 and RB1 double loss of function mutation is a common molecular pattern in SCLC, while in our case, we did not see clear evidence of TP53 and RB1 double loss of function mutation or SCLC pathology evidence.

After developing osimertinib resistancy, this patient still benefited from Pemetrexed/Cisplatin (PP) chemotherapy for 3 months. Multiple minor MAF unknown significance mutations were listed in this patient while all the clinical treatments were mostly depleted. The heterogeneity of EGFR mutations at the later stage may attribute to her resistance to all available treatments. Findings in this report are promising for applying multidisciplinary approach to treating cancer patient in clinical practice.

Conclusion

This report provides insight into a successful empirical treatment with erlotinib for a lung adenocarcinoma patient in the presence of pretreatment T790M at small colony. Furthermore, this case report a primary resistant of osimertinib treatment with T790M and L858R double mutation. After osimertinib resistance, the patient received clinical benefit for three months with Pemetrexed/Cisplatin chemotherapy in the situation that no further treatment is available.

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Ethics Approval and Consent to Participate

The study was reviewed and approved by the "The First Affiliated hospital of Xi'an Jiaotong University Ethics committee", and the project implementation process was in line with the ethical principples. The patient provided written informed consent to participate in the study.

Consent for publication

We obtained written informed consent of the patient for the publication of the case report and accompanying images.

Competing interests

Yizhou Ye, Hua Dong, Lin Shi, Dalei Wang and Fugen Li are employees of 3DMed, Inc. Zheng Wang is employee of AstraZeneca China. The remaining authors declare no competing interest.

Availability of data and material

The data generated and/or analyzed during this study are included in this published article and the additional files.

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Authors' Contributions

ZX, DH and YY are responsible for generating and analyzing the data, and drafting the manuscript; ZL, WL and CZ for patient sample collection; WZ, SL and WD for related data confirmation and interpretation; YJ, LP and ZX for pathology review of the patient tissues; YY, DH, LF and YJ for study design and technical discussion; drafting and finalizing the manuscript. All authors have read and approved the final version.

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