

Effect of Seasonal Malaria Chemoprevention (SMC) with Sulfadoxine-Pyrimethamine (SP) and Amodiaquine (AQ) on the Acquisition of anti-AMA1 and anti-MSP1₄₂ Antibodies among Children under 10 Years Living in the Southern part of Senegal (Velingara)

Khadime Sylla^{1*}, Roger Clément Kouly Tine¹, Doudou Sow¹, Magatte NDiaye¹, Aissatou Sarr¹, Marie Louise Tshibola Mbuyi², Ibrahima Diouf¹, Jean Louis Abdourahim Ndiaye², Daouda NDiaye¹, Oumar Gaye¹ and Babacar Faye¹

¹Service of Parasitology-Mycology, Pharmacy and Dentistry, University Cheikh Anta Diop, Dakar, Senegal

²Department of Parasitology-Mycology, University of Health Sciences, Libreville, Gabon

Abstract

Background: In developing countries, malaria is still a leading cause of morbidity and mortality and children are the most affected individuals. In order to strengthen malaria control, new intervention such as Seasonal Malaria Chemoprevention (SMC) has been developed. This strategy is very effective in preventing malaria clinical episodes but its effect on children's immunity is not well documented. This study aimed to evaluate the effects of SMC on the acquisition of anti-AMA1 and anti-MSP1₄₂ antibodies among children fewer than 10 years living in the southern part of Senegal (Velingara).

Patients and methods: The study was nested in a cluster randomized trial assessing the impact of SMC with a single dose of Sulfadoxine-Pyrimethamine (SP) and 3 doses of Amodiaquine (AQ). Two cross-sectional surveys were carried out (October 2010) and (September 2011) to assess the effect of SMC on children's immunity. Thick and thin blood smears were performed to assess malaria parasitemia prevalence. Blood was collected on filter paper for serological measurement by ELISA to measure IgG anti-MSP1₄₂ and anti-AMA1. Logistic regression analysis was performed to assess factors associated with the production of antibodies.

Results: A total number of 1611 children under 10 years old were included in two surveys (866 children in 2010 and 745 children in 2011). Malaria prevalence was 10.39% at baseline (2010) and 5.03% one year after intervention (2011). The seroprevalence of anti-MSP1₄₂ anti-AMA1 antibodies was higher in 2010 compared to 2011 providing a significant reduction of IgG production at 11.4 AU (95%CI [8.3-14.4]) for MSP1₄₂ and 7.2 AU (95%CI [4.5-9.9]) for AMA1. Seroprevalence increased with age and *Plasmodium falciparum* carriage while it decreased according to the area and study period.

Conclusion: SMC is an effective strategy for malaria prevention in children under 10 years. The strategy can as well induce a decrease of IgG anti-AMA1 and anti-MSP1₄₂ which are protective against malaria. Consequently, this strategy needs to be renewed each year in areas where malaria is highly seasonal to avoid a resurgence of malaria, while promoting the use of other antimalarial interventions.

Keywords: Malaria; *Plasmodium falciparum*; SMC; Children; Immunity; Senegal

Introduction

Malaria is still a leading cause of morbidity and mortality despite all the efforts made to control the disease. Over 80% of malaria cases and 90% of malaria deaths occur in Africa and mainly in children [1]. In order to strengthen the fight against malaria in children, a new preventive strategy was recently developed: intermittent preventive treatment currently called Seasonal Malaria Chemoprevention (SMC). This strategy is defined as the administration of therapeutic doses of antimalarial drugs at monthly interval during malaria transmission period in areas where malaria is endemic. Several studies in Africa have shown that this intervention, cost-effective, safe, and feasible for the prevention of malaria among children in areas with highly seasonal malaria transmission. In Senegal, Cissé et al. showed 86% of reduction of malaria incidence among children who received seasonal intermittent preventive treatment [2]. A study in Mali showed a 67.5% efficacy of IPT of 67.5% against clinical malaria episodes [3]. In Ghana, Kweku et al. found 69% efficacy of in terms of reducing malaria incidence [4]. In Tanzania, Schellenberg et al. showed a protective effect of 36% of intermittent preventive treatment in children [5]. WHO adopted in March 2012, Seasonal Malaria Chemoprevention (SMC) previously

referred to as intermittent preventive treatment for malaria prevention strategy in children living in the Sahel sub in regions of Africa [6,7].

This strategy is certainly effective, but it may induce an effect on immunity by reducing production of malaria antibodies such as anti-MSP1₄₂ (Merozoite Surface Protein) and anti-AMA1 (Apical Membrane Protein) which are associated with protection against malaria. Immunization with these antigens provides protection against

***Corresponding author:** Khadime Sylla, Service of Parasitology-Mycology, Faculty of Medicine, Pharmacy and Dentistry, University Cheikh Anta Diop, Dakar, Senegal, Tel: +221338251998; E-mail: khadimesylla@yahoo.fr

Received December 29, 2016; **Accepted** February 10, 2017; **Published** February 17, 2017

Citation: Sylla K, Tine RCK, Sow D, Diaye MN, Sarr A, et al. (2017) Effect of Seasonal Malaria Chemoprevention (SMC) with Sulfadoxine-Pyrimethamine (SP) and Amodiaquine (AQ) on the Acquisition of anti-AMA1 and anti-MSP1₄₂ Antibodies among Children under 10 Years Living in the Southern part of Senegal (Velingara). Malar Chemoth Cont Elimination 6: 155. doi: 10.4172/2470-6965.1000155

Copyright: © 2017 Sylla K, et al. This is an open-access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

malaria. Several studies have shown that SMC decreased the production of protective antibodies against malaria [8,9].

This study was conducted to evaluate the effects of SMC with Sulfadoxine-Pyrimethamine and (SP) and Amodiaquine (AQ) on the acquisition of anti-AMA1 and anti-MSP1₄₂ antibodies in children under 10 years living in the southern part of Senegal (Velingara).

Methodology

Study area

This study was carried out in Velingara health district located in the South-eastern part of Senegal, 500 km from the capital city of Dakar. In this district the study was conducted in Bonconto health post headed by a nurse and has 8 functional health huts staffed with community health workers, serving a total population of 10,016 inhabitants. Malaria transmission is seasonal during rainy season (from July to November) with a peak between Octobers to November. *Plasmodium falciparum* is the most predominant parasite species. Malaria control strategies implemented by the National Malaria Control Program (NMCP) were represented by the case management of uncomplicated malaria cases using RDTs and ACTs; intermittent preventive treatment in pregnant women; universal coverage of long lasting insecticide treated net and indoor residual spraying.

Study design

The study was nested in a cluster randomized trial assessing the impact of SMC with a single dose of Sulfadoxine-Pyrimethamine (SP) and 3 doses of Amodiaquine (AQ) on the incidence of malaria clinical episodes, malaria parasitaemia and anaemia prevalence at the end of transmission season. Children in intervention area were assigned to receive SMC plus community case management of malaria (CCm) while in the control area children had only access to CCm. Details for the cluster randomized trial procedures are described previously [10]. For the immunological assessment, a controlled before and after study was performed. Two cross sectional surveys were conducted, one in October 2010 (baseline) and a second survey a year after intervention period in September 2011. Ends points included (i) sero-prevalence of anti-MSP1₄₂ from baseline to end line both in intervention and control areas, (ii) seroprevalence of anti-AMA1 from baseline to end line both in intervention and control areas, (iii) malaria parasitaemia prevalence at cross sectional surveys.

Data collection method

A questionnaire was administered to collect individual's socio-demographic data (age, gender, weight, height, area of residence), clinical information, and access to antimalarial interventions such as bed net. Anthropometric data were collected as previously described [11]. Blood samples were collected using finger prick blood for malaria diagnostic, haemoglobin and antibody measurement. Haemoglobin level was measured using Hemo-Cue machine (Hemocue® Hb 301). Anemia was defined as Hb concentration below 11 g/dl.

Evaluation of anti-*Plasmodium falciparum* IgG antibodies by ELISA

Three drops of blood were collected onto Whatman 3MM filter paper, which was sealed and stored dry with desiccant at room temperature. Reconstituted sera were obtained from filter paper blood spots described elsewhere [12,13].

Sera were tested for anti-MSP1₄₂ IgG antibodies and anti-AMA1 IgG antibodies by indirect ELISA. Apical membrane antigen (AMA1)

was from the *Pichia pastoris* expressed ectodomain of *Plasmodium falciparum* FVO strain comprised amino acids 25–545 [14]. MSP1₄₂ protein was from the C-terminal MSP1₄₂ amino acid sequence of the Uganda-Palo Alto (FUP) *P. falciparum* isolate expressed in *Escherichia coli* (*Ec*) system [15]. Samples were also tested on freeze thawed *P. falciparum* Schizont Extract (concentration of 1×10^8 /ml), which was coated onto ELISA plates at 1/500.

Briefly, 96 well ELISA plates were coated with 100 μ l/well of 0.1 μ l/well of MSP1 and 0.026 μ l/well of AMA1 in coating buffer (1.59 g Na₂CO₃, 2.93 g NaHCO₃, 1 liter distilled water, pH 9.4). The plates were incubated overnight at 4°C. After incubation, plates were washed at three times using PBS (5.7 g NaH₂PO₄, 16.7 g Na₂HPO₄, 85 g NaCl in 1 liter distilled water) plus 0.05% Tween 20 (PBS/T) and blocked with 1% (w/v) skimmed milk powder in PBS/T for one hour at 37°C. Eluates were removed from 4°C just before use. Following three more washes, eluates were diluted at a ration 1/100 in PBS/T and added 100 μ l in duplicate in a well plate.

For each plate three types of control were used: deep well without serum but with a second antibody to measure the non-specific binding, pool of sera from patients with *Plasmodium falciparum* malaria (positive control) and pool of sera from non-infected subjects (negative control) from Copenhagen. Plates were incubated one hour at 37°C. After three more washes 100 μ l of horseradish peroxidase-conjugated rabbit anti-human IgG (SouthernBiotech) (1/5000 in PBS/T) was added to all wells.

After incubation for one hour at 37°C, plates were developed with TMB/E (Upstate, Chemicon et Linco, Millipore) as substrate for 30 minutes at room temperature and the reaction was stopped by the addition of 50 μ l/well of 2M H₂SO₄. Optical density was read at 450 nm against a 620 nm with an ELISA TECAN SUNRISE reader.

Statistical Methods

Sample size calculation

For each cross sectional survey the total number of children to examine was calculated at 800, based on a prevalence of malaria parasitaemia at 20% in the study area (Senegal MIS 2009) a confidence level at 95% with a precision of 5%, power level at 90% and assuming a percentage of 20% of withdrawal.

Data management and data analysis

Data were entered in Excel software and analysis was performed using Stata software version IC 12.1 software. For serological assessment, the optical density was obtained by subtracting the average OD of duplicate wells from that of the corresponding blank wells. Values were converted into arbitrary units (AUs), as previously described [16]. Quantitative variables were described in terms of means, standard deviation. Inter group comparisons were done using ANNOVA test or Student *t* test where appropriate; otherwise non parametric tests such as Man withney or Kruskall Wallis) were used. For descriptive data, percentage was used to each outcome. Antibodies seroprevalence was calculated and expressed by percentage with their 95% confidence intervals. Proportions were compared using chi-square test or the Fisher exact test (univariate analysis). A stepwise logistic regression analysis was done to assess factors associated with *Pf* antibodies carriage. Significance level of the different tests was set at 5%.

Ethical Considerations

Informed consent was required prior the participation in the

study. Ethical approval was obtained from the Senegalese National Ethical Committee (**Conseil National d’Ethique et de Recherche en Santé**). Approval number 027/MSP/DS/CNRS, 18/03/2010. The study was registered at the Pan African Clinical Trial Registry: registration number: PACTR201305000551876.

Results

Baseline characteristics of study population

A total of 1611 children under 10 years were included in this study (866 children in 2010 and 745 children in 2011); 450 children in 2010 and 470 children in 2011 received SMC while 416 children in 2010 and 275 children in 2011 were included in control area.

The mean age of study participants was 4.5 ± 2.7 years and 4.02 ± 2.3 years respectively in 2010 and in 2011. Study population was mainly represented by children over 5 years old (60.64%) in 2010 and children under 5 years (56.16%) in 2011. The sex ratio was 0.95 in 2010 and 1.01 in 2011.

The mean hemoglobin level was 8.5 ± 3.4 g/dl and 9.3 ± 1.8 g/dl respectively in 2010 and 2011. The prevalence of anemia (Hb <11 g/dl) was 77.14 % and 77.81% respectively in 2010 and 2011.

Prevalence of stunting, underweight and wasting in 2010 was respectively 35.44%, 26.65% and 10.51%. In 2011, stunting, underweight and wasting represented 33.02%, 27.66% and 10.08% (Table 1).

Malaria prevalence

Overall *P. falciparum* malaria prevalence was 10.39% in 2010 and 5.03% in 2011. At baseline study (2010), malaria prevalence was higher in control zone (11.78%) than in intervention zone where it was 9.11%. The difference was not significant ($p=0.19$).

In 2011, *P. falciparum* malaria prevalence was 3.27% in the control area and 6.03% in the intervention area ($p=0.09$) (Figure 1).

Anti-plasmodium IgG responses

In 2010 the level of IgG anti-MSP1₄₂ and IgG anti-AMA1 was respectively 23.2 AU and 17.2 AU while in 2011 it was 11.8 AU and 9.9 AU for both antibodies.

Comparing 2010 and 2011, these results showed a decrease of IgG anti-MSP1₄₂ and IgG anti-AMA1. The difference was significant ($p<10^{-3}$). The mean difference of IgG anti-MSP1₄₂ and IgG anti-AMA1 between 2010 and 2011 was respectively 11.4 AU (95% CI(8.3-14.4)) and 7.2 AU (95% CI(4.5-9.9)). At baseline (2010), in 2010, the level of IgG anti-MSP1₄₂ and anti-AMA1 was respectively 25.2 AU and 22.1 AU in SMC area while it was respectively 17.9 AU and 10.6 AU in the control area ($p<10^{-3}$). A year after intervention 2011, the level of IgG anti-MSP1₄₂ and anti-AMA1 was lower in SMC area compared to the control area (Table 2). The difference was significant ($p<10^{-3}$) (Table 3).

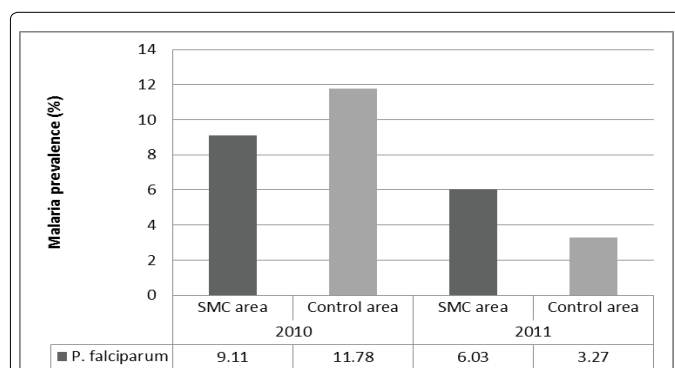
In control area a decrease of IgG anti-MSP1₄₂ was noted between 2010 and 2011. The mean difference was 7.2 AU ($p<10^{-3}$). An increase of IgG anti-AMA1 was noted between 2010 and 2011. The mean difference was -1.3 AU ($p=0.38$).

In SMC area, a significant decrease of IgG anti-AMA1, and IgG anti-MSP1₄₂ was not observed between 2010 and 2011 ($p<10^{-3}$). Mean differences observed were 12.713 AU and 0.4 AU respectively both antibodies (Table 4).

Overall, the seroprevalence rate of anti-MSP1₄₂ and anti-AMA1

	2010 (N=866)	2011 (N=745)
Zone		
SMC (+)	450	470
SMC (-)	416	275
Mean age (year)	4.5 ± 2.7	4.02 ± 2.3
Age group		
< 1 year	86 (58.9%)	83 (11.23%)
(1-4 years)	344 (34.4%)	415 (56.16%)
(5-10 years)	436 (60.64%)	241 (32.61%)
Gender		
Female	443 (51.15%)	341 (46.14%)
Male	423 (48.85%)	345 (46.68%)
Missing	-	53 (7.17%)
Hb mean (g/dl)	8.5 ± 3.4	9.3 ± 1.8
Anemia (Hb<11 g/dl)		
Yes	668 (77.14%)	575 (77.81%)
No	198 (19.8%)	164 (22.19%)
Nutritional status		
Stunting	661 (35.44%)	530 (33.02%)
Underweight	497 (26.65%)	444 (27.66%)
Wasting	196 (10.51%)	210 (13.08%)

Table 1: Characteristics of study population.



Note: *Pf* malaria prevalence in 2010 and 2011 in SMC area and Control area. Malaria prevalence was more important in 2010 compared to 2011. After intervention *P. falciparum* carriage was higher in SMC area compared to Control area. χ^2 test was used to compare the prevalence between two areas. The difference was not significant ($p = 0.19$). Black: SMC area, Grey: Control area.

Figure 1: *Pf* malaria prevalence in 2010 and 2011 in SMC and control area.

antibodies was higher in 2010 (at baseline) compared to 2011 (a year after intervention). Anti-MSP1₄₂ and anti-AMA1 antibody prevalence was respectively 53.12% and 46.3% in 2010. While in 2011 the seroprevalence of both antibodies was 20.03% and 19.8%.

Regarding the area (SMC area / Control area) the seroprevalence of anti-MSP1₄₂ and anti-AMA1 antibody was higher in control zone. In 2010, the seroprevalence of both antibodies was respectively 62.76% and 54.81% in the intervention area while it was respectively 44.22% and 38.44% in the control area. The difference was significant ($p<10^{-3}$). In 2011, a year after SMC implementation, the seroprevalence rate of anti-MSP1₄₂ and AMA1 antibodies was 24.36% and 22.18% respectively in control area. However in the SMC zone, the seroprevalence of both antibodies was 17%.

In SMC zone, the seroprevalence of anti-MSP1₄₂ antibody decreased by 2.5 folds between 2010 and 2011. For anti-AMA1 antibodies the seroprevalence rate decreased by 0.5 folds (Figure 2).

	Mean (AU)	CI (95%)	P value
IgG Anti-MSP1			
2010	23.2	20.2-26.2	
2011	11.8	11.1-12.4	
Mean difference	11.4	8.3-14.5	<10 ⁻³
IgG anti-AMA1			
2010	17.2	14.6-19.6	
2011	9.9	8.9-10.8	
Mean difference	7.2	4.5-9.9	<10 ⁻³

Table 2: Level of IgG anti-MSP1₄₂ and anti-AMA1 in 2010 and 2011.

2010			
Mean (AU)	SMC area (N=450)	Control area (N=416)	p value
IgG anti-MSP1 (95% CI)	25.2 (21.8 – 28.5)	17.9 (13.8 – 22.1)	<10 ⁻³
IgG anti-AMA1 (95% CI)	22.1 (18.3 – 25.9)	10.6 (8.5 – 12.6)	<10 ⁻³
2011			
	SMC area (N=464)	Control area (N=275)	p value
IgG anti-MSP1 (95% CI)	10.6 (9.7 – 11.5)	12.7 (11.5 – 13.4)	<10 ⁻³
IgG anti-AMA1 (95% CI)	8.7 (7.7 – 9.7)	11.9 (9.9 – 13.8)	<10 ⁻³

Table 3: Level of IgG anti-MSP1₄₂ and anti-AMA1 in 2010 and 2011 depending on the area (SMC + / SMC-).

Control area			
	Mean	95% CI	P value
IgG anti-MSP1			
2010	17.9	9.78–11.5	
2011	10.6	13.8–22.05	
Mean difference	7.2	2.1–12.3	<10 ⁻³
IgG anti-AMA1			
2010	10.6	8.5–12.6	
2011	11.9	9.9–13.8	
Mean difference	-1.3	-4.2–1.6	0,38
SMC area			
	Mean	95% CI	P value
IgG anti-MSP1			
2010	22.1	18.3–25.9	
2011	8.7	7.7–9.7	
Mean difference	13.74	9.5–13.3	<10 ⁻³
IgG anti-AMA1			
2010	25.2	21.8–28.5	
2011	12.5	11.5–13.4	
Mean difference	12.7	9.3–16.1	<10 ⁻³

Table 4: Level of IgG anti-MSP1₄₂ and anti-AMA1 in SMC area and control area depending to the year.

Multivariate analysis showed that seroprevalence of anti-AMA1 and anti-MSP1₄₂ antibodies increases with age and *Plasmodium falciparum* carriage. Seroprevalence of anti-MSP1₄₂ and anti-AMA1 antibodies was higher in children over 5 years old with respectively 39.7% (aOR=1.07; 95% CI (0.75 - 1.54), p value=0.68) and 39% (aOR=1.17; 95% CI (0.81 - 1.68), p value=0.38). The seroprevalence of

anti-MSP1 and anti-AMA1 antibodies was more important in children with malaria infection compared to subject without malaria infection. In children with malaria infection, seroprevalence of anti-MSP1₄₂ and anti-AMA1 was respectively 40.94% (aOR=1.13; 95% CI (0.77 - 1.63); p value=0.22) and 37.01% (aOR=1.16; 95% CI (0.81-1.69); p value=0.44). However, a decrease of anti-AMA1 and anti-MSP1 antibodies was observed according to the area and the study period. The seroprevalence of antibody was lower in SMC area compared to control area. Prevalence of anti-MSP1 and anti-AMA1 was respectively 30.63% (aOR=0.53; 95% CI (0.43 - 0.66), p value<10⁻³) and 27.68% (aOR =0.56; 95% CI (0.45 -0.71), p value<10⁻³) in SMC area. These result shown a protective effect of seasonal malaria chemoprevention to 47% for MSP1₄₂ and 44% for AMA1. In control zone, the seroprevalence of both antibodies was 47.47% and 41.82%.

The seroprevalence of both antibodies was higher in 2010 compared to 2011. In 2011, the prevalence of anti-MSP1₄₂ antibodies was 20.03% (aOR=0.23; 95% CI (0.18 - 0.28), p value<10⁻³). For anti-AMA1 antibody, it was 19.08% (OR=0.29; 95% CI (0.24 - 0.37), p value<10⁻³). These results showed a protective effect of IPT around 80% (Tables 5 and 6). No significant correlation was observed between seroprevalence of anti-MSP1₄₂ and anti-AMA1 antibodies and sex, anemia and nutritional status.

Discussion

In developing country malaria is still a leading cause of morbidity and mortality in children. This situation represents a major public health problem. To reduce malaria burden in children, WHO recommend in March 2012 Seasonal Malaria Chemoprevention (SMC) as a malaria preventive strategy in children [6,7]. This strategy has been shown to be effective, cost-effective, safe, and feasible for the prevention of in endemic areas. However, the influence of SMC on acquired immunity

	Number (%)	OR (95 % CI)	ORa (95 % CI)	p value
Age group				
< 1 year	66 (39.05%)	1	1	
(1-4 years)	273 (35.9%)	0.87 (0.62-1.23)	0.88 (0.63-1.25)	0.49
(5-10 years)	269 (39.7%)	1.03 (0.73-1.45)	1.07 (0.75-1.54)	0.68
Gender				
Female	298 (37.82%)	1	1	
Male	293 (38.35%)	1.02 (0.8-1.25)	1.01 (0.83-1.26)	0.81
Nutritional status				
Stunting	228 (43.02%)	1.4 (1.1-1.7)	0.9 (0.68-1.18)	
Underweight	162 (36.5%)	0.92 (0.73 (1.15)	0.91 (0.68-1.22)	0.55
Wasting	55 (26.19%)	0.54 (0.39-0.75)	0.83 (0.56-1.22)	0.37
Anemia				
No	130 (35.91%)	1	1	
Yes	478 (38.46%)	1.1 (0.87-1.42)	1.08 (0.82-1.43)	0.55
Malaria Pf				
No	556 (37.62%)	1	1	
Yes	52 (40.94%)	1.15 (0.79-1.66)	1.13 (0.77-1.63)	0.22
Area				
Control area	328 (47.47%)	1	1	
SMC area	280 (30.63%)	0.48 (0.39-0.6)	0.53 (0.43-0.66)	<10 ⁻³
Study period				
2010	460 (53.12%)	1	1	
2011	148 (20.03%)	0.22 (0.17-0.27)	0.23 (0.18-0.28)	<10 ⁻³

Table 5: Multivariate adjusted analysis for the risk factors of anti-MSP1₄₂ antibodies.

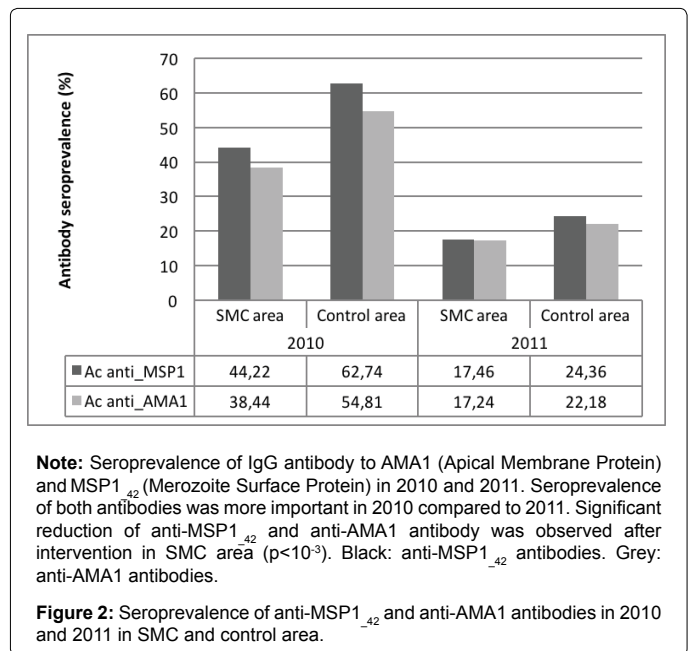
	Number (%)	OR (95 % CI)	ORa (95 % CI)	p value
Age group				
< 1 year	59 (34.91%)	1	1	
(1-4 years)	219 (28.85%)	0.75 (0.53-1.07)	0.75 (0.52-1.07)	0.11
(5-10 years)	264 (39%)	1.19 (1.84-1.69)	1.17 (0.81-1.68)	0.38
Gender				
Female	273 (34.64%)	1	1	
Male	258 (33.77%)	0.96 (0.77-1.18)	0.96 (0.78-1.19)	0.75
Nutritional status				
Stunting	209 (39.43%)	1.45 (1.16-1.8)	1.09 (0.83-1.44)	0.49
Underweight	143 (32.21%)	0.9 (0.72-1.14)	0.96 (0.7-1.31)	0.81
Wasting	47 (22.38%)	0.52 (0.37-0.74)	0.79 (0.53-1.2)	0.28
Anemia				
No	131 (36.19%)	1	1	
Yes	411 (33.07%)	0.87 (0.68-1.11)	0.97 (0.75-1.27)	0.87
Malaria Pf				
No	495 (33.49%)	1	1	
Yes	47 (37.01%)	1.16 (0.8-1.69)	1.16 (0.81-1.69)	0.44
Area				
Control area	289 (41.82%)	1	1	
SMC area	253 (27.68%)	0.53 (0.41-0.65)	0.56 (0.45-0.71)	<10 ⁻³
Study period				
2010	401 (46.3%)	1	1	
2011	141 (19.08%)	0.27 (0.22-0.34)	0.29 (0.24-0.37)	<10 ⁻³

Table 6: Multivariate adjusted analysis for the risk factors of anti-AMA1 antibodies.

is not well documented [8]. This aspect needs to be well documented because some molecules such as Sulfadoxine and Artesunate used during strategy may have immunosuppressive effects [9,13].

It was in this context, we conducted this study in the South-eastern part of Senegal (Velingara) in order to assess the immunological effects of Seasonal Malaria Chemoprevention (SMC) with Sulfadoxine-Pyrimethamine (SP) and Amodiaquine (AQ) among children under 10 years. The results of this study showed a decrease of *P. falciparum* malaria prevalence between 2010 and 2011. These results were reduction in malaria prevalence is confirmed by results from the national malaria surveys [17,18].

A decrease of anti-MSP1₄₂ and anti-AMA1 antibodies was observed between 2010 and 2011 and it was higher in children who received SMC with SP plus AQ compared to those in control area. Overall, the seroprevalence of malaria antibodies decreased between 2010 and 2011. One year after SMC implementation, the seroprevalence of malaria antibodies was more important in control y area compared to SMC area where seroprevalence of antibodies was lower. Similar results were found by other authors. The low prevalence of malaria antibodies in SMC was in line with results observed by Boulanger et al. when assessing the immunological consequences of intermittent preventive



treatment in Senegalese children compared to control group [8]. In Gambia a decrease of malaria antibodies production was observed in children who received IPT with Pyrimethamine and Dapsone [19].

Our findings were similar with what was observed in Ghanaian children six months after intermittent preventive treatment with single dose of SP [20]. However, previous studies have shown no significant difference between IPT group and control group in terms of antibody production [21-23].

Overall, a year after SMC implementation a decrease of protective antibodies against malaria was noted. This situation increases the susceptibility of children to malaria infection. Indeed, a year after SMC implementation *Plasmodium falciparum* malaria, prevalence was higher in areas where children had access to SMC compared to control area. Similar findings concerning the rebound effect of SMC were described previously. In Ghana an increase of malaria incidence by 62% in children who had access to intermittent preventive treatment a year after intervention was observed [19].

Multivariate analysis, showed that seroprevalence of anti-MSP1₄₂ and anti-AMA1 antibodies is correlated with age, *Pf* carriage and the study period and area. Antibody level increases with age and *Pf* carriage. Similar results were found previously [24-27]. Seroprevalence of anti-MSP1₄₂ and anti-AMA1 antibodies increase with age. These findings were in line with what was observed in the in northern part of Senegal with an increase of IgG according to the age [28]. Previous study conducted in Senegal showed that antibodies level increases with age, specifically in children over 5 years [29]. These results are in line with those found in Ghana that showed IgG levels increased with age [30].

Antibody prevalence increases with *Plasmodium falciparum* carriage. Seroprevalence of anti-MSP1₄₂ and anti-AMA1 antibodies was more important in children with *Plasmodium falciparum* infection compared to children without *Plasmodium falciparum* infection. Similar results were found in Ghana and Mozambique with a high level of antibodies in subjects with active malaria [21,31].

The seroprevalence of anti-MSP1₄₂ and anti-AMA1 antibodies was lower in 2011 than in 2010. The low prevalence of malaria antibodies may be due by the decrease of malaria prevalence and the effect of seasonal malaria chemoprevention.

Conclusion

SMC is an effective strategy for malaria prevention in children under 10 years. The strategy can as well induce a decrease of IgG anti-AMA1 and anti-MSP1₄₂ which are associated with protection against malaria. Consequently, this strategy needs to be renewed each year in areas where malaria is highly seasonal to avoid a resurgence of malaria, while promoting the use of other antimalarial interventions.

Competing Interests

The authors declare that they have no competing interests.

Authors' Contributions

KS, RCT, DS, MN, AS, MLT, ID, DN, OG, and BF conceived and designed the study. KS and RCT monitored the data collection. KS, MN, AS, and MLT collected data in the site. KS analyzed the data and wrote the first draft of the manuscript. All authors read and approved the final manuscript.

Acknowledgments

We thank all patients who agreed to participate in the study. We also acknowledge Dr Patrick Duffy and Dr Richard Shimp from NIH/NIAID (National Institutes of Health/National Institute of Allergy and Infectious Diseases) and Daniel Dodo, from Noguchi Memorial Institute for Medical Research, University of Ghana, Legon, Ghana) who provided the antigens.

References

1. WHO (2013) World malaria report. World Health Organisation, Geneva.
2. Cisse B, Cheikh S, Denis B (2006) Seasonal intermittent preventive treatment with artesunate and sulfadoxine pyrimethamine for prevention of malaria in Senegalese children: A randomised, placebo-controlled, double-blind trial. *Lancet* 367: 659-667.
3. Dicko A, Sagara I, Sissoko MS, Guindo O, Diallo AI, et al. (2008) Impact of intermittent preventive treatment with sulphadoxine-pyrimethamine targeting the transmission season on the incidence of clinical malaria in children in Mali. *Malar J* 7: 123.
4. Kweku M, Liu D, Adjuik M, Binka F, Seidu M, et al. (2008) Seasonal intermittent preventive treatment for the prevention of anaemia and malaria in Ghanaian children: A randomized, placebo controlled trial. *PLoS One* 3: 4000.
5. Schellenberg D, Menendez C, JAponte JJ, Kahigwa E, Tanner M, et al. (2005) Intermittent preventive antimalarial treatment for Tanzanian infants: follow-up to age 2 years of a randomised, placebo-controlled trial. *Lancet* 365: 1481-1483.
6. WHO (2012) Seasonal Malaria Chemoprevention (SMC) for Plasmodium falciparum malaria control in highly seasonal transmission areas of the Sahel sub regions in Africa.
7. Cairns M, Roca-Feltre A, Garske T, Wilson AL, Diallo D, et al. (2012) Estimating the potential public health impact of seasonal malaria chemoprevention in African children. *Nat Commun* 3: 881.
8. Boulanger D, Sarr JB, Fillo L F, Sokhna C, Cisse B, et al. (2010) Immunological consequences of intermittent preventive treatment against malaria in Senegalese preschool children. *Malar J* 9: 363.
9. Bygbjerg IC, Flachs H (1986) Effect of chloroquine on human lymphocyte proliferation. *Trans R Soc Trop Med Hyg* 80: 231-235.
10. Tine RCK, Faye B, Ndour CT, Ndiaye JL, Ndiaye M, et al. (2011) Impact of combining intermittent preventive treatment with home management of malaria in children less than 10 years in a rural area of Senegal: a cluster randomized trial. *Malar J* 10: 358.
11. Tine RC, Faye B, Ndour CT, Sylla K, Sow D, et al. (2013) Parasitic infections among children under five years in Senegal: Prevalence and effect on anaemia and nutritional status. *ISRN Parasitology* 272701: 1-6.
12. Corran PH, Cook J, Lynch C, Leendertse H, Manjurano A, et al. (2008) Dried blood spots as a source of anti-malarial antibodies for epidemiological studies. *Malar J* 7: 195.
13. Drakeley CJ, Corran PH, Coleman PG, Tongren JE, McDonald SL, et al. (2005) Estimating medium- and long-term trends in malaria transmission by using serological markers of malaria exposure. *Proc Natl Acad Sci USA* 102: 5108-5113.
14. Kocken CH, Withers-Martinez C, Dubbeld MA, Van Der WA, Hackett F, et al. (2002) High-level expression of the malaria blood-stage vaccine candidate *Plasmodium falciparum* apical membrane antigen 1 and induction of antibodies that inhibit erythrocyte invasion. *Infect Immun* 70: 4471-4476.
15. Shimp RL, Martin LB, Zhang Y, Henderson BS, Duggan P, et al. (2006) Production and characterization of clinical grade *Escherichia coli*-derived *Plasmodium falciparum* 42-kDa merozoite surface protein 1 (MSP142) in the absence of an affinity tag. *Protein Expr Purif* 50: 58-67.
16. Guitard J, Cottrell G, Magnouha NM, Salanti A, Li T, et al. (2008) Differential evolution of anti-*VAR2CSA*- IgG3 in primigravidae and multigravidae pregnant women infected by *Plasmodium falciparum*. *Malar J* 7: 10.
17. National Agency for Statistics and Demography (2009) National malaria survey 2008-2009, Senegal.
18. National Agency for Statistics and Demography (2013) Demographic and health survey continues in Senegal 2012-2013.
19. Otoo LN, Snow RW, Menon A, Byass P, Greenwood B (1988) Immunity to malaria in young Gambian children after a two-year period of chemoprophylaxis. *Trans R Soc Trop Med Hyg* 82: 59-65.
20. Schreiber N, Kobbe R, Adjei S, Adjei O, Klinkert MQ, et al. (2007) Immune responses after single-dose sulphadoxine-pyrimethamine indicate underestimation of protective efficacy of intermittent preventive treatment in infants. *Trop Med Int Health* 12: 1157-1163
21. Quelhas D, Puyol L, Quinto L, Serra-Casas E, Nhampossa T, et al. (2008) Impact of intermittent preventive treatment with sulfadoxine-pyrimethamine on antibody responses to erythrocytic-stage *Plasmodium falciparum* antigens in infants in Mozambique. *Clin Vaccine Immunol* 15: 1282-1291.
22. Quelhas D, Jiménez A, Quinto L, Serra-Casas E, Mayor A, et al. (2010) IgG against *Plasmodium falciparum* variant surface antigens and growth inhibitory antibodies in Mozambican children receiving intermittent preventive treatment with sulfadoxine-pyrimethamine. *Immunobiology* 7: 793-802.
23. Nhabomba AJ, Guinovart C, Jiménez A, Manaca MN, Quinto L, et al. (2014) Impact of age of first exposure to *Plasmodium falciparum* on antibody responses to malaria in children: A randomized, controlled trial in Mozambique. *Malar J* 13: 121.
24. Cook J, Reid H, Iavro J, Kuwahata M, Taleo G, et al. (2010) Using serological measures to monitor changes in malaria transmission in Vanuatu. *Malar J* 9: 169
25. Aguirre AR, Cuentos A, Speybroeck N, Cook J, Mancilla J, et al. (2013) Assessing malaria transmission in a low endemicity area of north-western Peru. *Malar J* 12: 339.
26. Doodoo D, Aikins A, Kusi KA, Lamptey H, Remarque E, et al. (2008) Cohort study of the association of antibody levels to AMA1, MSP119, MSP3 and GLURP with protection from clinical malaria in Ghanaian children. *Malar J* 7: 142.
27. Oduro RA, Conway DJ, Schellenberg D, Satoguina J, Greenwood BM, et al. (2013) Sero-epidemiological and parasitological evaluation of the heterogeneity of malaria infection in the Gambia. *Malar J* 12: 222.
28. Sarr JB, Remoue F, Samb B, Dia I, Guindo S, et al. (2007) Evaluation of antibody response to *Plasmodium falciparum* in children according to exposure of *Anopheles gambiae* or *Anopheles funestus* vectors. *Malar J* 6: 117.
29. Perraut R, Richard V, Varela ML, Trape JF, Guillotte M, et al. (2014) Comparative analysis of IgG responses to *Plasmodium falciparum* MSP1p19 and PF13-DBL1 α 1 using ELISA and a magnetic bead-based duplex assay (MAGPIX@Luminex) in a Senegalese meso-endemic community. *Malar J* 13: 410.
30. Doodoo D, Atuguba F, Bosomprah S, Ansah N, Ansah O, et al. (2011) Antibody levels to multiple malaria vaccine candidate antigens in relation to clinical malaria episodes in children in the Kasena-Nankana district of Northern Ghana. *Malar J* 10: 108.
31. Quelhas D, Puyol L, Quinto L, Serra-Casas E, Nhampossa T, et al. (2008) Impact of intermittent preventive treatment with sulfadoxine-pyrimethamine on antibody responses to erythrocytic-stage *Plasmodium falciparum* antigens in infants in Mozambique. *Clin Vaccine Immunol* 15: 1282-1291.