

## Effect of Imidacloprid on Reproduction of Female Albino Rats in Three Generation Study

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### Abstract

In the present study, the effect of oral administration of imidacloprid over three generations on biochemical, histological and physiological alterations in female rats was assessed. Female rats were divided into three groups. Group 1 was control and was given corn oil, group 2 was administered imidacloprid at the rate of 10 mg/kg bw/day, group 3 was administered imidacloprid at the rate of 20 mg/kg bw/day. F0 and F1 generation female albino rats were dissected for this study. Weight of ovary decreased significantly at higher dose of treated female rats of F0 and F1 generation. Histopathology of ovary of group 2 and group 3 revealed different stages of follicles. The level of acid phosphatase (ACP) and alkaline phosphatase (ALP) increased significantly at higher dose in the ovary of females of both the generations. In either generation, non-significant changes were observed in fertility index, live birth index, gestation lengths and sex ratio. Female F1 pups in the 20 mg/kg/day group showed a significant decreased body weight on postnatal day 21 as compared to F0 pups on day 21.

**Conclusion:** The lower dose of imidacloprid (10 mg/kg/day T1) had no effect on various reproductive parameters of female rats and higher dose (20 mg/kg/day T2) of imidacloprid had some significant effects on feed consumption and reproductive parameters for three generation reproductive study.

**Keywords:** Imidacloprid; Female albino rats; Fertility index; Sex ratio; Reproductive parameters; Lower dose; Higher dose; Three generations

### Introduction

Imidacloprid, a neonicotinoid the newest class of major insecticide has outstanding potency and systemic action for crop protection against piercing and sucking insect pests and also highly effective for control of flea on cats and dogs [1]. Few cases of acute human poisoning have been reported following ingestion of imidacloprid formulations [2,3]. There are some reports that show imidacloprid has an adverse effect on the reproductive tract [4], also this compound has been identified as having teratogenic [5], mutagenic [6] and carcinogenic [7] effects in animals and humans. Many pesticides having endocrine disruptor properties are also known to adversely impair the reproductive competence of males. Imidacloprid may adversely affect reproduction and cause developmental delays as a result of maternal toxicity. The effects of imidacloprid on reproduction and development were examined in a two generation, two-litter study in Wistar rats (30/sex/dose in the parental generation, P1). The dietary doses were 100, 250 and 700 ppm [8]. Maternal toxicity at 700 ppm included decreased body weight gain and food consumption with a marked reduction during lactation. In one study, pregnant rats fed technical grade imidacloprid throughout pregnancy and lactation at doses of 0, 100, 250 and 750 ppm revealed no effects other than significantly reduced food consumption (14% relative to controls) in the mother rats [9]. Imidacloprid was not found to affect reproductive variables or cause birth defects. However, reduced mean body weight and body weight gain relative to controls was observed in the males and females of all generations at the highest dietary concentration tested (700 ppm). At this concentration, parental animals also had reduced body weights, relative to controls, in association with reduced food consumption [10]. In spite of a number of studies on the effects of imidacloprid on reproductive behavior of animals, no information is available on the effect of imidacloprid on the two generation reproduction of rats when females were treated with imidacloprid. Aminotransferases and phosphatases are important and critical enzymes in the liver metabolic activity and are responsible

for detoxification processes. So any interference in various enzyme levels lead to biochemical impairment and lesions of the tissue. The liver is the principal target of imidacloprid toxicity, as demonstrated by its elevated serum transaminase, alkaline phosphatase and/or glutamate dehydrogenase activities; and alterations of other clinical parameters. Therefore, this study was designed to investigate the effects of imidacloprid on three generation reproduction of rats (when only females were treated with imidacloprid).

### Materials and Methods

#### Chemical

Commercial product of imidacloprid (Confidor, 17.8%, w/w imidacloprid as active ingredient) used in this study was purchased from the local market in Ludhiana, India.

#### Animals and experimental design

The study was conducted on sexually mature female albino rats, 3 months of age, weighing 100-150 g obtained from Guru Angad Dev Veterinary and Animal Sciences University (GADVASU), Ludhiana. The animals were housed in groups of two rats per cage. The rats were acclimatized for one week before using them for experimentation. The rats were maintained under controlled conditions of temperature ( $22 \pm 2^\circ\text{C}$ ) and humidity (30-70%) with 12 h light and dark cycle. After acclimatization for one week, healthy rats were subjected to this study,

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with 6 female rats constituting each group. Animals were divided into three groups. Group I served as control and given corn oil orally. Group II rats were given 10 mg/kg bw/day dose of imidacloprid. Group III rats were given 20 mg/kg bw/day dose of imidacloprid. The animals were given standard diet containing pelleted food and water *ad libitum*. The experimental protocol met the National guidelines on the proper care and use of animals in the laboratory research. The Institutional Animal Ethics Committee (IAEC) approved this experimental protocol. The study began with 6 female rats/group (F0 generation), and they were exposed to imidacloprid orally at 10 and 20 mg/kg bw/day. After 8-week administration of imidacloprid, each treated female was mated with a normal male having no dosage group and pregnant females were allowed to deliver and nurse their pups. F0 females were necropsied after weaning of their pups. Administration of imidacloprid was continued throughout the mating, gestation and lactation periods until necropsy. For the second generation, 6 female weanlings in each group of F0 generation were selected as F1 parents on post natal days 21-25 to equalize the mean body weights among groups as much as possible. The day on which F1 parental animals were selected was designated as day 0 of dosing for the F1 generation. F1 selected rats were given imidacloprid orally dissolved in corn oil mated after 8-week administration. They were allowed to deliver and nurse their F2 pups.

## Dosing

Imidacloprid was dissolved in corn oil to obtain the desired test concentrations and given orally to female rats for two generations. Two doses of imidacloprid 10 and 20 mg/kg bw/day were given to females of F0 and F1 generation, and control rats of each generation were given corn oil. F0 females mated with normal males to get F1 generation and F1 females mated with normal rats to get F2 generation. 10 mg/kg/day dose was reported as NOEL (No Observed Effect Level Dose) by Bhardwaj et al. and Kapoor et al. [11,12] and other higher than NOEL was taken in the present study. Administration to F0 parental female animals was started at 3 months of age. Administration to F0 females lasted until necropsy through 10 weeks or more of the pre-mating, mating, gestational, lactational periods and during weaning of the F1 offspring. Administration to F1 parental animals was started from age of 6 weeks old, until necropsy through 8 weeks or more of the pre-mating, mating, gestational, lactational periods and during weaning of the F2 offspring.

## Parental data (F0 and F1)

**Clinical observations:** Throughout the study, all F0 and F1 parental female rats were observed at least twice daily and cages were inspected daily for evidence of ill health or reaction to the treatment. F0 females underwent physical examinations beginning on the day treatment commenced, and weekly until mating. After mating, the physical examinations occurred on days 1, 7, 14, 21 and 28. The same schedule of examinations was used for the selected F1 rats. F0 females were weighed on the day treatment commenced, at weekly intervals until mating was detected, on days 0, 6, 13 and 20 after mating, on days 0, 4, 7, 14 and 21 of lactation and prior to necropsy. After selection, the F1 animals were weighed following the same schedule as the F0 animals.

**Body weight and food consumption:** All adult rats were observed daily for clinical signs of toxicity; food consumption and body weight were recorded weekly during mating, gestation and lactation period. For F0 females mean weekly food consumption was calculated for individual animals prior to mating. Mean daily food consumption was calculated for each F0 female based on the data recorded for the period days 0-5, 6-12 and 13-21 post-mating, and days 1-3, 4-6, 7-13 and 14-20

of lactation. Food consumption for the F1 animals was recorded at the same frequency following selection.

**Mating procedures:** At the age of 12 weeks after the 8 week administration period starting from six week old in the F0 animals, and at the age of 14-15 weeks after 8 week oral administration period from six weeks of age in the F1 animals, females were moved to the cages of male partners in the evening, and the males and females cohabited at a sex ratio of 1:1 through the mating period. Existence of a vaginal plug or sperm in the vaginal smear was examined every morning from the following day. When either was detected, it was judged that copulation had occurred and the day was defined as gestation day (GD) 0. The mating period was limited to two weeks. Pregnancy was confirmed by existence/absence of delivery and/or by investigating implantation sites at the time of necropsy. Beginning on day 20 after mating, females were inspected 3 times daily for evidence of parturition. The progress and completion of parturition was monitored, the numbers of live and dead offspring were recorded. Individual F1 offspring's were numbered; within each litter day 1-postpartum. The selected F1 generation was allocated to its specific treatment group when they were 6 weeks old.

Vaginal smears were obtained from the female animals everyday in the morning to examine the estrous cycle during four weeks before mating; starting from 12 weeks of age for the F0 parents and from 11 weeks old for the F1 parents, and the mean days of estrous cycle were calculated. Cases with estrous cycle other than 4 to 6 days were regarded as abnormal. F0 and F1 parental females were anaesthetized by chloroform. The following organs were weighed in all the F0 and F1 females: the liver, ovaries and uterus.

## Biochemical analysis

After sacrifice, the tissue sample of liver was homogenized in the phosphate buffer saline. The activities of aspartate aminotransferase (AST), alanine aminotransferase (ALT) were estimated by the method of Reitman and Frankel as described by Bergmeyer and acid phosphatase (ACP), alkaline phosphatase (ALP) was estimated by method of Bessey et al.

## Processing of tissues for histopathology

All F1 and F2 rats were weighed and sacrificed by chloroform anesthesia. For histopathological studies, ovaries of treated and control rats were fixed in 10% formalin. After routine processing and dehydration of each tissue, paraffin sections were cut at 5 um and stained with hematoxylin-eosin for microscopic examination. Serial sections of ovary were studied for various observations like total number of follicles, number of normal and atretic follicles, oocyte and nucleus as described by Kaur and Guraya.

## Offspring evaluation (F1 and F2)

All pups derived from F0 and F1 parents (F1 and F2 litters, respectively) were examined as soon as possible on the day of birth to determine the number and sex of pups, the number of live born and stillborn members of each litter and gross abnormalities. All individual offspring's were examined approximately 24 h after birth (day 1) and daily thereafter for any evidence of ill health. Litter size and mortality were recorded daily from days 1 to 21. The sex ratios of each litter were recorded. Individual body weights were recorded on days 0,4,7,14,21. The dams were removed and offspring weaned on day 21 of age, and the selection of the F1 generation was made on day 25. For the selected F1 females, sexual maturation was assessed daily from day 25 of age until vaginal opening occurred. Females that littered and reared offspring to

weaning were euthanized on day 28 post-partum (after weaning). For all F0 and the selected F1 adult females, detailed necropsies involving full macroscopic examinations weighing of the liver, ovary and uterus was carried out.

**Reproductive performance of F0 and F1 rats:** The reproductive parameters from this study were expressed in terms of indices, weights and ratios that considered all stages from conception to weaning [13]. These parameters were defined below:

Fertility index (%)=(Number of females delivering /number of females cohabited) × 100

Live birth index (%)=(Number of live pups at day 0/number of pups born) × 100

4-day survival index (%)=(Number of live pups on day 4/ number of pups alive on day 0) × 100

21-day (weaning) survival index (%)=(Number of pups alive on day 21/ number of pups alive on day 4) × 100

Litter size=Number of pups/number of pregnant females

Body weights of pups=The body weight of pups were recorded on days 0, 4, 7, 14 and 21

### Statistical analysis

All values were presented as the mean ± standard error of means (S.E.M). Comparisons were made between control and treated groups using “Analysis of Variance (ANOVA)” as a statgraphics statistical package. The body weight of parental animals, food consumption, length of estrous cycle, organ weight was evaluated by student t test for pairwise comparisons between control and individual treatment groups.

## Results

### Data for parental animals

**Clinical signs and body weight:** The most consistent finding was aggression/hyperactivity which was observed throughout the study in both F0 and F1 females of imidacloprid treated groups. Some females of the low and high dose groups had vaginal discharges. Mean estrous cycle days were significantly reduced in higher dose (T2) treated group of F0 females (4.06 ± 0.03, P<0.01) as compared to control (5 ± 0.21). In F1 females there was non-significant change in mean estrous cycle days. Body weight data of F0 and F1 females are shown in Table 1. The imidacloprid treated F0 and F1 females experienced reduction in body weight. The total weight gain of females reduced non significantly in T1 and T2 group (Table 2).

**Organ weights of F0 and F1 parental rats:** Organ weights of ovary and uterus for F0 and F1 parental animals are summarized in Table 3. The relative ovarian weight was significantly reduced in higher dose (T2)

group in F0 and F1 generation. Weight of uterus increased in both F0 and F1 females.

**Food consumption:** During the pre-pairing phase, compared to controls, food consumption was significantly decreased in imidacloprid treated females as compared to control of both the generations (Table 4). Decreased food consumption in F1 generation was significant as compared to F0 generation. During most of gestation and lactation, feed intakes were significantly increased in higher dose (20 mg/kg/d) and non-significant increase was also seen in the 10 mg/kg/day dose group females (Tables 5 and 6). Average gestation length was slightly increased by 1-2 days in the higher dose group (23 days); all gestation lengths were within the expected range of 22-23.5 days. There was non-significant increase in length of gestation days in treated groups as compared to control in both the generations.

### Biochemical analysis

There was significant increase in activity of liver ALT in F1 and F2 females. AST, ACP and AKP enzyme activity increased non-significantly in F1 and F2 females (Tables 7 and 8).

### Offspring evaluation (F1 and F2)

**Reproductive performance of F0 and F1:** The reproductive performance of F0 and F1 rats were evaluated by looking at various parameters that included fertility index, live birth index, weaning index, litter size, body weight of pups and sex ratio. In F0 rats, imidacloprid treatment caused a non-significant change on the gestation length, litter size and the viability indices at the higher dose group compared to control (Table 9). Fertility index decreased in imidacloprid treated groups as compared to control in F0 and F1 generation.

Similarly, in F1 generation rats, no imidacloprid treatment related effect on gestation, viability and sex ratio was observed in the higher dose group compared to control. The body weight of F0 pups increased significantly, and at day 21 body weight of F1 pups was significantly lowered in high dose group as compared to control (Table 10).

### Hormone levels

Progesterone and estrogen levels were estimated in plasma of control and treated rats of F1 and F2 generation (Table 5). In higher dose treated group there is non-significant decrease in progesterone level in F1 and F2 generation. Estrogen level showed non-significant increase in treated groups in both the generations (Table 11).

### Histology

Histologically, sections of ovary of control rats showed different stages of follicles and sections of higher dose of treated rats showed more number of atretic follicles as compared to control rats. All the stages of follicular development viz. primary, secondary, tertiary, early

Weeks	F0			F1		
	Control	T 1	T 2	Control	T 1	T 2
I	101.67 ± 1.05	103.3 ± 1.52	103.33 ± 1.66	103.33 ± 1.67	105 ± 1.82	102.5 ± 2.14
II	107.5 ± 1.12	109.16 ± 1.40	110 ± 3.16	110 ± 1.29	110 ± 1.82	110 ± 1.82
III	117.5 ± 2.14	115 ± 1.17	115 ± 1.17	120.83 ± 2.38	120 ± 1.29	120 ± 1.82
IV	129.17 ± 2.00	136.67 ± 4.95	129.17 ± 3.98	139.17 ± 1.53	130 ± 1.29	130.83 ± 2.00
V	144.16 ± 3.33	141.6 ± 3.33	136.6 ± 4.95	149.1 ± 5.54	140.83 ± 1.53	141.6 ± 6.54

Values are Mean ± SE of 6 animals in each group

**Table 1:** Effect of imidacloprid on weekly body weight of female albino rats before pairing of F0 and F1 generation as compared to control.

Traits	F0			F1		
	Control	T 1	T 2	Control	T 1	T 2
Mating	141.6 ± 3.06	152.5 ± 3.81	156.66 ± 3.07	158.33 ± 3.33	152.5 ± 6.02	152.5 ± 1.11
Gestation	152.5 ± 4.95	156.6 ± 4.21	173.33 ± 3.57*	169.16 ± 1.53	167.5 ± 2.50	166.6 ± 3.80
After delivery	146.6 ± 1.67	155 ± 3.65*	166.6 ± 2.47	158.33 ± 3.33	159.16 ± 4.36	145.8 ± 7.12
During lactation	165.8 ± 2.38	173.3 ± 8.33	186.6 ± 4.59**	185 ± 2.88	181.66 ± 3.07	176.6 ± 4.94

Values are Mean ± SE of 6 animals in each group

\*Significantly different from control at P<0.05

\*\*Significantly different at P<0.01

**Table 2:** Effect of imidacloprid on body weight of female albino rats after pairing of F0 and F1 generation as compared to control.

	Control	T 1	T 2
<b>F 0 females</b>			
No. of females examined	6	6	6
Final body weight (g)	129.16 ± 3.982	141.7 ± 3.333	156.7 ± 3.073
Ovary (g)	0.030 ± 0.002	0.029 ± 0.001	0.020 ± 0.001**
Uterus (g)	0.135 ± 0.021	0.172 ± 0.036	0.182 ± 0.031
<b>F 1 females</b>			
No. of females examined	6	6	6
Final body weight (g)	131.7 ± 4.013	140.8 ± 3.515	153.3 ± 7.490**
Ovary (g)	0.037 ± 0.001	0.036 ± 0.004	0.031 ± 0.001*
Uterus (g)	0.137 ± 0.020	0.206 ± 0.043	0.191 ± 0.015

Values represent the mean ± SE of 6 animals in each group

\*Significantly different from control at P<0.05

\*\*Significantly different from control at P<0.01

**Table 3:** Relative organ weights for F0 and F1 females.

antral and antral were observed in control and imidacloprid treated rats in F1 and F2 generation (Figures 1 and 2).

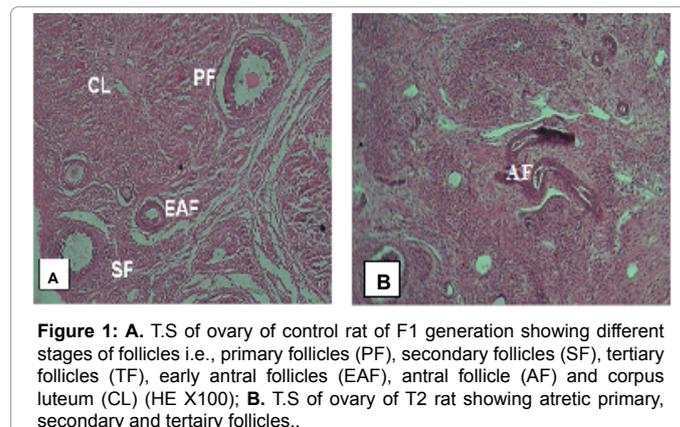
## Discussion

The present three generation reproductive study was performed to provide general information concerning the effects of imidacloprid on the performance of female reproductive system, and on the growth and development of the offspring. Mean estrous cycle days were significantly reduced in T2 treated group of F0 females (4.06 ± 0.03, P<0.01) as compared to control (5 ± 0.21). Studies of Borgeest et al. [14] experienced a significant increase in the percentage of days in estrous phase compared with control and methoxychlor treated mice. It is obvious that monitoring of body weight provides information on health level of animals which can also be important interpretation of reproductive effects [15]. Body weight of females increased significantly during gestation and lactation period of F0 and F1 generation.

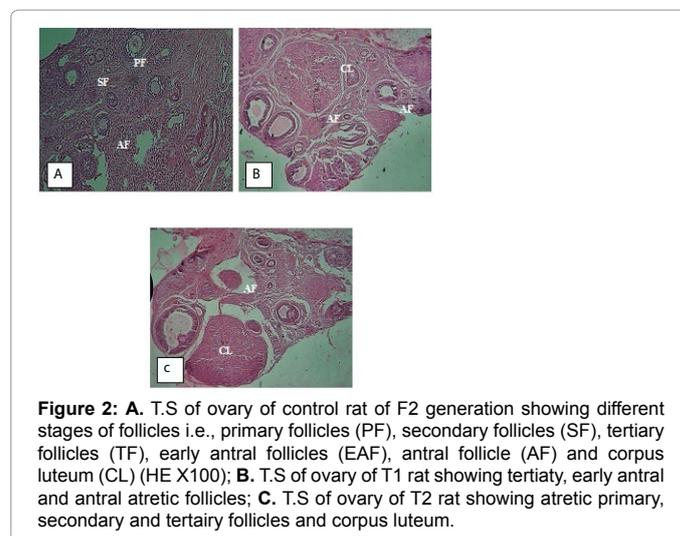
The relative ovarian weight was significantly reduced in higher dose (T2) group in F0 and F1 generation. Reduction in body weight at 20 mg/kg/day dose level as observed in the present study may be correlated with decreased sex organ weight (ovary weight) in both the generations, which reflects the effect of imidacloprid on the reproductive system. Organophosphates like methyl parathion, dimethioate and monocrotophos given to female albino rats have also resulted in significant decrease in the ovarian weights [16].

Food consumption of females before pairing for the F0 and F1 generation decreased significantly in treated females of both the generations. The decrease in feed consumption is correlated with decrease in body weight gain in all the treated groups [17] in the two generations. There was significant increase in food consumption of females during

gestation and lactation in F0 and F1 generation. Significant increase in food consumption was observed on 13-21 days of gestation in both the generations. During the period of gestation and lactation metabolism increases by 82.5% on average and the female assimilates additionally 304 kcal (including 60.5 kcal for gestation and 243.5 kcal for lactation) [18]. Lactation makes considerably greater demands on the mother's body than pregnancy does in species where the young are helpless at birth and depend on mother's milk for a comparatively long time. The requirements for the production of milk are met partly from within, by the mobilization of the mother's body tissues, and partly from without, by an increased food intake [19]. Similar results were found by Margaret et al. [20] that at different intervals throughout the gestation and lactation periods, increased food consumption was observed in F0 generation females of the mid- and high-dose groups of stanol esters,



**Figure 1:** A. T.S of ovary of control rat of F1 generation showing different stages of follicles i.e., primary follicles (PF), secondary follicles (SF), tertiary follicles (TF), early antral follicles (EAF), antral follicle (AF) and corpus luteum (CL) (HE X100); B. T.S of ovary of T2 rat showing atretic primary, secondary and tertiary follicles..



**Figure 2:** A. T.S of ovary of control rat of F2 generation showing different stages of follicles i.e., primary follicles (PF), secondary follicles (SF), tertiary follicles (TF), early antral follicles (EAF), antral follicle (AF) and corpus luteum (CL) (HE X100); B. T.S of ovary of T1 rat showing tertiary, early antral and antral atretic follicles; C. T.S of ovary of T2 rat showing atretic primary, secondary and tertiary follicles and corpus luteum.

Week	Control	T 1	T 2
<b>F0 generation</b>			
1	18 ± 0.090	17 ± 0.271*	16.76 ± 0.391*
2	25 ± 0.000	22.95 ± 0.66	17.23 ± 0.030
3	30.5 ± 1.295	23.43 ± 0.632	19.14 ± 1.351
4	30.5 ± 2.108	29.29 ± 1.355	20.43 ± 0.271
<b>F 1 Generation</b>			
1	13.56 ± 2.306	12.89 ± 0.548 <sup>a</sup>	12.28 ± 0.456
2	26.24 ± 2.741	24.76 ± 1.029	23.71 ± 1.355
3	33.86 ± 1.078	32.67 ± 1.046 <sup>a</sup>	30.33 ± 0.475
4	34.42 ± 0.579	31.15 ± 0.811	30.65 ± 0.968

Values represent the mean ± SE of 6 animals in each group

\*Significantly different from control at P<0.05

<sup>a</sup>Significantly different from T1 of F0 at P<0.01

**Table 4:** Food consumption of females before pairing for the F0 and F1 generation (g/animal/week).

F0 generation	Control	T 1	T 2
<b>Gestation</b>			
0-5 days	26.80 ± 0.000	26.80 ± 0.000	30.93 ± 1.307*
6-12	20.62 ± 0.688	24.15 ± 0.200	26.87 ± 0.295
13-21	24.09 ± 0.482	27.10 ± 0.662	27.97 ± 0.191**
<b>Lactation</b>			
1-3 days	28.54 ± 0.940	31.36 ± 1.574	33.19 ± 2.79*
4-6	29.30 ± 0.953	31.90 ± 1.364	33.62 ± 2.330
7-13	28.90 ± 1.079	31.55 ± 1.547	33.06 ± 2.551
14-20	28.75 ± 0.857	31.33 ± 1.526	32.96 ± 2.380

Values represent the mean ± SE of 6 animals in each group

\*Significantly different from control at P<0.05

\*\* Significantly different from control at P<0.01

**Table 5:** Food consumption of females during gestation and lactation for the F0 generation (g/animal/day).

F1 generation	Control	T 1	T 2
<b>Gestation</b>			
0-5 days	43.13 ± 1.499	38.93 ± 1.850	39.00 ± 1.820
6-12	43.52 ± 0.740	39.48 ± 0.314*	43.57 ± 1.065
13-21	33.87 ± 1.114	43.74 ± 0.625**	44.63 ± 1.042**
<b>Lactation</b>			
1-3 days	42.22 ± 1.921	42.06 ± 1.471	43.44 ± 1.240
4-6	38.89 ± 1.890	43.78 ± 1.210	45.94 ± 1.381*
7-13	43.55 ± 1.164	45.79 ± 0.375	46.62 ± 0.792
14-20	43.57 ± 0.620	42.55 ± 1.221	44.31 ± 1.240

Values represent the mean± SE of 6 animals in each group

\*Significantly different from control at P<0.05

\*\*Significantly different from control at P<0.01

**Table 6:** Food consumption of females during gestation and lactation for the F1 generation (g/animal/day).

Parameters (µmole/g)	Dosage/mg/kg/d		
	0 (Control)	10	20
ALT	27.362 ± 3.34	57.85 ± 2.84***	66.27 ± 7.72***
AST	53.15 ± 8.90	59.88 ± 4.38	69.34 ± 4.13
AKP	51.85 ± 21.16	61.00 ± 0.93	65.33 ± 9.21
ACP	27.29 ± 4.06	33.12 ± 1.19	34.23 ± 2.78

Values represent the mean ± SE of 6 animals in each group

\*\*\*Significantly different from control at P<0.001

**Table 7:** Liver biochemical data of female rats of F1 generation orally administered imidacloprid.

while increased food consumption was noted in F1 generation females of the mid- and high-dose groups during gestation. Such increases in

Parameters (µmole/g)	Dosage/mg/kg/d		
	0 (Control)	10	20
ALT	26.80 ± 3.59	53.54 ± 5.71	63.73 ± 9.59**
AST	68.00 ± 9.89	61.08 ± 4.28	70.30 ± 3.35
AKP	50.70 ± 24.35	63.06 ± 2.50	69.00 ± 8.94
ACP	28.70 ± 3.96	35.23 ± 1.17	39.43 ± 2.10

Values represent the mean ± SE of 6 animals in each group

\*\*Significantly different from control at P<0.01

**Table 8:** Liver biochemical data of female rats of F2 generation orally administered imidacloprid.

F 0 parents/F1 offspring	Control	T 1	T 2
Number of pairs at the start	6	6	6
Number of pregnant	6	5	5
Litter size	6	5	6.6
Fertility index of F0 female	100	83.33	83.33
Live birth index (%)	100	100	96.96
4-day survival index (%)	100	100	93.75
21-day survival index (%)	100	100	100
F 1 parents/F2 offspring	Control	T 1	T 2
Number of pairs at the start	6	6	6
Number of pregnant	6	5	5
Litter size	7	7.2	6.1
Fertility index of F0 female	100	83.33	83.33
Live birth index (%)	100	83.33	81.08
4-day survival index (%)	100	93.33	90
21-day survival index (%)	100	92.85	96.29

**Table 9:** Developmental findings for F1 and F2 pups.

food consumption are expected as a result of the animals' attempt to compensate for the reduced caloric value of the test diet compared to controls. Cripps and Williams [21] measured feed consumption during lactation in Sprague-Dawley rats and found an approximately 3- to 5-fold increase in daily feed consumption between post natal day 1 and post natal day 21. Similar increased food consumption during lactation in rats was reported by Shirley [22] and Arnold et al. [23].

There was non-significant increase in length of gestation days in treated groups as compared to control in both the generations. A slight increase in gestation length was observed in two generation study by Tyl et al. [24]. In F0 rats, no imidacloprid treatment related effect on the litter size and the viability indices (day 0 and 4) was observed at the higher dose group compared to control. Fertility index was slightly decreased in treated groups of both the generations. Similarly, in F1 generation rats, imidacloprid treatment caused non-significant change on fertility, survival index and litter size in the higher dose group compared to control. Suter et al. [8] also reported a mild effect or no effect on reproductive performance after exposure to imidacloprid. There were no effects on mating indices, fertility, gestation, litter size, mortality and no evidence of pathology at any dose level [25].

There was significant increase in ALT and AST activity in the liver. Bhardwaj et al. [11] also observed that oral administration of imidacloprid in female rats at the rate of 5, 10 and 20 mg/kg bw/day

Live pup weight (g)	Control	T 1	T 2
F 0 parents/F1 pups			
Day 0	3.90 ± 0.59	4.13 ± 0.24	6.08 ± 1.32**
Day 4	9.06 ± 0.59	9.05 ± 1.00	10.5 ± 2.27
Day 7	16.38 ± 0.50	16.01 ± 0.95	18.94 ± 4.01
Day 14	23.9 ± 1.19	22.83 ± 1.35	25.29 ± 5.38
Day 21	31.95 ± 6.78	30.44 ± 1.48	29.24 ± 1.48
F1 parents/F2 pups			
Day 0	2.73 ± 0.22	2.64 ± 0.19	2.81 ± 3.05
Day 4	6.38 ± 0.42	6.31 ± 0.10	6.62 ± 0.43
Day 7	13.09 ± 0.73	12.1 ± 0.93	12.67 ± 0.94
Day 14	17.29 ± 0.74	18.30 ± 0.86	17.60 ± 1.41
Day 21	29.93 ± 0.86	23.08 ± 2.46	20.74 ± 2.20**

Values represent the mean ± SE of 6 animals in each group

\*\* Significantly different from control at P<0.01

**Table 10:** Effect of imidacloprid on the body weight gain of F1 and F2 pups.

Hormones	F1		
	Control	T 1	T 2
Progesterone (ng/ml)	19.8 ± 7.29	19.4 ± 7.46	7.3 ± 1.82
Estrogen (pg/ml)	253.3 ± 47.23	256.6 ± 34.02	256.6 ± 28.007
F2			
Progesterone (ng/ml)	10.6 ± 6.90	8.6 ± 2.71	8.5 ± 2.44
Estrogen (pg/ml)	201.6 ± 11.66	218.3 ± 19.73	235.0 ± 16.48

**Table 11:** Effect of imidacloprid on hormones of F1 and F2 generation.

for 90 days resulted in elevation of serum ALT, AST, glucose, Blood Urea Nitrogen (BUN). It has been suggested that an increase in alkaline phosphatase (ALP) level occurs due to the damage of the cells of liver, kidney, small intestine, and bone resulting in the liberation of this enzyme in the blood systems [26].

The body weight of F0 pups increased significantly on day 0, and at day 21 body weight of F1 pups was significantly lowered in higher dose group as compared to control. The body weight of F1 pups at day 21 in high dose group was significantly lowered compared to control. Svetlana [25] reported a decrease in body weight gain of offspring's up to 13% compared to control until weaning at postnatal day 21. Similar results were found by Leslie et al. that body weight gain of offspring was significantly decreased from days 21-25 for Han Wistar rat females receiving 7500 ppm and 25,000 ppm rebaudioside A. Pup body weight and weight gains were reduced throughout lactation, with statistically identified lower weights on post natal day 21 in all generations at 100 mg/kg/day [27].

Oral administration of cypermethrin to female rats has resulted in significant decrease in plasma progesterone levels [28]. Earlier studies of Mani et al. also observed significant reduction in testicular enzyme 17 β-hydroxysteroid dehydrogenase, responsible for testosterone biosynthesis in male rats exposed to fenvalerate which may ultimately be leading to net decrease in testosterone concentration in group of rats [29].

Two doses of imidacloprid to female rats over two successive generations resulted in some effects on weight gain, food consumption, fertility index and showed no effects on gestation index, weaning index, sex ratio, live birth index of F1 and F2 pups [30,31]. Higher dose of imidacloprid (20 mg/kg bw/day) in both the generations showed a reduction in their body weight and food consumption and some effects on reproduction. Lower dose of imidacloprid (10 mg/kg bw/day) showed non-significant effects. Thus 10 mg/kg/day dose of imidacloprid which has been reported as NOEL (No Observed Effect Level) dose

has no adverse effects on either generation. Studies on imidacloprid have indicated 10 mg/kg/day as No Observed Effect Level (NOEL) as evidenced by various biochemical, hematological, neurobehavioral and oxidative stress parameters and produced significant changes at high dose levels (20 mg/kg/day) [11,12]. There have been no studies reported in the literature concerning the effects of imidacloprid on multigenerational reproduction after oral exposure to imidacloprid in female rats [32]. In our multigenerational experiment, the results indicated only minimal effects upon reproductive performance of rats. Imidacloprid exposure caused reduction in fertility index of both the generations. Suter et al. [8] also reported a mild effect or no effect on reproductive performance after exposure to imidacloprid.

### Conflict of Interest Statement

The authors declare that there are no conflicts of interest.

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### References

1. Yamamoto I, Tomizawa M, Saito T, Miyamoto T, Walcott EC, et al. (1998) Structural factors contributing to insecticidal and selective actions of neonicotinoids. Arch Insect Biochem Physiol 37: 24-32.
2. Wu IW, Lin JL, Cheng ET (2001) Acute poisoning with the neonicotinoid insecticide imidacloprid in N-methyl pyrrolidone. Clin Toxicol 39: 617-621.
3. Proença P, Teixeira H, Castanheira F, Pinheiro J, Monsanto PV, et al. (2005) Two fatal intoxication cases with imidacloprid: LC/MS analysis. Forensic Sci Int 153: 75-80.
4. Rouchard J, Gustinm F, Wauters A (1994) Soil organic matter aging and its effect on insecticide imidacloprid soil biodegradation in sugar beet crop. Toxicol Environ Chem 45: 149-155.
5. Pike KS, Reed GL, Graf GT, Allison D (1993) Compatibility of imidacloprid with fungicides as a seed-treatment control of russian wheat aphid (Homoptera: Aphidae) and effect on germination, growth, and Yield of Wheat Barley. J Econ Entomol 86: 586-593.
6. Placke FJ, Weber E (1993) Method of determining imidacloprid residues in plant materials. Pflanzenschutz-Nachrichten Bayer 46: 109-182.
7. Scholz K, Spittler M (1992) Influence of groundcover on the degradation of 14C-imidacloprid in soil. Proceeding of the Brighton Crop Protection Conference, pp: 883-888.
8. Suter P, Biedermann K, Luetkemeier H, Wilson J, Terrier C (1990) NTN 33893 Technical. Multiple Generation Study in Rats. RCC, Research and Consulting Company AG. Itingen, Switzerland. Study No. 100647. DPR Vol. 51950-0019 # 119496.
9. Sheets L (2001) A Developmental Neurotoxicity Screening Study with Technical Grade Imidacloprid in Wistar Rats: Lab Project Number: 99-D72-DV: 110245.
10. Solecki R (2001) Toxicological evaluations, pesticide residues in food. Pest Bio div Federal institute for health protection of consumers and veterinary medicine, Berlin, Germany.
11. Bhardwaj S, Srivastava MK, Kapoor U, Srivastava LP (2010) A 90 days oral toxicity of imidacloprid in female rats: morphological, biochemical and histopathological evaluations. Food Chem Toxicol 48: 1185-1190.
12. Kapoor U, Srivastava MK, Srivastava LP (2011) Toxicological impact of technical imidacloprid on ovarian morphology, hormones and antioxidant enzymes in female rats. Food Chem Toxicol 49: 3086-3089.
13. Oser BL, Oser M (1956) Nutritional studies on rats of diets containing high levels of partial ester emulsifiers. II. Reproduction and lactation. J Nutr 60: 489-505.
14. Borgeest C, Symonds D, Mayer LP, Hoyer PB, Flaws JA (2002) Methoxychlor may cause ovarian follicular atresia and proliferation of the ovarian epithelium in the mouse. Toxicol Sci 68: 473-478.

15. Aly HA, Domènech O, Abdel-Naim AB (2009) Aroclor 1254 impairs spermatogenesis and induces oxidative stress in rat testicular mitochondria. *Food Chem Toxicol* 163: 1733-1738.
16. Kaur S, Dhanju CK (2005) Biochemical effects of some organophosphorus pesticides on the ovaries of albino rats. *Indian J Physiol Pharmacol* 49: 148-152.
17. Bhadaniya AR, Joshi DV, Patel BJ, Kalaria VA, Padodara RJ, et al. (2012) Toxicopathological studies on experimentally induced acephate toxicity in wistar rats (*Rattus norvegicus*). *Wayamba J Animal Sci* 4: 451 -458.
18. Migula P (1969) Bioenergetics of pregnancy and lactation in European Common Vole. *Acta theriologica* 14: 169-179.
19. Widdowson EM (1976) Changes in the body and its organs during lactation: nutritional implications. *Ciba Found Symp* 45: 103-118.
20. Whittaker MH, Frankos VH, Wolterbeek AP, Waalkens-Berendsen DH (1999) Two-generation reproductive toxicity study of plant stanol esters in rats. *Regul Toxicol Pharmacol* 29: 196-204.
21. Cripps AW, Williams VJ (1975) The effect of pregnancy and lactation on food intake, gastrointestinal anatomy and the absorptive capacity of the small intestine in the albino rat. *Br J Nutr* 33: 17-32.
22. Shirley B (1984) The food intake of rats during pregnancy and lactation. *Lab Anim Sci* 34: 169-172.
23. Arnold DL, Bryce FR, Clegg DJ, Cherry W, Tanner JR, et al. (2000) Dosing via gavage or diet for reproduction studies: a pilot study using two fat-soluble compounds-hexachlorobenzene and aroclor 1254. *Food Chem Toxicol* 38: 697-706.
24. Tyl RW, Myers CB, Marr MC, Sloan CS, Castillo NP, et al. (2008) Two-generation reproductive toxicity study of dietary bisphenol A in CD-1 (Swiss) mice. *Toxicol Sci* 104: 362-384.
25. Svetlana EK, Numay RR, Thomas BM (2006) Imidacloprid. Risk Characterization Document. Dietary and Drinking Water Exposure. California EPA (Environmental Protection Agency).
26. Zimmerman HJ (1969) Serum enzymes determination as an aid to diagnosis. In: *Clinical diagnosis by Laboratory methods*. Dawidson I, Henry JB (eds). WB Saunders Company, Philadelphia, USA, p: 719.
27. Hanley TR Jr, Breslin WJ, Quast JF, Carney EW (2002) Evaluation of spinosad in a two-generation dietary reproduction study using Sprague-Dawley rats. *Toxicol Sci* 67: 144-152.
28. Saleem FM, Elmonsoury EGA, Yousef AE (1996) Reproductive performance in rats and ewes treated with pyrethroid (ectomin). *Assiut Vet Med J* 34: 103-123
29. Chao S, Casida JE (1997) Interaction of imidacloprid metabolites and analogs with the nicotinic acetylcholine receptor of mouse brain in relation to toxicity. *Pestic Biochem Physiol* 58: 77-88.
30. Curry LL, Roberts A, Brown N (2008) Rebaudioside A: two-generation reproductive toxicity study in rats. *Food Chem Toxicol* 46 Suppl 7: S21-30.
31. Baligar PN, Kaliwal BB (2002) Reproductive toxicity of carbofuran to the female mice: effects on estrous cycle and follicles. *Ind Health* 40: 345-352.
32. Wilen R, Bastomsky CH, Naftolin F (1981) Control of puberty in female rats: the effect of PTU-induced hypothyroidism and systematic undernutrition. *Pediatr Res* 5: 169-171.