

Comparison of HPLC and UV Spectrophotometric Methods for the Determination of Cefaclor Monohydrate in Pharmaceutical Dosages

Alfeen Mohammad A*

Analytical Chemistry Department, Faculty of Science, Al-Baath University, Homs, Syria

Abstract

This paper describes the development and evaluation of a HPLC and UV spectrophotometric methods to quantify Cefaclor Monohydrate in Oral suspensions and Capsules. HPLC analysis were carried out using a C18 Knauer column and a mobile phase composed of Triethylamine:methanol:Acetonitrile:water (2:10:20:68) v/v%, with a flow rate of 1.0 mL/min and UV detection at 265 nm. For the spectrophotometric analysis, water was used as solvent and the wavelength of 264 nm was selected for the detection. Both methods were found to quantify Cefaclor monohydrate in Oral suspensions and Capsules accurately. Therefore HPLC and UV methods presented the most reliable results for the analyses of Oral suspension and Capsules.

Keywords: HPLC; Cefaclor monohydrate; Spectrophotometric methods; Antibacterial activity

Introduction

Cefaclor monohydrate (CAS 56238-63-2) (Figure 1) is a second generation cephalosporin with high antibacterial activity; it has enhanced *in vitro* activity against clinically important Gram-positive and Gram-negative microorganisms [1]. The chemistry of cephalosporins has been widely explored because of their extensive medical applications [2]. Several analytical procedures are available in literature for the analysis of antimicrobial. These methods are spectrophotometry [3-13], high performance liquid chromatography [14-19], capillary electrophoresis [20], fluorimetry [21-24], polarography [25-29], titrimetry [30], and bioassay [31,32]. Spectrophotometric assay for determination of other cephalosporins as ceftazidime has been described [33] but no method for Cefaclor monohydrate had been previously described.

The purpose of this study was to develop and validate analytical methods to quantify Cefaclor monohydrate in Capsules and Oral suspensions, using HPLC and UV spectrometry. The results obtained by these methods were statistically compared, by using analysis of variance (ANOVA). In addition, the reliability and feasibility of them were evaluated focusing on routine quality control analysis.

Experimental

Reagents and materials

Cefaclor monohydrate reference standard was kindly donated by Parabolic Indian Ltd. The Capsules and Oral Suspensions were purchased from Medico Labs-Homs-Syria and Oubari Company-Aleppo-Syria. Ultra Pure Water was purified by using a Millipore system (Bedford, MA). Methanol, Acetonitrile, and Triethylamine (HPLC grade) was obtained from Merck (Fairfield, OH).

Instruments and analytical conditions

All HPLC measurements were made on a Waters 1525 Binary HPLC Pump, consisting of a 7725i manual injector with a 20 μ L loop (Rheodyne, Torrance, CA), integrated UV detector UV-vis (Milford, MA). The system employed a 250 mm \times 4.6 mm C18 column Wat 054275 (Milford, MA) and particle size of 5 μ m guard column. The detector was utilized at 265 nm and UV spectra from 200 to 400 nm were recorded on line for peak identification. The mobile phase consisted of Triethylamine:methanol:Acetonitrile:Ultra Pure water (2:10:20:68)

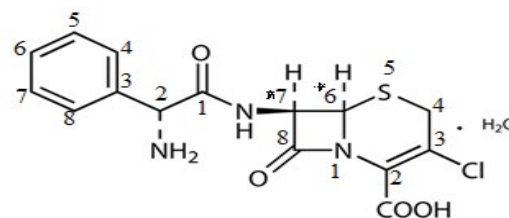


Figure 1: Structure of Cefaclor monohydrate.

v/v%, at a flow rate of 1.0 mL/min. The injection volume was 20 μ L. Ultraviolet spectrophotometric analyses were carried out on a UV-Vis Shimadzu UV mini 1240 (Shimadzu, Kyoto, Japan) spectrophotometer, in a 1 cm quartz cubette. The wavelength of 264 nm was selected for the quantitation of Cefaclor monohydrate and the measurements were obtained against water as a blank.

Preparation of standard and sample solutions

The standard stock solutions were prepared by dissolving 10 mg of Cefaclor monohydrate reference standard in 10 mL of water to get a concentration of 1 mg/mL. An aliquot of 100 μ L of the obtained solution was transferred to a 10 mL volumetric flask. The volume was adjusted with Ultra Pure water for spectrophotometric and chromatographic analysis, resulting in solutions of 10 μ g/mL.

The sample solutions were prepared by dissolving 10 mg of Cefaclor monohydrate powder for Capsules or Oral suspensions in 10 mL of water to get a concentration of 1 mg/mL. An aliquot of 100 μ L of this

*Corresponding author: Mohammad Anas Alfeen, Lecturer and Chemist, Department of Analytical Chemistry, Faculty of Science, Al-Hamra Street, Homs, Syrian Arab Republic, Tel: 963968628582; E-mail: chem_anas@yahoo.com

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solution was transferred to a 10 mL volumetric flask. The volume was adjusted with water for spectrophotometric analysis or mobile phase for chromatographic analysis, to obtain a solution at 10 µg/mL of Cefaclor.

Validation

The optimized spectrophotometric and chromatographic methods were completely validated according to the procedures described in ICH guidelines Q2 (R1) for the validation of analytical methods [34].

Linearity

Standard solutions containing 1000 µg/mL of Cefaclor monohydrate in water were prepared, in triplicate. Aliquots of these solutions were diluted in water. Eight different concentrations, corresponding to 1.0, 5.0, 10.0, 20.0, 30.0, 40.0, 50.0 and 60 µg/mL of Cefaclor (for UV analysis) and Twelve different concentrations, corresponding to 0.1, 0.5, 1.0, 5.0, 10.0, 20.0, 30.0, 40.0, 50.0, 60.0, 70.0 and 80.0 µg/mL of Cefaclor (for HPLC analysis). Calibration curves with concentration versus peak area or absorbance were plotted for each method and the obtained data were subjected to regression analysis using the least squares method.

Precision

The intra-day precision was evaluated by analyzing six samples (n=6), at the test concentration of 10 µg/mL, using the UV and the HPLC methods. Cefaclor monohydrate contents and the relative standard deviations (RSD) were calculated.

Accuracy

Cefaclor monohydrate reference standard was accurately weighed and added, at three different concentrations. At each concentration, samples were prepared in triplicate and the recovery percentage was determined by UV and HPLC methods.

Robustness

The robustness of the method was determined by the variation of the analyst and mobile phase flow rate. The flow rate was checked in 0.8 mL to 1.0 mL.

Analysis of cefaclor monohydrate powder for capsules and oral suspension

Samples of Medaclor, Oraclor were analyzed by the validated HPLC and UV methods. The sample solutions for the HPLC and UV analyses were prepared as described previously. The Cefaclor monohydrate contents were determined by using the two methods and the obtained results were statistically compared by using ANOVA test and Tukey's multiple comparison test, applied at 0.05 significance level.

Results and Discussion

During the chromatographic method development, Ultra Pure Water showed to be a more adequate organic solvent than Methanol, regarding the Cefaclor monohydrate retention. A typical chromatogram obtained is as shown by Figure 2.

After the evaluation of the Cefaclor monohydrate UV spectrum in various solvents (Ultra Pure water, methanol, (Ultra Pure Water: Methanol) (50:50) v/v%, hydrochloric acid 0.1M, and sodium hydroxide 0.1 M). In the range of 200-400 nm (Figure 3), the wavelength of 264 nm was chosen due to the adequate molar absorptivity of Cefaclor monohydrate in this region and to minimize possible interference from other compounds and solvents in the samples.

Validation

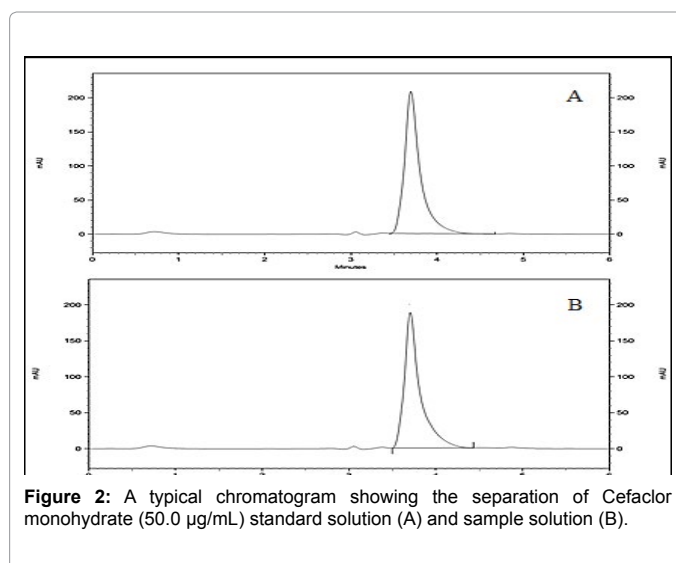


Figure 2: A typical chromatogram showing the separation of Cefaclor monohydrate (50.0 µg/mL) standard solution (A) and sample solution (B).

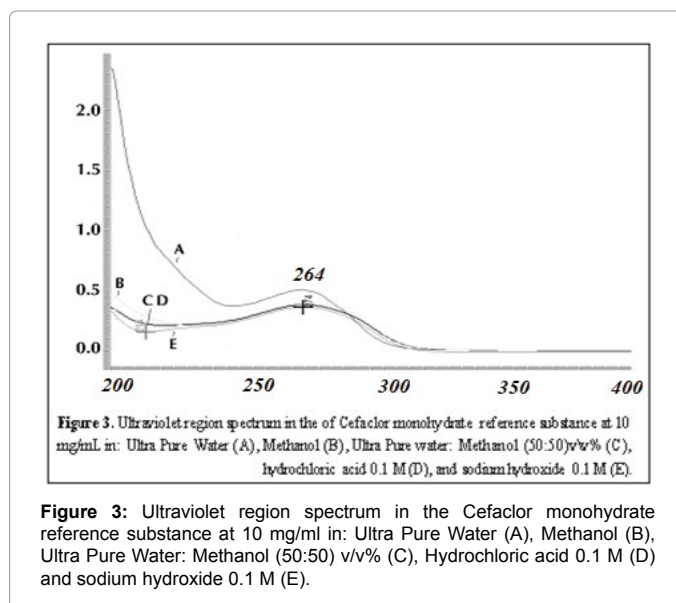


Figure 3: Ultraviolet region spectrum in the Cefaclor monohydrate reference substance at 10 mg/ml in: Ultra Pure Water (A), Methanol (B), Ultra Pure Water: Methanol (50:50) v/v% (C), Hydrochloric acid 0.1 M (D) and sodium hydroxide 0.1 M (E).

A linear relationship was found between the Cefaclor monohydrate concentrations and the response of both HPLC and UV methods. The regression analysis data are presented in Table 1. High regression coefficient (r^2) values were obtained (0.9995 and 0.9996, respectively). A random pattern of the regression residues was found and no significant deviation of linearity was detected in the assayed range.

The precision data obtained for the evaluated methods are demonstrated in Table 2. Both methods presented RSD values lower than 2.0%, assuring a good precision.

Accuracy (Table 2) was investigated by means of a standard addition experiment. Both chromatographic and spectrophotometric methods exhibited mean recoveries (n=9) close to 100% demonstrating an adequate accuracy.

The difference in the retention time, the peak area and the analyst (for a given Cefaclor monohydrate concentration) caused by the aforementioned minor alterations were insignificant (Table 2).

Regression parameters	HPLC	UV
Regression coefficient (r^2)	0.9995	0.9996
Slope \pm standard error	0.199 \pm 0.20	0.025 \pm 0.0017
Intercept \pm standard error	0.205 \pm 0.11	0.006 \pm 0.010
Relative standard error (%)	1.13	1.78
Concentration range ($\mu\text{g/mL}$)	0.1-80.0	1.0-60.0
Number of points	12	8

Table 1: Overview of the Linearity Data Obtained for Cefaclor monohydrate by the Chromatographic and Spectrophotometric Methods

Validation parameters	HPLC	UV
Intra-day precision, n=6 (RSD%)	1.13	1.78
Accuracy, n=9 (mean recovery, %) (10 $\mu\text{g/mL}$)	100.1	100.82

Table 2: Validation Parameters of the Evaluated Methods for Cefaclor monohydrate Determination.

Analyst	Area	Mean \pm SEM	RSD (%)
1	691545	688479 \pm 0.27	0.72
	682258		
	685912		
	691089		
	699896		
2	680178	634243 \pm 1.25	3.31
	691563		
	613157		
	599899		
	651955		
	630085		
	618799		

RSD=Relative Standard Deviation
SEM=Standard Error Mean

Table 3: Robustness of the HPLC Method for Cefaclor monohydrate by Varying the Analyst.

Analysis of capsules and oral suspensions cefaclor monohydrate

The validated chromatographic and spectrophotometric methods were applied to the analysis of Cefaclor monohydrate in Medaclor, Oraclor (Table 3). ANOVA test revealed a statistically significant difference between the results obtained for injectable samples, from the distinct methods, at a confidence level of 0.05. Chromatographic analysis showed to be the most sensitive and selective method, and might be applied successfully for Cefaclor monohydrate trace analysis and quantitation in biological matrices. We cannot discharge, however, the analyses time and cost. The spectrophotometric method is clearly less expensive and requires shorter analysis time, besides the ease of handling and lower residues generation.

Since the use of Cefaclor monohydrate as a potent antimicrobial drug is widespread, the development and validation of simple and reliable methods are essential to assure the quality of the raw materials and pharmaceutical formulations marketed nowadays. A simple method to identify and precisely quantify these drugs may be an important tool to avoid treatment inefficacy and development of resistance due to the exposition to sub therapeutic doses [35].

Conclusion

HPLC and UV spectrophotometry were found to be adequate methods to quantify Cefaclor monohydrate in Capsules and Oral suspensions solutions; the chromatographic and spectrophotometric methods presented the most reliable results. Since these methods are fast

and simple, they may be successfully applied to quality control analyses, with the aim of quantifying and identifying Cefaclor monohydrate in pharmaceutical products.

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References

- Brunton LL, Lazo JS, Parker KL (2006) Goodman and Gilman: As Bases Farmacológicas da Terapêutica, 11th edn. McGraw-Hill Book Co., New York, USA.
- Okamoto MP, Nakahiro RK, Chin A, Bedikian A, Gill MA (1994) Cefepime: a new fourth-generation cephalosporin. *Am J Hosp Pharm* 51: 463-477.
- Gutierrez Navarro P, Martinez de las Parras PJ, Marquez Garcia A (1991) Reaction of sodium amoxicillin with Cu(II) ion in a methanolic medium. *J Pharm Sci* 80: 904-907.
- Zuhri AZA, Rady AH, El-Shahawi MS, Al-Dhaheer S (1994) Spectrophotometric determination of ampicillin by ternary complex formation with, 10-phenantroline and copper(II). *Microchem J* 50: 111-115.
- Dimitrovska A, Andonovski B, Stojanoski K (1996) Spectro- photometric study of copper(II) ion complexes with cefaclor. *Int J Pharm* 134: 213-221.
- Ayad MM, Shalaby AA, Abdellatef HE, Elsaid HM (1999) Spectrophotometric determination of certain cephalosporins through oxidation with cerium (IV) and 1-chlorobenzotriazole. *J Pharm Biomed Anal* 20: 557-564.
- Al-Momani IF (2001) Spectrophotometric determination of selected cephalosporins in drug formulations using flow injection analysis. *J Pharm Biomed Anal* 25: 751-757.
- Mohamed GG (2001) Spectrophotometric determination of ampicillin, dicloxacin, flucloxacillin and amoxicillin antibiotic drugs: ion- pair formation with molybdenum and thiocyanate. *J Pharm Biomed Anal* 24: 561-567.
- Gallo Martinez L, Campins Falco P, Sevilano Cabeza A (2002) Comparison of several methods used for the determination of cephalosporins. Analysis of cephalixin in pharmaceutical samples. *J Pharm Biomed Anal* 29: 405-423.
- Salem H, Askal H (2002) Colourimetric and AAS determination of cephalosporins using Reineck's salt. *J Pharm Biomed Anal* 29: 347-354.
- El-Mamml MY (2003) Spectrophotometric determination of flucloxacillin in pharmaceutical preparations using some nitrophenols as a complexing agent. *Spectrochim Acta A Mol Biomol Spectrosc* 59: 771-776.
- Amin AS, Ragab GH (2004) Spectrophotometric determination of certain cephalosporins in pure form and in pharmaceutical formulations. *Spectrochim Acta* 60: 2831-2835.
- Aly HM, Amin AS (2007) Utilization of ion exchanger and spectrophotometry for assaying amoxycillin and flucloxacillin in dosage form. *Int J Pharm* 338: 225-230.
- Myers CM, Blumer JL (1983) Determination of ceftazidime in biological fluids by using high-pressure liquid chromatography. *Antimicrob Agents Chemother* 24: 343-346.
- Joshi S (2002) HPLC separation of antibiotics present in formulated and unformulated samples. *J Pharm Biomed Anal* 28: 795-809.
- Adamis G, Papaioannou MG, Giamarellos-Bourboulis EJ, Gargalianos P, Kosmidis J, et al. (2004) Pharmacokinetic interactions of ceftazidime, imipenem and aztreonam with amikacin in healthy volunteers. *Antimicrob Agents Chemother* 23: 144-149.
- Zivanovic L, Ivanovic I, Vladimirov S, Zecevic M (2004) Investigation of chromatographic conditions for the separation of cefuroxime axetil and its geometric isomer. *J Chromatogr B Analyt Technol Biomed Life Sci* 800: 175-179.
- Tozo GC, Salgado HR (2006) Determination of lomefloxacin in tablet preparations by liquid chromatography. *J AOAC Int* 89: 1305-1308.
- Moreno Ade H, Salgado HR (2008) Development of a new high-performance liquid chromatographic method for the determination of ceftazidime. *J AOAC Int* 91: 739-743.
- Flurer CL (2005) Analysis of antibiotics by capillary electrophoresis. *Electroph*

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- 18: 2427-2437.
21. Fabre H, Blanchin MD, Lerner D, Mandrou B (1985) Determination of cephalosporins utilising thin-layer chromatography with fluorescamine detection. *Analyst* 110: 775-778.
22. Farrell CD, Rowell FJ, Cumming RH (1995) A rapid fluorescence ELISA for ceftazidime. *Anal Proc* 32: 205-206.
23. Aly FA, Hefnawy MM, Belal F (1996) A selective spectro-fluorimetric method for the determination of cephalosporins in biological fluids. *Anal Lett* 29: 117-130.
24. Yang JH, Zhou GJ, Jie NQ, Han RJ, Lin CG, et al. (1996) Simultaneous determination of cephalexin and cephadroxil by using the coupling technique of synchronous fluorimetry and H- point standard additions method. *Anal Chim Acta* 325: 195-200.
25. Sengun FI, Ulas K, Fedai I (1985) Analytical investigations of cephalosporins-II. Polarographic behaviour of ceftriaxone, cefuroxime, cefotaxime and ceftizoxime and assay of their formula- tions. *J Pharm Biomed Anal* 3: 191-199.
26. Altinoz S, Ozer D, Temizer A (1994) Determination of ceftriaxone in aqueous humor and serum samples of differential-pulse adsorptive stripping voltammetry. *Analyst* 119: 1575-1577.
27. Billová S, Kizek R, Jelen F, Novotná P (1994) Square-wave voltametric determination of cefoperazone in a bacterial culture, pharmaceutical drug, milk and urine. *Anal Bioanal Chem* 377: 362-369.
28. Reddy GV, Reddy SJ (1997) Estimation of cephalosporin antibiotics by differential pulse polarography. *Talanta* 44: 627-631.
29. Ozkan SA, Erk N, Uslu B, Yilmaz N, Biryol I (2000) Study on electrooxidation of cefadroxil monohydrate and its determination by differential pulse voltammetry. *J Pharm Biomed Anal* 23: 263-273.
30. Fogg AG, Abadia MA, Henriques HP (1982) Titrimetric determina- tion of the yield of sulphide formed by alkaline degradation of cephalosporins. *Analyst* 107: 449-451.
31. Salgado HR, Tozo GC (2007) Microbiological assay for cefoxitin sodium in dosage form. *J AOAC Int* 90: 452-455.
32. De Haro MA, Salgado HR (2007) Microbiological assay for ceftazidime injection. *J AOAC Int* 90: 1379-1382.
33. AOAC (1990) Official Methods of Analytical Chemists of AOAC. 15th edn.
34. ICH (2005) International Conference on Harmonization. Validation of analytical procedures: methodology, Q2B (CPMP/ICH/281/95).
35. Souza MJ, Rolim CM, Melo J, Souza FPS, Bergold AM (2007) Development of a microbiological assay to determine the potency of ceftiofur sodium powder. *J AOAC Int* 90: 1724-1728.