

# Challenges and Drawbacks of DNA Sampling Using Swabs: A Comprehensive Analysis

Thomsen Robson\*

Department of Pure and Applied Chemistry, University of Strathclyde, Glasgow G1 1RD, UK

## Introduction

DNA sampling using swabs has become an indispensable tool in various fields such as forensic science, medical diagnostics, and genetic research. Swab-based DNA collection offers a non-invasive and relatively simple method for obtaining genetic material from individuals. However, despite its widespread use and advantages, this technique is not without its drawbacks and difficulties. In this article, we delve into the challenges associated with DNA sampling using swabs and explore potential solutions to mitigate these issues [1]. One of the primary challenges of DNA sampling using swabs is the risk of contamination. Swabs can easily pick up DNA from external sources, such as skin cells, dust particles, or environmental contaminants. Contamination can occur during sample collection, handling, or processing, leading to erroneous results and false conclusions.

To address contamination risks, strict protocols must be followed during sample collection and processing. This includes using sterile swabs, wearing gloves, and minimizing contact with potential sources of contamination. Additionally, laboratory facilities must adhere to stringent quality control measures to prevent cross-contamination between samples [2]. Another limitation of swab-based DNA sampling is the potential for low DNA yield, particularly in cases where the sample size is small or the DNA concentration is low. This can pose challenges in forensic investigations or genetic studies where the amount of DNA available for analysis is limited. To improve DNA yield from swab samples, researchers have developed various techniques and technologies, such as amplification methods like Polymerase Chain Reaction (PCR) or Whole Genome Amplification (WGA). These methods allow for the amplification of DNA from minute samples, enabling more comprehensive analysis and detection of genetic markers.

Even when DNA is successfully collected using swabs, there is a risk of incomplete DNA recovery. Factors such as improper sampling techniques, inadequate swabbing, or suboptimal storage conditions can result in partial or degraded DNA samples, limiting the accuracy and reliability of downstream analysis [3]. To enhance DNA recovery from swab samples, researchers are exploring novel extraction methods and improving sample processing protocols. For example, advancements in DNA extraction kits and automated extraction systems have shown promise in maximizing DNA recovery efficiency and minimizing sample loss.

## Literature Review

Swab samples collected from certain biological substrates may contain

\*Address for Correspondence: Thomsen Robson, Department of Pure and Applied Chemistry, University of Strathclyde, Glasgow G1 1RD, UK; E-mail: thomsenrobson@gmail.com

Copyright: © 2024 Robson T. This is an open-access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

Received: 24 January, 2024, Manuscript No. jfr-23-129346; Editor Assigned: 26 January, 2024, PreQC No. P-129346; Reviewed: 08 February, 2024, QC No. Q-129346; Revised: 14 February, 2024, Manuscript No. R-129346; Published: 24 February, 2024, DOI: 10.37421/2157-7145.2024.15.598

inhibitors that interfere with DNA analysis. Common inhibitors include substances like hematin, humic acids, or other contaminants present in blood, saliva, or environmental samples. These inhibitors can inhibit enzymatic reactions, affect PCR amplification, and compromise the integrity of DNA analysis. To mitigate the effects of inhibitors, researchers employ various strategies, such as sample dilution, enzymatic treatment, or purification techniques. Additionally, using specialized DNA extraction kits designed to remove inhibitors can help improve the quality of DNA extracted from challenging samples [4]. Maintaining the stability and integrity of swab samples during storage is crucial for preserving DNA quality and preventing degradation. However, inadequate storage conditions, such as exposure to temperature fluctuations, humidity, or UV radiation, can compromise DNA stability and lead to sample deterioration over time.

## Discussion

To ensure sample stability, proper storage protocols must be followed, including storing samples in appropriate containers, at optimal temperatures, and in controlled environments. Additionally, the use of preservatives or stabilizing agents in swab collection kits can help prolong DNA shelf life and maintain sample integrity during storage and transportation [5]. In addition to technical challenges, DNA sampling using swabs raises ethical and legal considerations regarding privacy, consent, and data protection. Collecting DNA samples from individuals without their informed consent or using genetic information for purposes other than those disclosed may infringe upon individual rights and raise concerns about potential misuse or discrimination. To address ethical and legal concerns, strict guidelines and regulations govern the collection, storage, and use of DNA samples for research or forensic purposes. These guidelines emphasize obtaining informed consent from participants, ensuring confidentiality and anonymity, and implementing robust data protection measures to safeguard genetic information [6].

## Conclusion

DNA sampling using swabs has revolutionized various fields, offering a convenient and non-invasive method for obtaining genetic material. However, this technique is not without its challenges and drawbacks, including contamination risks, low DNA yield, incomplete DNA recovery, presence of inhibitors, storage issues, and ethical considerations. By addressing these challenges through innovative technologies, rigorous protocols, and adherence to ethical standards, researchers can maximize the potential of swab-based DNA sampling while ensuring accurate and reliable results for various applications.

## Acknowledgement

None.

## Conflict of Interest

There is no conflict of interest by author.

---

## References

1. Van Oorschot, Roland AH, Timothy J. Verdon and Kaye N. Ballantyne. "Collection of samples for DNA analysis." *Forens DNA Typing Proto* (2016): 1-12.
2. Verdon, Timothy J., Robert J. Mitchell and Roland AH van Oorschot. "Swabs as DNA collection devices for sampling different biological materials from different substrates." *JFS* 59 (2014): 1080-1089.
3. Vashist, Vedant, Neil Banthia, Swapnil Kumar and Prajwal Agrawal. "A systematic review on materials, design, and manufacturing of swabs." *Ann 3D Print Med* 9 (2023): 100092.
4. Lee, Rose A., Joshua C. Herigon, Andrea Benedetti and Nira R. Pollock, et al. "Performance of saliva, oropharyngeal swabs, and nasal swabs for SARS-CoV-2 molecular detection: A systematic review and meta-analysis." *J Clin Microbiol* 59 (2021): 10-1128.
5. Van Oorschot, R. A. H., D. G. Phelan, S. Furlong and G. M. Scarfo, et al. "Are you collecting all the available DNA from touched objects?." *Int Cong Ser* 1239 (2003): 803-807.
6. Bruijns, Brigitte B., Roald M. Tiggelaar and Han Gardeniers. "The extraction and recovery efficiency of pure DNA for different types of swabs." *JFS* 63 (2018): 1492-1499.

**How to cite this article:** Robson, Thomsen. "Challenges and Drawbacks of DNA Sampling Using Swabs: A Comprehensive Analysis." *J Forensic Res* 15 (2024): 598.