Open Access

Association of Serum and Tumor Tissue microRNA Profile with Aggressiveness of Papillary Thyroid Carcinoma in an Iranian Population

Sara Nazari¹, Ahmad Majd¹, Iraj Heydari², Mohammad Reza Mohajeri Tehrani³, and Reza Nekouian^{4*}

¹Department of Cellular and Molecular Biology, Faculty of Biological Sciences, Islamic Azad University, North Tehran Branch, Tehran, Iran ²Endocrine Research Center, Institute of Endocrinology and Metabolism, Iran University of Medical Sciences, Tehran, Iran ³Endocrinology and Metabolism Research Center, Endocrinology and Metabolism Clinical Sciences Institute, Tehran University of Medical Sciences, Tehran, Iran ⁴Pediatric Growth and Development Research Center, Institute of Endocrinology and Metabolism, Iran University of Medical Sciences, Tehran, Iran

Abstract

Objectives: Papillary thyroid carcinoma (PTC) is the most common malignancy of thyroid. We aimed to investigate the association of *let-7f*, *miR-146b-5p*, *miR-34b*, *miR-16* and *miR-877-5p* expression in blood circulation and tumor with aggressiveness of PTC.

Methods: A total of 18 patients with aggressive PTC and 18 patients with non-aggressive PTC were studied. The microRNAs expressions were evaluated using real-time PCR. Fold changes (FC) of the miRs in aggressive PTC patients were calculated via calibration with mean of expression of the miRs in non-aggressive groups.

Results: *MiR-16* showed significant up regulation in blood (FC=2.85; P=0.024), *miR-34* showed significant down regulation in blood (FC=0.19; P<0.001) and tumor tissue (FC=0.19; P<0.001), *miR-146* showed significant up regulation in blood (FC=48.10; P<0.001) and tumor tissue (FC=60.61; P<0.001), *miR-877* showed significant down regulation in blood (FC=0.22; P<0.001), and *let-7* showed significant down regulation in blood (FC=0.13; P<0.001).

Conclusion: In general, our study in an Iranian population supported the previous results. Up regulation of *miR*-146 was associated with aggressiveness of PTC.

Keywords: Papillary thyroid carcinoma • microRNA • Aggressive tumor • mir-146b

Introduction

Papillary thyroid carcinoma (PTC) is the most common malignancy of thyroid gland originating from thyroid follicular cells. PTC has generally a good survival rate; however the cases having certain clinico-pathological parameters have poorer prognosis [1]. A population-based cohort study showed that incidence of total thyroid cancers increased from 3.6 per 100000 in 1973 to 8.7 per 100000 in 2002. No significant change was reported for incidence of the less common types including follicular, medullary, and anaplastic carcinomas. In other words the entire increase is attributable to an increase in incidence of PTC, which increased from 2.7 to 7.7 per 100000. Crude death rate of PTC was approximately 0.5 deaths per 100000 individuals from 1973 to 2002 [2]. Point mutations in proto-oncogenes including rearranged during transformation (*RET*), *BRAF*, *V-Raf* and rat sarcoma viral oncogene homolog (*RAS*) were frequently observed in PTC [3].

microRNAs (miRs) are endogenous single stranded non coding RNAs that bind to the 3' non coding region of the target mRNAs, resulting in their selective degradation or inhibition of translation. Therefore miRs are involved in regulation of biological functions [4]. *Let-7* family is known as the first group of discovered miRs in human. It has been observed that down regulation

*Address for Correspondence: Nekouian R, Pediatric Growth and Development Research Center, Institute of Endocrinology and Metabolism, Iran University of Medical Sciences, Tehran, Iran, E-mail: nekouian.r@iums.ac.ir

Copyright: © 2021 Nazari S, et al. This is an open-access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

Received 14 December 2020; Accepted 22 January 2021; Published 29 January, 2021

of these miRs is associated with malignancies [5]. *Let-7* miRs bind to RAS oncogenes and result in their down regulation. Therefore they have been investigated in PTC. In addition, this family has been observed in normal thyroid gland suggesting that it may play a role in function of thyroid gland [6]. Other than *let-7* family, other miRs were notable. MiR-146b regulates signal transduction of transforming growth factor-beta (TGF- β) and therefore may play a role in PTC. In addition, oncogene activation has resulted in increase in expression of miR-146b [7]. It has been observed that patients with *BRAF* mutation have a higher expression level of miR-146b in comparison to patients with wild type of *BRAF* [8]. MiR-34b is involved in oncogenesis. In has been observed that it was down regulated in cancer cells [9]. A study on different cancer cell lines showed that many genes such as *RAS* family are targeted by miR-16 [10]. MiR-877 is another cancer related miR which is down regulated in hepatocellular carcinoma via targeting cyclin dependent kinase (CDK) 14 [11].

Previously diagnostic role of miRs in PTC have been investigated [12]. However, other than diagnosis, it is important to have ability to differentiate aggressive cases from the non-aggressive ones. Therefore, the present study was designed to investigate the association of *let-7f*, *miR-146b-5p*, *miR-34b*, *miR-16* and *miR-877-5p* expression in blood serum and tumor tissue with aggressiveness of PTC in an Iranian population.

Materials and Methods

Study design and patients

The present work was a case control study to compare blood and tumor tissue expression of the miRs between aggressive and non-aggressive patients of PTC using reverse transcriptase quantitative polymerase chain reaction (RT-qPCR). A total of 18 patients with aggressive PTC were considered as the case group and 18 patients with non-aggressive PTC were considered as the control group. The samples were collected from 3 centers Toos, Shariyati and Firoozgar (Tehran, Iran) from February to September 2019 through convenient sampling. This study was approved by the ethics committee of Shariyati hospital (IR.TUMS.EMRI.REC.1398.008). Written informed consents were obtained from all the participants.

Blood and tumor tissue samples collection

Fresh tissue specimens of aggressive and non-aggressive PTC tissues were obtained from tumors and immediately kept at -80°C until analysis. All the samples were diagnosed by two independent pathologists. Confirmation of PTC was through light microscope using hematoxylin and eosin staining. Clinical staging was according to prognostic factor for recurrence in N1b [13]. From each participant, 2 ml of peripheral blood was collected in EDTA containing tubes and immediately kept at -80°C until analysis.

Total RNA extraction

Total RNA was obtained from the tumors and serum with columnar extraction method using total RNA purification kit (Norgen, Canada) according to the manufacturer's instructions. Total RNA quantification was performed using nanodrop spectrophotometry (Thermo Scientific Wilmington, DE, USA). Sample quality control using cel-mir-39 Spike-In Kit (Norgen, Canada) offers a quantified synthetic RNA (cel-miR-39) for spike-in during RNA extraction procedures and subsequent normalization in RT-qPCR assays. The amount of cel-miR-39 RNA recovered after RNA extraction is directly correlated with the amount of total RNA recovered. Therefore cel-miR-39 was used as the reference to report Δ Ct.

miR reverse transcriptase PCR

cDNA was synthesized with poly A polymerase method using microScript microRNA cDNA Synthesis kit (Norgen, Canada). Then the cDNA was subjected to be used for real-time reverse transcriptase polymerase chain reaction (RT-PCR) using SYBR Green mastermix (Norgen, Canada) according to the manufacturer's instructions (after a first 30 min at 37°C, then 30 min at 50°C and finally 15 min at 70°C). All RT-PCR reactions were performed in triplicates. Levels of miR expression were calculated by relative quantification using Rotor Gene Q Real-Time PCR SDS 2.3.1 software (Applied Biosystems Inc., Foster city, CA). The results were presented as normalized Ct values. Previously published literatures were used for primary selection of the miRs. miR cancer database was used to find associations of miRs with cancers according to the literatures of PubMed. The candidate miRs were let-7f, miR-146b-5p, miR-34b, miR-16 and miR-877-5p. Mirbase database was used to find the sequences of the miRs in order to convert them to primer sequence. The used primers are shown (Table 1). miRDB database was used to find the targeted genes. According to this, these miRs can target RAS and Braf. Mirandola database was used to understand whether the miRs could be detected in circulating blood and the tissue. Bioinformatics characteristics of the used miRs are shown (Table 2).

Statistical analysis

Rest 2009 was used to investigate relative expression. Fold changes of each miR were calculated via the formula $2^{-\Delta\Delta CT}$ in Excel 2013 calibrating with the mean of expression in non-aggressive PTC patients. Significance of

Table 1. Sequences of the primers used in this study.

miR	Primers		
Mir16-5p	TAGCAGCGTAAATATTGGCG		
Mir34b-5p	TAGGCAGTGTCATTAGCTGATTG		
Mir146b-5p	TGAGAACTGAATTCCATAGGCTG		
Mir877-5p	GTAGAGATGGCGCAGGG		
Let7f-5P	TGAGGTAGTAGATTGATAGTT		

Table 2. Bioinformatics of the used miRs.

MicroRNA	Location	Target	Score	Position	Sequence	
Mir-16-5p	Chr 13: 50,048,973- 50,049,061	PLSCR4	70	1568-1575 of PLSCR4 3' UT R	UAGCAGCACGUAAAUAUUGGCG	
		ANO3	98	2517-2523 of ANO3 3' UTR		
		PHF19	100	1377-1383 of PHF19 3' UTR		
		AQP11	65	227-234 of AQP11 3' UTR		
	chr10: 102,436,512- 102,436,584	NOVA1	98	682-689 of NOVA1 3' UTR		
Mir-146b-5p		TRAF6	100	1272-1279 of TRAF6 3' UTR	UGAGAACUGAAUUCCAUAGGCUG	
MII-1460-5p		CD80	90	839-846 of CD80 3' UTR	UGAGAACUGAAUUCCAUAGGCUG	
		SEC23IP	99	3634-3641 of SEC23IP 3' UTR		
	chr6: 30584332- 30584417	ZNF174	89	329-336 of ZNF174 3' UTR		
Mir-877-5p		TP53INP2	80	3069-3075 of TP53INP2 3' UTR	GUAGAGGAGAUGGCGCAGGG	
wiii-o <i>i i</i> -op		RP11-204N11.1	80	2289-2296 of RP11-204N11.1 3' UTR	GUAGAGGAGAUGGCGCAGGG	
		TMCC2	67	1200-1207 of TMCC2 3' UTR		
	chr11: 111512938- 111513021	TENM1	100	1314-1321 of TENM1 3' UTR		
Mir 2/1h En		ELMOD1	98	1100-1106 of ELMOD1 3' UTR	UAGGCAGUGUCAUUAGCUGAUUG	
Mir-34b-5p		RFX3	98	3622-3628 of RFX3 3' UTR	UAGGCAGUGUCAUUAGCUGAUUG	
		DLL1	97	294-300 of DLL1 3' UTR		
Let-7f-5p	chr9: 94176347- 94176433	STARD13	100	Position 2362-2368 of STARD13 3' UTR	UGAGGUAGUAGAUUGUAUAGUI	
		C14orf28	100	437-444 of C14orf28 3' UTR		
		LIN28B	100	44-51 of LIN28B 3' UTR	UGAGGUAGUAGAUUGUAUAGUU	
		BZW1	92	84-90 of BZW1 3' UTR		

individual fold changes was investigated with one sample t test (fold change=1 was the null hypothesis) and comparison of the fold changes between blood serum and tumor tissue was through independent t test. SPSS 24 software (IBM, US) was used for data analysis. Two-tailed P value less than 0.05 was considered as the significance level.

Results

A total of 36 Iranian patients of PTC with age range 20-72 were investigated. The range of the tumor size among the PTC patients was 0.5-8.0 cm with number of lymph node metastasis ranged 0-5 (Table 3). Real-time RT-PCR was performed and after approving the melting curves, fold changes were compared between blood serum and tumor tissue expression. Up regulation and down regulation of the miRs based on relative expression were calculated with rest program. This relative expression was calculated for expression of the miRs as aggressive versus non-aggressive groups (Table 4). Fold changes of the miRs in aggressive PTC patients were calculated via calibration with mean of expression of the miRs in non-aggressive groups using one sample t test. According to this, MiR-16 showed significant up regulation in blood (fold change [FC]=2.85; P=0.024), miR-34 showed significant down regulation in blood (FC=0.19; P<0.001) and tumor tissue (FC=0.19; P<0.001), miR-146 showed significant up regulation in blood (FC=48.10; P<0.001) and tumor tissue (FC=60.61; P<0.001), miR-877 showed significant down regulation in blood (FC=0.22; P<0.001), and let-7 showed significant down regulation in blood (FC=0.09; P<0.001) and tumor tissue (FC=0.13; P<0.001). Fold change of each miR (blood expression versus tumor tissue expression) was compared using independent t test. MiR-16 showed a significant more up regulation in aggressive PTC patients (2.85 vs. 0.92; P=0.020). No significant difference was observed between blood and tumor tissue fold changes for other miRs (Table 5 and Figure 1).

Discussion

The present study was designed in order to find the role of circulating and tumor tissue miRs in invasion of PTC. At this level we could find significant association of 3 miRs with invasion of PTC in both blood and tumor tissue, and association of 2 miRs with invasion of PTC in blood. *MiR-16* showed significant up regulation in blood, *miR-34* showed significant down regulation in blood and tumor tissue, *miR-146* showed significant up regulation in blood and tumor tissue, *miR-146* showed significant up regulation in blood and tumor tissue, *miR-146* showed significant up regulation in blood, and *let-7* showed significant down regulation in blood, and *let-7* showed significant down regulation in blood and tumor tissue. Expression change of *miR-16* was significantly more dominant in blood in favor of up regulation. Since there was no significant difference between the fold changes of the miRs in blood and tissue (except *miR-16*), blood serum expression study of these miRs can be used in clinics as an available and representative source instead of tumor tissue.

PTC is a cancer with good prognosis. The necessity of total thyroidectomy may be affected by its aggressive behavior. Since aggressive behavior of PTC was hard to predict, many studies were trying to have research on biomarkers. Linwah et al. studied 17 aggressive and 15 non-aggressive PTC patients in USA in order to find the role of tissue miR signature. They found up regulation for *miR-146b*, -221, -222, -155, -31, and down regulation for *miR-1*, -34b, -130b, -138 in aggressive PTC. Our study supported this finding for the common *miRs* [14]. Yang et al. studied 20 aggressive and 20 non-aggressive patients in China. They found up regulation of miR 146b 5p and *miR* 221/222 and down regulation of *miR* 16 and *miR* 613 in aggressive PTC. In contrast,

Table 3. Clinico-pathological features of the participants.

Variables	Group 1 (Aggressive PTC)	Group 2 (Non-aggressive PTC)	
	Gender		
Female	13	13	
Male	5	5	
	Age		
45>	4	6	
45<	14	12	
	Pathological characteristics		
Metastatic lymph node	03-May	0-4	
Tumor size	2.5-8 cm	0.5-4 cm	
TNM staging	III, IV, V	I, II	
Lobectomy	2	3	
Thyroidectomy	16	15	

Table 4. Comparison of relative expressions with their 95% confidence intervals are shown. *Significant at P<0.05.

miR	Source	Relative expression (95% CI)	P value (Rest)	Effect direction
Mis10 En	Tissue	1.206 (0.206-6.571)	0.407	Non-significant
Mir16-5p —	Blood	1.162 (0.214-5.852)	0.485	Non-significant
Mir24h En	Tissue	0.289 (0.010-4.225)	0.001*	Down regulation
Mir34b-5p —	Blood	0.294 (0.018-5.513)	0.002*	Down regulation
Nix1//Ch En	Tissue	6.834 (1.007-62.775)	<0.001*	Up regulation
Mir146b-5p –	Blood	4.289 (0.350-35.198)	<0.001*	Up regulation
Mir077 En	Tissue	1.096 (0.082-13.296)	0.774	Non-significant
Mir877-5p —	Blood	0.858 (0.056-20.990)	0.661	Non-significant
L at 76 ED	Tissue	0.555 (0.065-4.324)	0.028*	Down regulation
Let7f-5P —	Blood	0.108 (0.012-1.253)	<0.001*	Down regulation

Gro	up	Ν	Mean of fold change	Std. Deviation	Std. Error Mean	One sample t test P value	Independent t test P value	
mir_16	blood	18	2.854247	3.1674688	0.7465796	0.024*	0.020*	
	tissue	18	0.920071	1.0990411	0.2590465	0.761	0.020	
mir 0/	blood	18	0.198956	0.1896811	0.0447083	0.000*	- 0.377	
mir_34	tissue	18	0.286675	0.3702283	0.0872637	0.000*		
	blood	18	48.101287	30.1388044	7.1037843	0.000*	0.239	
mir_146	tissue	18	60.617704	32.4837004	7.6564816	0.000*		
	blood	18	0.220025	0.1853129	0.0436787	0.000*	0.000	
mir_877 -	tissue	18	0.700225	1.144368	0.2697301	0.282	0.088	
let_7	blood	18	0.093891	0.0760213	0.0179184	0.000*	0.005	
	tissue	18	0.136635	0.1406197	0.0331444	0.000*	0.265	

Table 5. Fold changes of the miRs in aggressive PTC and comparison of them in blood and tissue. *Significant at P<0.05.

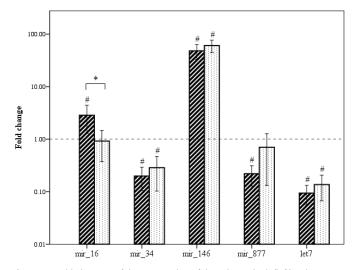


Figure 1. Fold changes of the expression of the miRs. Black (left) columns are for blood expression and white (right) columns are for tissue expression. The error bars indicate 95% confidence interval. The reference line fold change =1 shows calibration line (aggressive PTC vs non-aggressive PTC). *Significant at P<0.05; independent t-test. #Significant at P<0.05; one sample t test (the baseline fold change=1 is the null hypothesis).

our study did not show down regulation for *miR* 16 [4]. Lee et al. in Australia found that *miR-222* and *miR-146b* had over expression in PTC via comparing tumor tissue and plasma of 9 recurrent and 17 non-recurrent PTC patients [15]. Rosignolo et al. in Italy found association of *miR* 146*b*-5*p* and *miR* 222 3*p* up regulation with increased risk of recurrence [16]. Chou et al. introduced *miR-146b* as novel biomarker in PTC. They believed that this miR was associated with aggressiveness and prognosis. Different mechanisms had been suggested for the initiating role of *miR* 146*b* in oncogenic pathways. One of them was that *miR* 146*b* inhibited TGF- β anti-signal via down regulation of *SMAD4* and therefore inhibition of cell cycle arrest [17].

The most important limitation of this study and the previous studies was the study design. Although such studies named their groups as cohorts of PTC, but from the critical appraisal point of view in evidence-based medicine, they were not eligible cohort studies. Since the practical aim of this topic is prediction of aggressiveness, occurrence of PTC and its aggression should be subsequent to these biomarker changes.

Conclusion

In general, our study in an Iranian population supported the previous results. miRs can help to differentiate invasive PTC from non-invasive PTC. Briefly, *miR-16* and *miR-877* showed up regulation in blood, *miR-34* and *let-7*

showed down regulation in blood and tumor tissue, *miR*-146 showed significant up regulation in blood and tumor tissue, and *miR*-146 showed significant up regulation in blood and tumor tissue. Expression change of *miR*-16 was significantly more dominant in blood in favor of up regulation. Predicting role of these biomarkers should be investigated in well-designed cohort studies. Then the results can be used as personalized medicine in management of PTC.

Acknowledgments

We take it upon ourselves to appreciate all hospitals help us for sample collection. We acknowledge Firoozgar Clinical Research Development Center (FCRDC) of Iran University of Medical Sciences.

Funding

Funding is being done by Islamic Azad University, North Tehran Branch, as a PhD thesis.

References

- Nurul-Syakima Ab Mutalib, Sri Noraima Othman, Azliana Mohamad Yusofa and Shahrun Niza Abdullah Suhaimi, et al. "Integrated microRNA, gene expression and transcription factors signature in papillary thyroid cancer with lymph node metastasis." *Peer J* 4 (2016): e2119.
- Louise Davies, and Gilbert H. Welch. "Increasing incidence of thyroid cancer in the United States, 1973-2002." J Am Med Assoc 295 (2006): 2164-2167.
- Yasuko Kikuchi, Eiichi Tsuji, Koichi Yagia and Keisuke Matsusaka, et al. "Aberrantly methylated genes in human papillary thyroid cancer and their association with BRAF/RAS mutation." Front Genet 4 (2013): 271.
- Zhili Yang, Ziming Yuan, Youben Fana and Xianzhao Deng, et al. "Integrated analyses of microRNA and mRNA expression profiles in aggressive papillary thyroid carcinoma." *Mol Med Rep* 8 (2013): 1353-1358.
- Sarah Roush and Frank J. Slack. "The let-7 family of microRNAs." Trends Cell Biol 18 (2008): 505-516.
- Ewelina Perdas, Robert Stawski, Dariusz Nowak and Maria Zubrzycka. "The role of miRNA in papillary thyroid cancer in the context of miRNA Let-7 family." Int J Mol Sci 17 (2016): 909.
- Murilo Vieira Geraldo, Alex Shimura Yamashita and Edna Teruko Kimura. "microRNA miR-146b-5p regulates signal transduction of TGF-β by repressing SMAD4 in thyroid cancer." Oncogene 31 (2012): 1910-1922.
- Chen-Kai Chou, Rong-Fu Chen, Fong-Fu Choua and Hsueh-Wen Chang, et al. "miR-146b is highly expressed in adult papillary thyroid carcinomas with high risk features including extrathyroidal invasion and the BRAFV600E mutation." *Thyroid* 20 (2010): 489-494.

- Yi-Jia Li, Li Du, Grace Aldana-Masangkaya and Xiuli Wang, et al. "Regulation of miR-34b/c-targeted gene expression program by SUMOylation." *Nucleic Acids Res* 46 (2018): 7108-7123.
- Xin Yan, Hongwei Liang, Ting Deng and Kegan Zhu, et al. "The identification of novel targets of miR-16 and characterization of their biological functions in cancer cells." *Mol Cancer* 12 (2013): 1-11.
- Zhiming Tan, Chunguang Qiu, Jifan Sun and Wangxun Li. "MiR-877-5p suppresses cell growth, migration and invasion by targeting cyclin dependent kinase 14 and predicts prognosis in hepatocellular carcinoma." *Eur Rev Med Pharmacol Sci* 22 (2018): 3038-3046.
- Lutske Lodewijk, Anne M. Prins, Jakob W. Kist and Gerlof D. Valk, et al. "The value of miRNA in diagnosing thyroid cancer: a systematic review." *Cancer Biomark* 11 (2012): 229-238.
- 13. Young Jae Ryu, Jin Seong Cho, Jung Han Yoon and Min Ho Park. "Identifying

risk factors for recurrence of papillary thyroid cancer in patients who underwent modified radical neck dissection." *World J Surg Oncol* 16 (2018): 1-9.

- Linwah Yip, Lindsey Kelly, Yongli Shuai and Michaele J. Armstrong, et al. "microRNA signature distinguishes the degree of aggressiveness of papillary thyroid carcinoma." Ann Surg Oncol 18 (2011): 2035-2041.
- James C Lee, Jing Ting Zhao, Roderick J. Clifton-Bligh and Anthony Gill, et al. "microRNA-222 and microRNA-146b are tissue and circulating biomarkers of recurrent papillary thyroid cancer." *Cancer* 119 (2013): 4358-4365.
- Francesca Rosignolo, Lorenzo Memeo, Fabio Monzani and Cristina Colarossi, et al. "microRNA-based molecular classification of papillary thyroid carcinoma." Int J Oncol 50 (2017): 1767-1777.
- Chen-Kai Chou, Rue-Tsuan Liu and Hong-Yo Kang. "microRNA-146b: A novel biomarker and therapeutic target for human papillary thyroid cancer." Int J Mol Sci 18 (2017): 636.

How to cite this article: Sara Nazari, Ahmad Majd, Iraj Heydari and Mohammad Reza Mohajeri Tehrani, et al. "Association of Serum and Tumor Tissue microRNA Profile with Aggressiveness of Papillary Thyroid Carcinoma in an Iranian Population." *J Mol Biomark Diagn* 12 (2021): 448.