A Rapid, High-Throughput Iodometric Titration Method for the Determination of Active Chlorine Content of Topical Antiseptic Solutions

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Abstract

Objective: A considerable number of commercially available topical antiseptic solutions rely on free available chlorine as an active ingredient due to its broad-spectrum antimicrobial activity, but limited empirical knowledge exists of the degradation kinetics of chlorine-based solutions in contact with host tissue. To better inform clinical practice, we developed and qualified a rapid and sensitive semi-automated microtiter plate-based iodometric titration assay suitable for the rapid determination of free available chlorine in small samples of dilute antiseptic solutions following contact with biological materials.

Methods: The chlorine determination method described here utilizes a novel stepwise iodometric titration approach performed entirely on a plate reader spectrophotometer equipped with a standard automatic syringe dispenser module. In this method, both titrant addition and colorimetric monitoring steps are carried out automatically, providing significantly higher sample throughput with reduced technical error when compared to the manual titration approach typically used for this type of analysis. Assay qualification was performed by measuring free available chlorine in commercial Dakin’s solution at a range of concentrations during contact with human plasma in vitro and rat muscle tissue in a simulated wound model (ex vivo).

Results: The practical lower limit of quantitation for a 200 µl sample using this assay was found to be approximately 0.001% mass available chlorine and agreement was excellent between measured and nominal percent mass available chlorine over the range concentrations tested. Contact with biological material was found to cause loss of reactive chlorine in Dakin’s solution in seconds to minutes.

Conclusion: The semi-automated available chlorine determination method described here represents numerous improvements to traditional iodometric titration approaches by substantially decreasing required sample volume, drastically increasing throughput, and minimizing manual sample handling and error. We feel this novel method will be of value to other researchers investigating the degradation kinetics of chlorine-based solutions to improve clinical practice.

Keywords: Topical antiseptic; Chlorine activity; Automated titration; Dakin’s solution; Antimicrobial agent; High-throughput microtiter plate assay

Introduction

Numerous commercially available topical antiseptic solutions rely on free available chlorine as an active ingredient, typically in the form of dilute hypochlorous acid (HOCl) or sodium hypochlorite (NaClO), due to its broad-spectrum antimicrobial activity. Dakin’s solution (DS) is a buffered NaClO preparation that represents the prototypical example of this class of agents. DS was introduced more than a century ago to treat infected wounds in combat casualties during the First World War and remains available on the commercial market (Century Pharmaceuticals, Indianapolis, IN) in conjunction with newer hypochlorite and hypochlorous acid solutions. Modern NaClO and HOCl based preparations are typically marketed as antiseptic skin and wound cleansers or negative pressure wound therapy instillation solutions; current examples include PhaseOne Pure Hypochlorous SolutionTM (Integrated Healing Technologies, Franklin, TN), Vashe Wound SolutionTM (SteadMed Medical, Fort Worth, TX), Microcyn Negative Pressure Wound Therapy SolutionTM (Sonoma Pharmaceuticals, Petaluma, CA), Neutrophase Skin and Wound CleanserTM (Novabay Pharmaceuticals, Emeryville, CA), and Puracyn PlusTM (Innovacyn, Rialto, CA). Although systemic antibiotic therapy has supplanted topical antiseptics as a primary treatment modality for the management of skin and soft tissue infection, these agents remain in clinical use as adjunctive therapies for the management of chronic open wound [1-4] or in the case of DS, as a salvage therapy for Fournier’s gangrene [5] and invasive fungal infections of soft tissue wounds and burns of trauma patients [6].

The antimicrobial activity of chlorine-based solutions is specifically determined by free available chlorine content, i.e. the fraction of unreacted chlorine species available to oxidize or chlorinate organic biomolecules. Hypochlorous acid is the primary active chemical species in both “pure” HOCl preparations and sodium hypochlorite-
Materials and Methods

Mechanistically, the dysfunction of glucose metabolism and arrest of activity compared to anionic hypochlorite due to the greater ability of the uncharged species to penetrate microbial membranes [7,12]. Given the highly reactive nature of free chlorine, prudent use of NaClO and HOCl-based antimicrobial preparations implies a balance between antimicrobial activity and human tissue toxicity [19-21]. Limited empirical knowledge exists, however, of the degradation kinetics of chlorine-based solutions in contact with host tissue. In order to better inform clinical practice, we developed a rapid and sensitive semi-automated microtiter plate-based iodometric titration assay suitable for the rapid determination of free available chlorine in small samples of dilute antiseptic solutions following contact with biological materials. Results of the application of this high-throughput method to quantify the loss of Dakin's solution activity over time due to contact with human plasma in vitro, and rodent tissue ex vivo, are also described.

Materials and Methods

Sodium hypochlorite, potassium iodide, sodium thiosulfate standard solution, and partially saponified polyvinyl alcohol (MW 31,000-50,000; 87-89% hydrolyzed) were purchased from Sigma-Aldrich (St. Louis, MO). 0.9% sodium chloride solution was obtained from Baxter Healthcare Corporation (Deerfield, IL). Synergy 2 spectrophotometer equipped with an automatic microliter-scale syringe dispenser module (P/N 7090568) was purchased from BioTek Instruments (Winooski, VT). Commercial Dakin's solution (“Full Strength”; 0.5% NaClO) was purchased from Century Pharmaceuticals (Indianapolis, IN). Human plasma was purchased from Biological Specialty Corporation (Colmar, PA).

Free available chlorine assay

A 200 µl sample of dilute sodium hypochlorite solution (0.00025% to 0.125% mass NaClO) was mixed with 170 µl of a freshly prepared reaction mixture consisting of 0.5 ml of 2 M HCl, 1 ml of 0.6 M potassium iodide, and 200 µl of polyvinyl alcohol (PVA) indicator solution (5% PVA by weight in water). Reactions were carried out at room temperature in sealed opaque black 0.5 ml tubes in order to reduce photooxidation and chlorine loss through degassing to excessive headspace volume. Sample tubes were immediately mixed and centrifuged (1 min at 3000 RCF) prior to loading 100 µl of sample into triplicate wells of a Corning 96-well plate. The loaded plate was placed in a Biotek Synergy 2 spectrophotometer equipped with a Biotek automatic microliter-scale syringe dispenser module and baseline absorbance was measured at 490 nm. Sodium thiosulfate (Na$_2$S$_2$O$_3$) titrant (0.0001 M to 0.1 M, depending on the starting concentration of the NaClO solution being titrated) was added to each well in a stepwise fashion (5 µl per addition) using the integrated syringe dispenser module. The plate was mixed for 10 seconds following each titrant addition and absorbance was measured at 490 nm. The spectrophotometer control software was configured to automatically repeat the titrant addition, mixing and absorbance measurement steps sequentially for a total of 20 cycles. Equivalence was determined by monitoring the decrease in 490 nm absorbance following each automated titrant addition step. Absorbance of reaction mix control wells (without NaClO or sodium thiosulfate addition) was subtracted from sample values at each measurement time point to account for autoxidation of iodide by atmospheric oxygen at low pH. Available chlorine content of each sample, expressed as percent mass chlorine, was calculated via Equations 4 and 5 (after accounting for the dilution of the primary sample with reaction mix) using the known sodium thiosulfate titrant concentration and the volume needed to achieve equivalence, i.e., the number of titrant addition steps required for control-adjusted 490 nm absorbance to reach zero multiplied by the dispensation volume (Figure 1).

![Figure 1: Representative assay titration curve illustrating equivalence determination. Data represents mean ± SEM of N=3 technical replicates.](image-url)

Calculated active chlorine concentration was corrected for temperature-dependent iodine extraction efficiency using the method described by Yoshinaga et al. [22].

(Eq. 4)....

Concentration Cl$_2$=Volume (Na$_2$S$_2$O$_3$) x Concentration (Na$_2$S$_2$O$_3$)/Sample Volume x 2

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% Mass Available Cl₂ = Molar Mass (Cl₂) × Concentration (Cl₂)/Sample Specific Gravity × 10

Assay qualification

To evaluate the suitability of this assay for measuring the degradation kinetics of commercially available chlorine-containing solutions, the procedure described above was used to quantify the loss of free available chlorine in Dakin’s solution during contact with biological materials, both in vitro and ex vivo, under laboratory conditions. First, 30 ml of dilute DS (nominal 0.001% to 0.0125% mass NaClO in 0.9% normal saline) was reacted with human plasma (1% or 5% by volume) for 30 minutes at room temperature in opaque black 50 ml polypropylene tubes. Available chlorine levels were measured throughout the incubation time course by removing serial 200 µl samples of the DS/plasma mixture (beginning at 5 seconds) for processing and titration as described above. Time-matched control samples consisting of human plasma (1% or 5% by volume) in 0.9% normal saline were obtained and processed at each time point. Next, DS degradation during contact with skeletal muscle tissue was characterized using a rat ex vivo simulated wound model. This wound model consisted of a disarticulated hind limb of a male Sprague-Dawley rat, obtained under an approved tissue-sharing agreement, in which a 30 mm incision was created parallel with the femur to expose the gluteus maximus and biceps femoris. The incision was carried through to the level of the femoral periosteum and small tissue retractors were engaged at opposite ends of the incision to create a hollow pocket capable of holding approximately 3 ml of solution. The wound pocket was irrigated with saline and blotted dry with sterile gauze. 2.5 ml of DS equivalent in strength to commercial “dilute” Dakin’s solution (0.0125% mass NaClO) was applied as a single bolus and left in place for 30 minutes. Available chlorine was serially measured throughout the time course (5 seconds to 30 minutes) by assaying 200 µl samples of solution as above. Time-matched control samples were obtained from an identical hind limb wound model treated with 0.9% normal saline.

Results

Available chlorine assay

The practical lower limit of quantitation for a 200 µl sample using this assay was found to be approximately 0.001% mass available chlorine, as determined by the ability to reliably identify the 490 nm absorbance shift indicating equivalence. If required, this method could be easily adapted to measure the available chlorine content of substantially smaller samples (<50 µl) of more concentrated solutions (eg. ≥ 0.004% mass Cl₂). Agreement was excellent between measured and nominal percent mass available chlorine (r²=0.992) over a range of 0.001% to 0.125% mass sodium hypochlorite (Figure 2). Over this concentration range, the mean difference between measured and nominal values was less than 1% with no individual measured value more than 10% different from its respective nominal value (Figure 3). Accuracy and repeatability of titrant addition steps were within the limits of available instrumentation to determine differences in expected and measured weights of distilled, deionized water dispensed by the automated syringe dispenser module.

Assay qualification

Contact with biological material causes extremely rapid loss of reactive chlorine in solution. In the presence of 5% (v/v) human plasma in solution, 0.001% mass NaClO Dakin’s solution demonstrated 61% degradation after only 5 seconds (the practical time...
discrimination limit of this assay) with degradation progressing to 72% by 1 minute and 85% by 30 minutes (Figures 4A and 4B).

**Figure 4A:** Available chlorine degradation curves of Dakin's solution (DS) in contact with human plasma in vitro. 0.001% DS incubated with 1% or 5% (v/v) human plasma for 30 minutes.

**Figure 4B:** Available chlorine degradation curves of Dakin's solution (DS) in contact with human plasma in vitro. 60 second kinetic profile of 0.001% DS incubated with 5% (v/v) human plasma. Data represents mean ± SEM of N=3 replicate experiments; dashed line indicates measured concentration of unreacted DS prior to plasma exposure.

Total degradation was proportional to plasma concentration, with 1% (v/v) human plasma producing 13% degradation of 0.001% DS by 5 seconds, 32% degradation at 1 minute, and 55% degradation at 30 minutes (Figure 4B). Similar degradation profiles were observed at higher DS starting concentrations, e.g. 0.0125% DS exhibited 45% degradation after 5 seconds of contact with 5% human plasma which progressed to 62% degradation at 1 minute and 75% overall loss by 30 minutes (Figure 5). Contact with soft tissue in an ex vivo wound model resulted in a slightly slower loss of free available chlorine compared to the reaction with plasma in solution: 0.0125% DS demonstrated approximately 15% degradation after 5 seconds of tissue contact, 43% degradation after 1 minute and 72% degradation by 30 minutes (Figure 6). The short centrifugation step incorporated into the sample preparation procedure was found to be sufficient to clear precipitated protein that could interfere with absorbance measurement if left in suspension.

**Figure 5:** Available chlorine degradation curves of 0.0125% Dakin's solution (DS) incubated with 1% or 5% (v/v) human plasma in vitro over 30 minutes. Data represents mean ± SEM of N=3 replicate experiments; dashed line indicates measured concentration of unreacted DS prior to plasma exposure.

**Figure 6:** Reactive chlorine activity over time in 0.0125% Dakin's solution (DS) during contact with soft tissue of an ex vivo rat simulated wound model. 2.5 ml DS instilled into wound space as a single bolus without replenishment for 30 minutes. Data represents mean ± SEM of N=3 replicate experiments; dashed line indicates measured chlorine concentration of unreacted DS used for treatment.

**Discussion**

The semi-automated method for free available chlorine determination described here represents numerous improvements to the traditional iodometric oxidation/reduction titration approach (Equation 6) intended to decrease required sample volume (200 µl compared to 2-4 ml needed for conventional methods), drastically increase throughput, and minimize manual sample handling and error. This improved assay utilizes partially saponified polyvinyl alcohol (PVA) as a colorimetric iodine indicator (Equation 7) as previously validated by Yoshinaga et al. [23] to both substantially improve
sensitivity over the conventional starch indicator solution and enable one-step acidification and indicator addition suitable for automated titration. Furthermore, both the sodium thiosulfate titrant addition and colorimetric monitoring steps (Equation 8) are carried out automatically in this method, providing significantly higher sample throughput with reduced human error when compared to a typical manual titration assay. In testing, we found that 32 individual samples could be titrated in technical triplicate in approximately 27 minutes using this method.

(Eq. 6) \[ \text{Cl}_2 + 2 \text{KI} \rightarrow \text{I}_2 + 2 \text{KCl} \]

(Eq. 7) \[ \text{I}_2 + \text{PVA} \rightarrow \text{I}^- \cdot \text{PVA} \text{ Complex (} \lambda_{\text{max}}=490 \text{ nm)} \]

(Eq. 8) \[ \text{I}^- \cdot \text{PVA} \text{ Complex} + 2 \text{S}_2\text{O}_3^{2-} \rightarrow 2 \text{I}^- + \text{S}_4\text{O}_6^{2-} + \text{PVA} \]

This assay is particularly suitable for the measurement of active chlorine concentrations for in vivo studies because it is less susceptible to error caused by sample contamination and discoloration (eg. hemoglobin, precipitated protein, etc.) than previous direct spectrophotometric methods [7] as it operates via the detection an abrupt colorimetric shift that is largely independent of changes in background absorbance. Other rapid, high-throughput microtiter plate-based assays have been developed for the measurement of antiseptic agents, specifically hydrogen peroxide and peracetic acid [24], but this is the first assay for free available chlorine in which titrant addition and equivalence monitoring are carried out automatically without manual intervention.

As noted above, the summed concentrations of HOCl, OCl\(^-\) and dissolved Cl\(_2\) are collectively referred to in this work as percent mass available chlorine in solution. This nomenclature was specifically chosen for two distinct reasons: first, this term provides a practical means of describing the total oxidative potential of solutions of undefined pH or chemical composition (such as antiseptic solutions in contact with biological materials) in which the actual chlorine species profile (eg. [Cl\(_2\)] versus [HOCl ] versus [OCl\(^-\)]) is unknown. Second, this terminology directly relates measured active chlorine content to the commercial label "strength" of sodium hypochlorite-based solutions (such as DS), which is frequently expressed as percent mass NaClO. To clarify, the molar equivalency in oxidizing power between NaClO, HOCl, and Cl\(_2\) (Equations 1, 3 and 6), allows the nominal equivalent percent mass available chlorine for a given NaClO preparation to be calculated as follows:

% Mass Available Cl\(_2\) = \frac{\% Mass NaClO}{\text{MW NaClO}/\text{MW Cl}_2} = \frac{\% Mass NaClO}{74.44/70.91} = 1.05 \%

This close approximation between percent mass active chlorine and label NaClO concentration allows degradation kinetics to be evaluated as a direct loss of chlorine activity from a nominal starting point. While the mass equivalency between label strength and chlorine is less direct for pure, stabilized hypochlorous acid solutions (MW HOCl = 52.46 g/mol), the titration assay described here performs identically for all mixtures of Cl\(_2\), HOCl, OCl\(^-\), and NaClO in commercial antiseptic solutions.

The rapid degradation profile exhibited by Dakin's solution in contact with biological materials is consistent with the high protein reactivity previously reported for hypochlorous acid [25]. In our experiments, reactive chlorine was rapidly depleted, on the order of seconds to minutes, in solutions containing even low concentrations of aqueous human plasma. Degradation was slightly slower, however, during contact with a clean tissue surface in an ex vivo simulated wound model. This different reaction profile may be attributable to the limited availability of labile biomolecules at the tissue surface in conjunction with the relatively slow rate of solubilization of unreacted molecules from the tissue into the solution phase immediately after addition of Dakin's solution. The observed rate of degradation would likely be higher in living, perfused tissue where plasma proteins would actively be infused into an exposed wound space containing DS.

Conclusions

The novel method for the determination of active chlorine described here offers multiple improvements over existing methods, and will be of value to other researchers investigating the degradation kinetics of chlorine-based solutions in contact with biological materials. This assay, and the preliminary data reported here, form a foundation for future in vivo and clinical studies focused on optimizing the dosing of chlorine-based topical antiseptics solutions to maximize antimicrobial effectiveness with minimal host tissue toxicity to improve wound healing outcomes.

Disclosures

The opinions or assertions contained herein are the private views of the authors and are not to be construed as official or as reflecting the views of the Department of the Army or the Department of Defense. The authors declare no financial or nonfinancial conflicts of interest. The authors are employees of the U.S. Government, and this work was prepared as part of their official duties.

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