#### ISSN: 2952-8119

**Open Access** 

# A Note on Pathobiology and Endodontic Microbiology

#### Shaon Tiwari\*

Department of Microbiology, Maulana Azad Medical College, New Delhi, India

#### Introduction

Endodontic disease affects the pulp or periapex of the tooth and is caused by a wide variety of microorganisms that are ordinarily found in the oral cavity. Although many of the bacteria present in the necrotic pulp are commensal under normal circumstances, their makeup changes and many of them become more numerous and harmful. The lack of collateral circulation and the dental pulp's enclosure within the mineralized tissues of the tooth limit the dental pulp's ability to fight itself against advancing oral bacteria, despite the fact that it displays a strong immune response. A periapical lesion results as a result of the pulp losing vitality under these circumstances at rates that are higher than any other tissue in the body.

Endodontic disease affects the pulp or periapex of the tooth and is caused by a wide variety of microorganisms that are ordinarily found in the oral cavity. Although many of the bacteria present in the necrotic pulp are commensal under normal circumstances, their makeup changes and many of them become more numerous and harmful. The lack of collateral circulation and the dental pulp's enclosure within the mineralized tissues of the tooth limit the dental pulp's ability to fight itself against advancing oral bacteria, despite the fact that it displays a strong immune response. A periapical lesion results as a result of the pulp losing vitality under these circumstances at rates that are higher than any other tissue in the body.

## **Description**

Endodontic disease is primarily a microbiological condition that is originated and spread by a complex community of microorganisms that are typical members of commensal oral microflora, as opposed to one or several specific bacterial or fungal species. The key benefits of various microbial detection techniques. For many years, the gold standard for studying bacteria's pathogenicity, interactions, and susceptibilities to local and systemic treatment methods was to grow the bacteria (and to a lesser extent the fungi) from infected root canals or periapical abscesses. Despite the fact that culturing is still the method of choice for examining the phenotypic traits of bacteria and their susceptibility to antimicrobials, it has recently been evident that only approximately half of oral bacteria are cultivable [1,2].

Additionally, some bacteria that are typically easy to cultivate in the oral environment may become impossible to cultivate even though they are viable if the environment contains elements or circumstances that hinder growth in a lab [3]. This is particularly important in the endodontic setting, especially if bacteria are exposed to some endodontic chemicals used during treatment that may momentarily inhibit bacterial growth, including calcium hydroxide or antibiotics [4].

\*Address for Correspondence: Shaon Tiwari, Department of Microbiology, Maulana Azad Medical College, New Delhi, India; E-mail: tiwari.1983@gmail.com

**Copyright:** © 2022 Tiwari S. This is an open-access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

Date of Submission: 02 June, 2022, Manuscript No: jmbp-22-75169; Editor assigned: 04 June, 2022, PreQC No: P-75169; Reviewed: 21 June, 2022, QC No: Q-75169; Revised: 01 July, 2022, Manuscript No: R-75169; Published: 07 July, 2022, DOI: 10.37421/2952-8119.2022.6.160

Taking samples in an endodontic setting is quite difficult. It is well knowledge that bacteria build up on root canal walls as biofilms over the course of weeks, months, or even years, depending on how old the infection is. Planktonic bacteria are primarily collected during sampling using paper points inserted into uninstrumented canals. Even if files are employed to disrupt the biofilm, it is known that only a small portion of the canal wall surface area is actually touched by the files. Sampling tools hardly ever reach isthmi, fins, lateral canals, or dentinal tubules. Coronal and apical bacterial microflora likely have different effects on the development, manifestation, or healing resistance of diseases. As a result, sampling is likely to reveal a negligible amount of root canal bacteria present.

Significant progress has been achieved in molecular techniques for microbial detection, identification, and enumeration over the past 20 years. Molecular techniques enhance depth of coverage, taxonomic classification accuracy, and sensitivity (the capacity to detect very low amounts of bacteria). Beginning with the specific amplification of a microbial species, broad-range cloning and sequencing of the bacterial 16S gene to identify representatives of unknown bacteria present in the community, and deep sequencing that enable the detection of a significant portion of bacteria, including low-abundance taxa, have all been used in the study of molecular methods [5].

Modern microbiological analysis goes much beyond only identifying the bacterial or fungus species that are present. It is generally known that some bacterial strains are more virulent than others and that even members of the same species can behave differently depending on the environment and microbial composition. Additionally, when bacterial populations exceed a particular threshold, a characteristic known as quorum sensing, bacteria may begin to express critical virulence genes [2]. As a result, molecular microbiology today takes use of sequencing's widespread availability and low cost to look at the genetic profiles of all bacteria present. Shotgun sequencing, also known as metagenomic analysis, enables the discovery of numerous genes involved in taxonomy, structure, function, virulence, and antibiotic resistance. Identifying the genes simply indicates the possibility of specific capabilities being present, which is inadequate to determine the function. Proteomic analysis, transcriptomics, investigation of the metabolic pathways leading to particular activities, and analysis of the actual bacterial (and host) proteins expressed are more practical methodologies.

The capacity to identify particular microorganisms or discover germs in an endodontic case may be immediately beneficial clinically in a number of ways. Particular microorganisms may be linked to more symptomatic endodontic infections and antibiotic resistance; the presence of leftover bacteria at the moment of obturation, for instance, may be a predictor of long-term results. The presence of systemic viruses or local fungus may make infections more virulent and challenging to treat. These clinical investigations all involved the use of various bacterial detection techniques on infected teeth. Another study looked at different sizes of apical preparation in diseased canals but did not evaluate the microbiological condition of the prepared channel [6-8]. The findings revealed a dose-dependent improvement in healing for bigger apical sizes and a substantial improvement in healing when preparation was completed to three file sizes larger than the initial file. It is anticipated that larger diameters caused more disruption of the apical bacterial biofilm and more bacterial removal through disinfectant irrigation because all subjects in this study had large canals with apical periodontitis.

As was already mentioned, there is a wide variety of bacterial pathogens that are suspected to be involved in endodontic illness. These germs originate from the gingival microflora, which normally inhabits the mouth cavity. In situations of endodontic and periodontal disease, it has been found that the prevalence of particular endodontic and periodontal pathogens strongly correlates [9]. The quantity of bacteria in the root canal and periodontal environments of endodontic-periodontal diseases differs, despite similarities in overall makeup, according to a study conducted more recently utilising modern sequencing technologies.

Because of this, the numerous oral or periodontal bacteria that are present during the pathogenesis of endodontic infections are the first to enter the root canal and move toward the periapical tissues. The community that results eventually contains many of the initial taxa, but in varying proportions, according to ecological and nutritional forces. The extent to which this overlap occurs was demonstrated by one study that tracked the spread of germs in the same patient population from the normal oral environment to the necrotic root canal to the periapical abscess.

One of the most significant developments in host response research over the past ten years has been the understanding that structural cells including odontoblasts, osteoblasts, and endothelial cells can recognise and defend against microbial irritants as well as immune cells [10,11]. The odontoblast is a cell that exhibits what appears to be a wide range of immunologic capabilities, including the production of TLRs (TLR-2 and TLR-4), contact with dendritic cells, and secretion of chemokines like interleukin-8 (IL-8), which has a potent antimicrobial activity.

## Conclusion

Approximately two decades ago, investigations revealed that gene polymorphism may exist among people, resulting in variations in disease susceptibility, clinical presentation, and therapeutic response. Individual variations in gene sequences, even as small as single-nucleotide polymorphism, can be used to discover gene polymorphism. As previously mentioned, genetic variability across patients may cause variations in disease manifestation and post-endodontic pain perception among individuals. The influence that candidate genes may have on the healing of endodontic illness a year after treatment have also been studied in a number of studies, while adjusting for other factors.

## **Conflict of Interest**

There is no conflict of interest by author.

#### **Acknowledgement**

Not applicable.

#### References

- Narayanan, L. Lakshmi and C. Vaishnavi. "Endodontic microbiology." J Conserv Dent 13 (2010): 233.
- Jhajharia, Kapil. "Microbiology of endodontic diseases: A review article." Int J Appl Dent Sci 5 (2019): 227-230.
- Marshall, G., L. Canullo, RM. Logan and G. Rossi-Fedele. "Histopathological and microbiological findings associated with retrograde peri-implantitis of extra-radicular endodontic origin: a systematic and critical review." Int J Oral Maxillofac Surg 48 (2019): 1475-1484.
- 4. Fouad, Ashraf F., ed. Endodontic microbiology. John Wiley & Sons, 2017.
- Nair, P.N.R. "Strength of evidence in current endodontic microbial research." Oral Surg Oral Med Oral Pathol Oral Radiol Endod 105 (2008): 8-10.
- Craig, John R., Roderick W. Tataryn, Bruce Y. Cha and Pallavi Bhargava, et al. "Diagnosing odontogenic sinusitis of endodontic origin: A multidisciplinary literature review." *Am J Otolaryngol* 42 (2021): 102925.
- Siqueira Jr, José F., Isabela N. Rôças, Renata Souto and Milton de Uzeda, et al. "Microbiological evaluation of acute periradicular abscesses by DNA-DNA hybridization." Oral Surg Oral Med Oral Pathol Oral Radiol Endod 92 (2001): 451-457.
- Siqueira Jr, José F., and Isabela N. Rôças. "Dialister pneumosintes can be a suspected endodontic pathogen." Oral Surg Oral Med Oral Pathol Oral Radiol Endod 94 (2002): 494-498.
- 9. Nair, PNR. "Endodontic biofilm, technology and pulpal regenerative therapy: where do we go from here?" Int Endod J 47 (2014): 1003-1011.
- Maltz, M., SL. Henz, EF. De Oliveira, and JJ. Jardim. "Conventional caries removal and sealed caries in permanent teeth: A microbiological evaluation." J Dent 40 (2012): 776-782.
- 11. Kalfas, Sotirios, David Figdor and Göran Sundqvist. "A new bacterial species associated with failed endodontic treatment: identification and description of Actinomyces radicidentis." *Oral Surg Oral Med Oral Pathol Oral Radiol Endod* 92 (2001): 208-214.

How to cite this article: Tiwari, Shaon. "A Note on Pathobiology and Endodontic Microbiology." J Microbiol Pathol 6 (2022): 160.