A Hypothesis for Mechanisms of Weakness Distribution in Muscular Dystrophies

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Abstract

Objective: The distribution of muscle weakness in muscular dystrophies varies, involving proximal muscles, distal muscles, or both. The mechanism underlying the differences in distribution is unknown.

Method: We compare and analyze the location and function of the culprit proteins responsible for muscle diseases and show that muscle weakness often follows a common pattern.

Results: Muscular dystrophies causing proximal weakness are due to mutations in genes coding for membrane-bound proteins, while conditions causing a predominantly distal muscle weakness are due to abnormal intracellular functional proteins of the sarcomere, the Z-disk or the cell metabolism.

Conclusion: This classification could enable predicting the function of newly identified myopathy-related genes according to their clinical presentation.

Keywords: Muscle; Muscle dystrophy; Muscle weakness

Introduction

Weakness distribution is a key element in muscle diseases. It may involve proximal or distal muscles, and this division is sometimes characteristic of the disorder. However, the reason for the dichotomic distribution remains a challenging and enigmatic feature of primary muscle syndromes.

Recent years have shown an impressive increase in knowledge regarding the genetic, metabolic, physiologic and structural basis of muscles in health and disease. This includes understanding the inner structure and location of muscle proteins, their function, and their genetic predisposition to mutations [1]. Based on these new discoveries, we propose that the intracellular location and function of proteins involved in the pathogenesis of muscular dystrophies (MD) determines the anatomic distribution of weakness – proximal, distal, or both (Figure 1).

Literature Review

The hypothesis

We hypothesize that the intracellular location and function of proteins involved in the pathogenesis of MD determine the distribution of weakness. Those causing proximal weakness are due to abnormal proteins located on the muscle membranes, including outer (sarcolemma) and inner (sarcoplasmonic reticulum, the nuclear membrane) membranes. Those causing a predominantly distal muscle weakness connect to functional intracellular proteins of the sarcomere or the Z-disk. Thus, dysfunction in proteins associated with the protection of membranes, and those involved in the regeneration and repair after membrane injury, lead to proximal muscle weakness. On the other hand, proteins implicated in distal muscle weakness are involved in inner sarcomere mechanics and stability of the Z-disk.

Testing the hypothesis

Our premise, illustrated below, focuses on the location and function of the culprit gene products in the various muscle abnormalities.

Proteins of the proximal MD

Dystrophin is a membrane protein linking actin filaments to the sarcolemma that is thought to contribute to protecting muscle fibers from micro-tears during contraction [2]. Mutations in the dystrophin gene cause Duchene Muscular Dystrophy and Becker muscular dystrophy. The clinical phenotype is that of progressive proximal myopathy [3]. Sarcoglycans are proteins connected to dystrophin and the muscle cell membrane. In conjunction with dystrophin they build a dystrophin-glycoprotein complex that participates in muscle membrane stabilization during contraction, and its dysfunction leads to proximal weakness (Figure 2A). The caveolins are proteins involved in building invaginations observed in muscle membrane that are responsible for molecular signaling and lipid regulation [4]. Disease mechanisms include a reduced half-life of caveolin and the aggregation of mutated caveolin-3 in the Golgi apparatus. Mutations in caveolin-3 cause a proximal myopathy (Table 1).

Mutations in the gene for collagen VI underlie Bethlem myopathy, which causes proximal weakness. Collagen VI is located on the outer surface of muscle membrane, as part of the extracellular matrix, and probably functions as an anchoring protein between muscle basement membrane and the surrounding tissue [5]. Lamin A/C, located in the near nuclear membrane, is involved in membrane integrity and signaling. Lamin A/C dysfunction increases the vulnerability of the nuclear membrane to mechanical strain and induces changes in nuclear
Proteins of the distal MD

The ZASP protein, located at the Z-disk, is thought to play a role in connecting titin fibers to the Z-disk via α-actinin. ZASP gene mutations cause a slow progressive weakness of soleus and gastrocnemius muscles, with a later progression to the distal muscles of the upper extremities known as Markesbery-Griggs MD [8]. Myosin-7 is part of the heavy chain of slow muscle type (Type I) and is a constituent of the sarcomere. Mutations in the myosin 7 gene are responsible for Gower-Laing muscular dystrophy that consists of distal weakness of dorsiflexion beginning in childhood with slow progression [9]. αB-crystallin (CRYAB), located near the Z-disk, has high affinity to desmin and titin, and protects myosin from premature destruction. Several mutations in CRYAB were shown to cause distal MD (Figure 2B). Glucosamine-(UDP-N-acetyl)-2-epimerase/N-acetylmannosamine kinase (GNE) is an enzyme located in the cytoplasm, Golgi apparatus, and nucleus (Table 2). It is responsible for the synthesis of sialic acid and is involved in cellular signaling [10]. Mutations in the GNE gene are responsible for distal myopathy with rimmed vacuoles (Nonaka distal MD) [8].

VCP is involved in a number of essential cellular processes such as apoptosis, cell cycle control, membrane fusion, transcription activation and ubiquitin-mediated protein degradation. It causes structural changes in target proteins using mechanical force. Mutated VCP was shown to cause distal MD [11]. Kelch-like (KLHL) gene family is a group of proteins that are involved in antioxidant responses, organization of actin cytoskeleton and protein ubiquitination [12]. Abnormality in KLHL-9 causes distal MD. Matrin-3 is a protein located in the nucleus that is thought to stabilize mRNA transcripts and DNA repair. Mutations in the nuclear matrix protein matrin-3 cause vocal cord paralysis with pharyngeal weakness and distal myopathy (VCPDM) [13].

Defective proteins that involve both proximal and/or distal muscles

Mutations in several key proteins cause both proximal and/or distal muscle weakness. These combined muscle diseases can be divided into two types: i) Mutations presenting with either proximal or distal weakness. ii) Mutations causing both proximal and distal weakness. Titin is a giant protein responsible for muscle elasticity and sarcomere organization that protrudes between two adjacent sarcomeres. It is responsible for sarcomere support and signaling. Mutations in the titin gene can cause either distal tibial myopathy (TMD), or a proximal myopathy of limb girdle type (LGMD2J) [14]. During eccentric contraction, titin increases its stiffness and as a result there is an increase in muscle force upon stretch [15]. When this function is lost, there is extensive proximal muscle damage (Tables 3 and 4). The two phenotypes (distal and proximal weakness) can be explained by the different isoform expressions of titin, as distal muscle have longer titin isoforms with a trend to associate with faster muscles and higher passive tension compared to proximal muscles (Figure 2C).

Nebulin, like titin, is a giant sarcomere protein regulating the length of thin filaments. Mutations in the nebulin gene either lead to a distal MD or, as nemaline rod myopathy (NEM2), a primarily proximal weakness later progressing to distal involvement. Dysferlin is located at the sarcolemma and thought to be involved in muscle membrane repair, possibly by regulating Ca²⁺ induced membrane fusion. Mutations in the large dysferlin gene lead to either a distal MD, the Miyoshi Myopathy, or to a proximal MD, limb girdle muscular dystrophy type 2B (LGMD2B) [16]. The wide spectrum of mitochondrial abnormalities in patients and impaired vesicle migration in dysferlin null muscle fibers in vivo might explain the wide range of phenotypes in dysferlinopathies. Desmin is an intermediate filament that is bound to the Z-disk, to the muscle

Figure 1: A schematic cross section of a muscle fiber depicting the probable location of proteins causing different types of muscle dystrophies.

nuclear membrane, and to extracellular matrix. Normal eccentric muscle contractions cause extensive damage to desmin fibers. Thus, desminopathies have a wide range of phenotypes: proximal weakness with mitochondrial dysfunction of fast muscles; Emery-Dreifuss like syndrome; and even a myasthenic syndrome. Most common, however, is a progressive myopathy involving distal muscles. However, cases were also reported of severe diffuse weakness, myopathy with cardiomyopathy [17], isolated cardiomyopathy, and limb girdle pattern of weakness (LGMD1E) [18].

Myotilin is a Z-disk protein involved in sarcomere stabilization. Mutations in myotilin cause proximal myopathy similar to that of LGMD1A. A far more common phenotype of myotilin gene mutation is a late onset distal MD, probably attributed to the involvement of myotilin in myofibrillar remodeling after eccentric muscle contractions [19,20]. Filamins are proteins responsible for actin crosslinking in the cytoskeleton. Filamins also interact with γ- and δ-sarcoglycans at the sarcomere membrane [21]. Mutations at the N-terminus of filamin-C cause distal MD, but in four German families the presentation was mainly of proximal limb girdle manner.

Calpains are a group of proteases involved in protein ubiquitination. Mutations in the calpain-3 gene cause LGMD2A present as weakness of the glutei and hip adductors, together with involvement of soleus muscle and winging of the scapula [22].

Telethonin is one of the Z-line proteins. Its N-terminal is connected to titin and ot stabilizes the interaction between titin and the Z-disk. Telethonin has a role in muscle maturation by regulating the stiffness of slow twitching muscles and is involved in myostatin regulation pathway [23]. Hence, telethonin protein changes cause a wide range of phenotype presentations from limb girdle muscle dystrophy 2G (LGMD2G), to severe distal weakness with proximal muscle involvement, congenital muscle dystrophy, proximal muscle weakness with involvement of facial muscle with tibial sparing [24], as well as proximal weakness [25].

TRIM-32 functions as a part-E ubiquitin ligase and is expressed mainly in proximal muscles, as well as in the gastrocnemius. It binds to the head and neck of myosin molecules and participates in ubiquitination of actin. Muscle stem cell differentiation during muscle regeneration is influenced by TRIM-32. This dual mode of action may explain the clinical presentation of both proximal (LGMD2H) [26], and distal weakness.

Discussion and Conclusion

The distribution of muscle involvement and weakness in primary muscle disease is an enigma. Since distal and proximal muscles have different functions and physiology, it is tempting to hypothesize that the function of the culprit gene product determines which set of muscles is involved. The present analysis attempts to shed some light on the mechanisms responsible for this disparity.

Our disease-by-disease examination of the metabolic/genetic abnormalities underlying chronic muscle disease, based on their cellular location and physiological function, provides convincing arguments to show that there are three venues responsible for the clinical phenotype.

1) Muscle diseases with proximal muscle dysfunction (proximal myopathies) are due to damaged associated muscle fiber membranes, or to muscle nuclear membranes with defective membrane repair mechanisms (Figure 2A). As proximal muscles are mainly responsible for eccentric contraction, defective proteins supporting this action that malfunction will present with proximal muscle wasting and weakness. Indeed, proteins causing proximal weakness are those supporting eccentric contraction that are membrane-bound, and thus are located on the outer cell membrane (sarcolemma) or inner membranes (SR and nuclear membranes). Therefore, mechanical injury due to
The culprit protein | Disease | Protein location | Protein function | Mechanism of myopathy |
---|---|---|---|---|
Dystrophin | Duchene, Becker | Near muscle membrane | Links actin filaments to the inside surface of muscle fiber's plasma membrane (sarcolemma). | Instability of muscle membrane during contraction, leads to muscle damage during contraction. |
Sarcoglycan | LGMD2C-2F | Muscle membrane | Part of dystrophin-sarcoglycan complex. | Muscle membrane is over susceptible to mechanical injury because of dysfunction of dystrophin sarcoglycan complex. |
Caveolin-3 | LGMD1C | Muscle membrane | Interacts with dystrophin and dyserin, participates in preservation of stability of muscle membrane. | Abnormal interaction with dystrophin; retention of caveolin molecules in Golgi apparatus; defective interaction with dyserin; reduction of mechanoprotection of muscle membrane. |
Collagen VI | Bethlem myopathy | Outer surface of muscle membrane | Synthesized by fibroblasts, has regulatory functions. | Unknown. Possible mechanism: 1) Loss of collagen VI causes excessive muscle stiffness and abnormal muscle regeneration via satellite cell dysfunction 2) Defective autophagy |
Lamin A/C | LGMD1B | Inner part of nuclear membrane, part of nuclear envelope | Might be involved in determination of nuclear shape and size, organization of nuclear pores and chromatin structure and gene expression. Mutation in laminas may change desmin localization and mechanotransduction. | Unknown. Possible mechanism: 1) Loss of stability of nuclear envelope and changes in mechanotransduction. 2) Changes in gene transcription |
Emerin | Emery Dreifuss | Nuclear envelope | 1) Mediates nuclear membrane anchorage to the cytoskeleton. 2) Connected to lamin, BAF and Btf (death promoting transcription receptor). 3) May influence actin dynamics. 4) Interacts with gene proteins. | Unknown. Possible mechanism: 1) Changes in connection between lamin and cytoskeleton cause changes in nuclear membrane stability. 2) Changes signaling and in genetic content with destruction of DNA and satellite cell dysfunction. |

LGMD: Limb Girdle Muscular Dystrophy; ER: Endoplasmic Reticulum.

| The culprit protein | Disease | Protein location | Protein function | Mechanism of myopathy |
---|---|---|---|---|
ZASP | Markesbury- Griggs | Z-disk | Binds to α-actinin that crosslinks thin filaments of sarcomeres, protecting the integrity of sarcomeres, and also interacts with actin. | Unknown. Possible disruption of sarcomere integrity. |
Myosin heavy chain -7 | Gower-Laing | Main myosin isoform in type I slow fibers | Muscle contraction. | Unknown. Possible abnormal myosin assembly and creation of myosin aggregates. |
CRYAB | αB-crystalline mutated distal myopathy | Near sarcomere | Chaperon activity, heat shock protein, protection from oxidative stress | Unknown. Possibly impairs heat shock protein activity, defective protection from oxidative stress and cytotoxic injury, interaction with actin. |
GNE epimerase | Distal myopathy with rimmed vacuoles ("Nonaka") | Cytoplasm near Golgi apparatus and nucleus | Rate limiting enzyme in the sialic acid biosynthesis pathway that is essential in protein glycosylation, cell adhesion and signal transduction. | Unknown. Possibly causes hypoglycosylation, mitochondrial, sarcomere organization and ubiquitination pathway problems. |
VCP | VCP mutated distal myopathy | Expressed in different parts of the cell | VCP is involved in regulation of cell cycle, apoptosis and protein ubiquitination, and mitochondrial function. | Interferes in one or more processes carried by VCP. |
KLHL9 | KLHL9 mutated distal myopathy | Cytoplasm | KLHL9 binds to Cu3 and forms an E3 ubiquitin ligase complex. | Disrupts KLHL9 -Cu3 complex. |
Matrin-3 | VCPDM | Nucleus | DNA repair and RNA stabilization | Unknown. Possibly impairs DNA repair and RNA stabilization. |


Table 2: Proteins causing distal muscle dystrophy are briefly reviewed according to protein location, function, possible mechanism of myopathy and levels of proof of the pathophysiological mechanism/s. Exact pathological processes are obscure in most myopathies of this type.

eccentric contractions damages the membrane structures in the proximal muscles.

2) In muscle disorders that have a predominantly distal distribution, the responsible abnormality lies in a defective protein associated with sarcomere function, namely concentric contraction that causes muscle shortening. This action is mainly confined to distal muscles (Figure 2B).

3) Most intriguing are those MDs that involve both proximal and distal muscles. In our hypothesis this group was the "internal control". If indeed phenotype is the outcome of both function
and cellular location of culprit genes, proteins that are associated with proximal and distal wasting should have either dual location and/or double function. This was the case in most but not all instances (Tables 3 and 4). To this group also belong patients with defects in proteins with a wide range of intracellular function, such as sarcomere and muscle membrane remodeling and repair and regulation of ionic channels' apoptosis (Figure 2C). Because of these important and diverse functions, mutations in these proteins may impair key mechanisms in muscle function, causing combined type of weakness.

Still, there are exceptions to this rule; the most important is dysferlin, a multifunctional protein located on the muscle cell membrane that causes proximal myopathy. As this protein is also associated with primary distal involvement in Miyoshi myopathy, one may postulate that this phenotype is mediated through the action of calpain-3, located near the sarcomere.

It should be emphasized that our hypothesis attempts to explain the distribution of muscle weakness only in degenerative muscle disorders. The distribution of weakness in other types of muscle diseases (inflammatory, endocrine, etc.) is probably more complex because

Table 3: Mutations in these proteins cause different phenomenological presentation, some cause proximal and some distal weakness. Both types are briefly reviewed according to protein location, function, possible mechanism of myopathy and levels of proof for the pathophysiological mechanism/s. Best levels of proof exists for titin, dysferlin and nebulin caused myopathies. The cause of myotillin caused myopathy is unknown.

<table>
<thead>
<tr>
<th>The culprit protein</th>
<th>Disease</th>
<th>Protein location</th>
<th>Protein function</th>
<th>Mechanism of myopathy</th>
</tr>
</thead>
<tbody>
<tr>
<td>Titin</td>
<td>Tibial muscular dystrophy (distal)</td>
<td>Protrudes between two sarcomeres, connected to Z-line proteins and M-line proteins</td>
<td>Giant protein that connects to actin filaments and myosin and is responsible for muscle elasticity. Plays role in muscle contraction and probably in signaling.</td>
<td>Dyfunction of titin kinase. Explanations for dual phenotype includes: secondary calpain-3 deficiency and different titin isoform expression in different type of muscles.</td>
</tr>
<tr>
<td>Nebulin</td>
<td>Distal nebulin myopathy (distal)</td>
<td>Sarcomere</td>
<td>Giant protein that connects to actin and Z-disk, stabilizes thin filaments, is responsible for muscle elasticity, and influences muscle contraction by interacting with actin and tropomyosin.</td>
<td>Unknown, possibly causes changes in length of thin filaments and forces reduction and abnormal involvement in muscle contraction.</td>
</tr>
<tr>
<td>Desmin</td>
<td>Desmin myopathy (distal)</td>
<td>Intermediate filament. Located near Z-disk and sarcolemma</td>
<td>Connects sarcomeres to the nuclei, extracellular matrix and sarcolemma; and acts in muscle contraction.</td>
<td>Unknown. Possible destruction of mesh of intermediate filaments and toxic desmin aggregation; changes in transduction and muscle strength and mitochondrial dysfunction</td>
</tr>
<tr>
<td>Myotillin</td>
<td>Myotilin distal muscular dystrophy (distal)</td>
<td>Z-disk</td>
<td>Binding of α-actinin protein, crosslinks actin filaments of sarcomere and controls sarcomere assembly; also connected to sarcolemma via muscle specific filaments.</td>
<td>Unknown. Possible defective myofibril reorganization after exercise</td>
</tr>
<tr>
<td>Filamin-C</td>
<td>Distal ABD filaminopathy (distal)</td>
<td>Z-disk and near muscle membrane</td>
<td>Crosslinks actin filaments.</td>
<td>Unknown, possibly causes structural changes in filamin leading to changes in interaction with actin and aggregation of filamin and its associated proteins.</td>
</tr>
</tbody>
</table>

LGMD: Limb Girdle Muscular Dystrophy; ABD: Actin Binding Domain.

Table 4: Abnormal proteins that present with more or less simultaneous involvement of proximal and distal muscles are briefly reviewed according to: protein location and function, name of disease caused with possible mechanisms of myopathy, and levels of proof for the pathophysiological mechanism/s. Best levels of proof exists for titin, dysferlin and nebulin caused myopathies. The cause of myotillin caused myopathy is unknown.

<table>
<thead>
<tr>
<th>Protein</th>
<th>Disease</th>
<th>Type of myopathy</th>
<th>Protein location</th>
<th>Protein function</th>
<th>Mechanism of myopathy</th>
</tr>
</thead>
<tbody>
<tr>
<td>Calpain-3</td>
<td>LGMD2A</td>
<td>Proximal and distal</td>
<td>Near titin at the sarcomere area, nucleus and near muscle membrane</td>
<td>Participates in sarcomere and membrane remodeling via promotion of myofibrillar protein turnover; interacts with dyserlin and sarcoglycans via filamin-C.</td>
<td>Unknown. Possible mechanisms include interactions with dysferlin and titin, muscle remodeling, cytoskeleton regulation, excessed apoptosis and loss of proteolytic activity.</td>
</tr>
<tr>
<td>Telethonin</td>
<td>LGMD2G</td>
<td>Proximal &gt; distal</td>
<td>Z-disk</td>
<td>Is a substrate of titin kinase. Binds to the titin at its C-part at Z1-Z2 domains and may be critical to sarcomere assembly and cell regulatory functions.</td>
<td>Unknown, possibly elevated muscle stiffness and inhibition of muscle cell differentiation.</td>
</tr>
<tr>
<td>TRIM 32</td>
<td>LGMD2H</td>
<td>Proximal &gt; distal</td>
<td>Nucleus and cytoplasmic bodies expressed mostly in hamstrings, soleus and diaphragm. Very low expression at quadriceps and triceps</td>
<td>1) Ubiquitin ligase, that interacts with myosin head and ubiquinates actin, and is involved in muscle degradation and construction during remodelling. 2) Regulates muscle stem cell differentiation essential for muscle regeneration.</td>
<td>Unknown, possibly involved in abnormal protein ubiquitination, stem cell dysfunction and loss of ability for self-activation.</td>
</tr>
</tbody>
</table>

TRIM 32: Tripartite Motif-Containing Protein 32; LGMD: Limb Girdle Muscular Dystrophy.
additional factors are involved other than proper protein dysfunction and muscle cell signaling. Furthermore, the degenerative muscle diseases caused by more complicated mechanisms than isolated protein dysfunction, such as myotonic dystrophy and facioscapulohumeral muscular dystrophy, represent exceptions to our hypothesis.

Implications of the Hypothesis

This rule of distribution of weakness could be useful for deciphering the function of an abnormal muscle fiber protein associated with a clinical phenotype. When the condition is characterized by proximal muscle weakness and atrophy, it might be reasonable to speculate that its function and location is associated with membrane integrity. Likewise, abnormal proteins causing distal muscle disease should be suspected to contribute to sarcomere action. Thus, by identifying the distribution pattern of the muscle weakness in a patient, it may be possible to predict the intracellular location of the culprit protein.

References