

High Resolution Magic Angle Proton Magnetic Resonance Spectroscopy (HRMAS) in Intact Sentinel Node Biopsy from Breast Cancer Patients: A New Diagnostic Tool!

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Abstract

Introduction: The extent of axillary lymph node involvement is one of the most important prognostic markers in patients of breast cancer. However, axillary dissection is associated with significant morbidity. The intra-operative sentinel node biopsy (SNB) provides a basis for omitting the routine axillary clearance however; use of in-house frozen section histopathology is required in order to substitute later. We report the use of in vitro high resolution magic angle proton magnetic resonance spectroscopy (HRMAS) for assessing the axillary nodal status with increased sensitivity.

Methods: Freshly removed axillary lymph nodes (n=17) obtained during sentinel node biopsy from 17 different patients were bisected. One segment of the bisected node was snap frozen and transported to HRMAS laboratory and was blindly subjected to 400 MHz high resolution magic angle proton magnetic resonance spectroscopy. The other portion was sent for frozen section biopsy. The tissues specimens used for HRMAS analysis and remaining portion of bisected node were then formalin fixed, paraffin embedded and sent for histo-pathological examination in separate vials. The metabolic profiles of these nodes were correlated with the routine histo-pathological findings.

Results: On histo-pathological examination, 7 nodes were found to be positive for metastasis were as 10 nodes were negative. The spectra of nodes (n=7) found to be positive for malignant cells were exclusively dominated by signals from choline, choline containing compounds and lactate in the spectral region of 3.2 ppm and 4.12 ppm respectively. Overall the sensitivity and specificity of HRMAS in the present study was 100%.

Conclusion: Metastatic and non-involved lymph nodes in breast cancer can be accurately distinguished based on its metabolic profile. The technique of high resolution magic angle proton magnetic resonance spectroscopy can be utilized in enhancing the sensitivity and specificity of sentinel node biopsy and may replace frozen section histopathology

Keywords: High resolution magic angle spectroscopy; Sentinel node; Breast cancer

Introduction

The extent of axillary lymph node involvement in breast cancer is a dominant indicator for systemic failure [1]. Therefore an adequate axillary lymph node dissection (ALND) along with mastectomy or breast conserving procedures is an established way to achieve a cure. However the procedure carries a significant morbidity in the form of sensory neuropathy, loss of shoulder mobility, shoulder pain and lymphedema of breast and arm [2].

Intra operative frozen section histopathology of the sentinel node (s) is an alternative to the standard axillary clearance however it has variable sensitivity of 60% to 95% [3,4]. Various methods such as immuno- staining for cytokeratin have been included along with frozen section histopathology to increase the sensitivity of the sentinel node biopsy [5].

The diagnostic workups using patho-morphological changes to some extent can be substituted by molecular diagnostics techniques. Molecular techniques of proteonomics, genomics and metabonomics have emerged as possible alternative or adjudicative to histomorphological tests [6-8]. The term metabonomics is defined as "the quantitative measurement of the dynamic multiparametric metabolic response of living system to pathophysiological stimuli or genetic modification". Magnetic resonance spectroscopy has emerged out as one of the main techniques of metabonomics and has been widely

used to assess the health risk of particular drug/toxins. The *in vitro* and *in vivo* application of magnetic resonance spectroscopy for the diagnosis and therapeutic monitoring of various medical and surgical conditions like the hydatid diseases, leishmaniasis, diagnosis of malabsorption syndrome and liver graft dysfunction have been described in the past [9-12]. The conventional technique of magnetic resonance spectroscopy [MRS] required separation of analytes by preparing time consuming and labour intensive tissue extracts. Furthermore, the specimen gets consumed in the process and is unavailable for histopathological examination. The technique of high resolution proton magic angle spinning (HRMAS) spectroscopy however is yet another advancement to analyze the metabolic profile of an intact specimen and has been used successfully to differentiate malignant breast tissue from adjacent normal tissue on the basis of metabolic finger prints [13]. The quick

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processing time is an added advantage that prompted us to evaluate the value of HRMAS spectroscopy *vis-a-vis* frozen section histopathology in sentinel node assessment from T₁/T₂ breast carcinoma patient in a pilot study.

Method

The study was carried out at the Department of General Surgery, CSM (King George's) Medical University, Lucknow and NMR Laboratory, Sophisticated Analytical Institute Facility, Central Drug Research Institute, Lucknow. The study was duly approved by the institutional ethical committee.

Sentinel node biopsy was performed in 17 patients by injecting blue dye (*iso-sulphan blue 1%*) in the subareolar region in patients planned for local wide excision for T₁/T₂ lesions with clinically negative axillae. None of the patients had received chemotherapy.

One largest blue node was taken for this study (Figure 1). The node was thoroughly washed in normal saline, bisected and one part was used for frozen section biopsy (Figure 2) and other was put into a cryogenic vial with a unique ID code. The vial was immediately snap frozen and carried in a liquid nitrogen container to the NMR laboratory facility (1.5 kms). The node was thinly sliced using a sharp surgical blade. A large slice about 2 to 3 mm thick and weighing 35 – 40 mg was put into 4 mm HRMAS rotor. The lymph node slice was assembled inside the rotor and 20 micro liter of D₂O containing tri methyl silyl tetra deuterion propionic acid (TSP) was added as standard. The HRMAS experiments were carried out on a Bruker Avance 400 MHz FT NMR spectrometer equipped with 4mm ¹H and ¹³C dual HRMAS with magic angle gradient at 4°C. The samples were spun at 4.0 KHz in order to keep rotation side bands out of the acquisition window.

One dimensional proton NMR spectra with water pre-saturation were acquired using NOESY pulse sequence with a mixing time of τ_m 100 milli seconds. Total relaxation delay of 3.99 sec was used using 8250.8 Hz spectral width, 128 transients with a total recording time of 9.44 minutes. The one dimensional CPMG pulse sequence with water pre-saturation using an echo time of 200 milli seconds was used in order to filter off short T₂ lipid component. Each experiment took about 20 minutes and the spectra were available for study on the dedicated computer screen. Assignments of HRMAS spectra were done as per published data [14]. The HRMAS spectra were read by the NMR expert (RR) who was not aware of the histopathological diagnosis of the sample. He was asked to deliver and sign a report in the shortest possible time to mimic the per-operative frozen section histopathology scenario. It may be noteworthy that routine frozen section facility was not available in our institution at the time of study. Completion axillary dissection was carried out in all 17 subjects.

The tissue used for the study was retrieved from the HRMAS rotor it was then formalin fixed and was taken for histopathological examination by standard H & E staining. The histopathology results were generally available a week later. The remaining tissue slices were also formalin fixed and sent separately for histopathological examination.

Results

Seventeen lymph node specimens from 17 subjects undergoing sentinel node biopsy from T₁/T₂ clinically N0 breast carcinoma patients were taken for the study. Each patient in this group yielded only one worthwhile lymph node for the study. The mean size of the nodes used for HRMAS study was 0.68 cms. Of all the differences in the cellular metabolism detected the high peaks of the metabolites viz; choline (Ch) and choline containing compounds eg; phosphocholine and phosphatidyl choline in the region of 3.2 ppm in 7/17 slices were most prominently seen (Figure 3). The presence of lactate at 4.12

ppm suggesting a raised anaerobic metabolism was also seen in all these 7 nodes. Furthermore, high concentration of amino acids was also observed in all these nodes. A portion of stack plot of the CPMG spectra depicting the presence of amino acid; glycine is shown in Figure 4. In 10/17 slices that were examined using the same HRMAS experiment in a clear contrast did not show the above metabolites. The NMR laboratory results could be interpreted almost instantaneously and one was quick to point out the above described differences and overtly 2 types of spectra. The HRMAS study data print outs in all the specimens examined from time to time were available within 30 minutes of receiving the tissue from the operating room.

The tissue slices retrieved from the NMR rotor were subjected to histopathology using H & E stain. There was clear and unequivocal evidence of malignancy in 7/17 nodes and none in 10/17 nodes. The histopathology of the remaining lymph node tissue slices also corroborated with the core slice examined. Upon decoding and correlating the histomorphological data with HRMAS findings it was obvious that overall the technique of HRMAS was 100% sensitive and 100% specific in these experiments (Table 1).

Discussion

This study reports a rather new efficient method of detecting tumor in sentinel node biopsy specimen from early stage breast cancer patients. The *in-vitro* use of High Resolution Magic Angle Spectroscopy on intact lymph node slice within 30 minutes from the operating room may be an important alternative to frozen section histopathological examination. Molecular diagnostic markers are an emerging field in cancer diagnosis and prognostic predictions. The *in vitro* technique of intact tissue metabonomics using HRMAS studies was able to differentiate cancer from non-cancer in this small sample of lymph nodes.



Figure 1: Extracted Blue Node

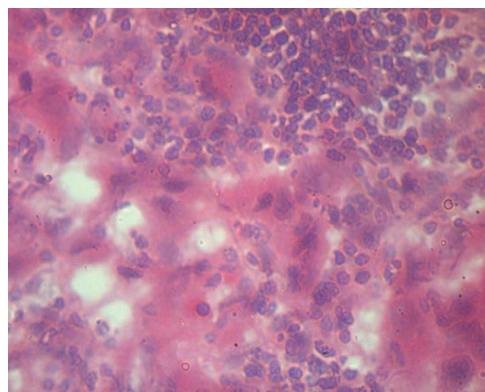


Figure 2: Frozen Section Biopsy of Involved Node.

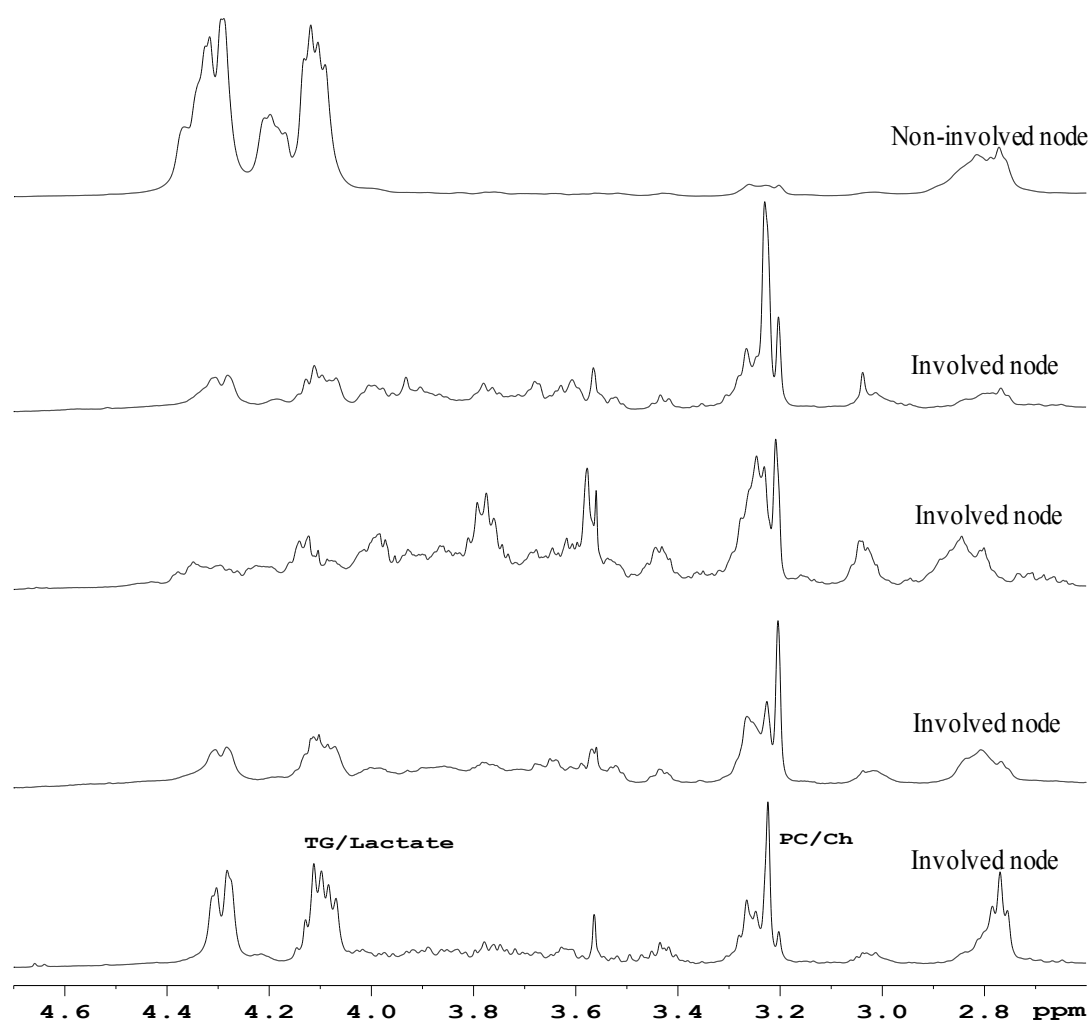


Figure 3: A Portion of 1D NOESY Stack Plot Spectra of Involved and Non-involved Lymph Nodes (PC- Phosphocholine, Ch- Choline, TG- Tri Glycide)

Choline and choline containing compounds are present in more than one tumor type but these metabolites are characteristically absent in normal and benign tissues [15,16]. Identification of cancerous secondary in an axillary sentinel lymph node as opposed to a non affected node indeed demonstrated a practical use of this knowledge. Thus, of all the alterations in the metabolic profile detected by the HRMAS spectra, the region of 3.2 ppm demonstrated the most useful metabolite choline, choline containing compound (phosphocholine) and lactate at 4.12 ppm. Choline and its derivatives are one of the important building blocks of cell physiology and represent accelerated cell proliferation in the presence of malignancy. Similarly choline has been detected in breast cancer tissue extract and ex-vivo MRS studies of breast cancer [17,18]. The choline and choline containing compounds signal were reported to be less prominent in patients of breast cancer treated with neo adjuvant chemotherapy [19].

Choline and choline containing compounds were also reported from lactating breast tissue [13,20]. Whether axillary lymph nodes in lactating women also show high choline peaks is unknown. The state of lactation is associated with increased choline metabolism because of the need to nourish the newborn with large amounts of choline (supplied in the milk predominantly as phosphatidylcholine, phosphocholine, glycerophosphocholine, and free choline). This may limit the specificity of choline and choline containing compounds as discriminating metabolite in lactating breast carcinoma, a rarity, however.

The second important discriminating metabolite in this study

was the lactate peaks at 4.12 ppm. In malignant cells the anaerobic metabolism of glucose is the major metabolic process and is thought to be responsible for the raised level of lactate [21].

The above described metabolites were indeed well known to occur in tissue extracts of cancers of several organs notably, breast, oral squamous cell carcinoma, urinary bladder and prostate cancer, brain tumours etc. Most of the earlier studies were done on cellular extracts of tumour in the early nineties. The metabolomics of intact tissue slices referred to as *ex vivo in vitro* tissue metabolomics is a rather recent development. The HRMAS technique was used to diagnose primary breast tumour and showed a high correlation with histopathology. The breast carcinoma tissue and juxta cancer tissue free from cancer were distinguished on the basis of these spectra [22]. Choline, lactate and other metabolites were significantly elevated in the malignant tissue. This study reported from China claimed it as a new technique for the diagnosis of human breast cancer in addition to histopathology. However there is a limitation for the use of lactate as a sole marker of malignancy in the detached specimens as it may be produced anaerobically and give a false positive result. Combined presence of choline and lactate along with other metabolites like creatinine, beta-glucose, GPC, glycine, myo-inositol and taurine were suggested as a sum total marker of malignancy with greater degree of confidence or diagnostic sensitivity in tissue samples.

The clinical radiologists can also do metabolites estimation from their standard *in vivo* magnetic resonance imaging (MRI) machine by simply using software. Such an *in-vivo* HRMAS in which the patient

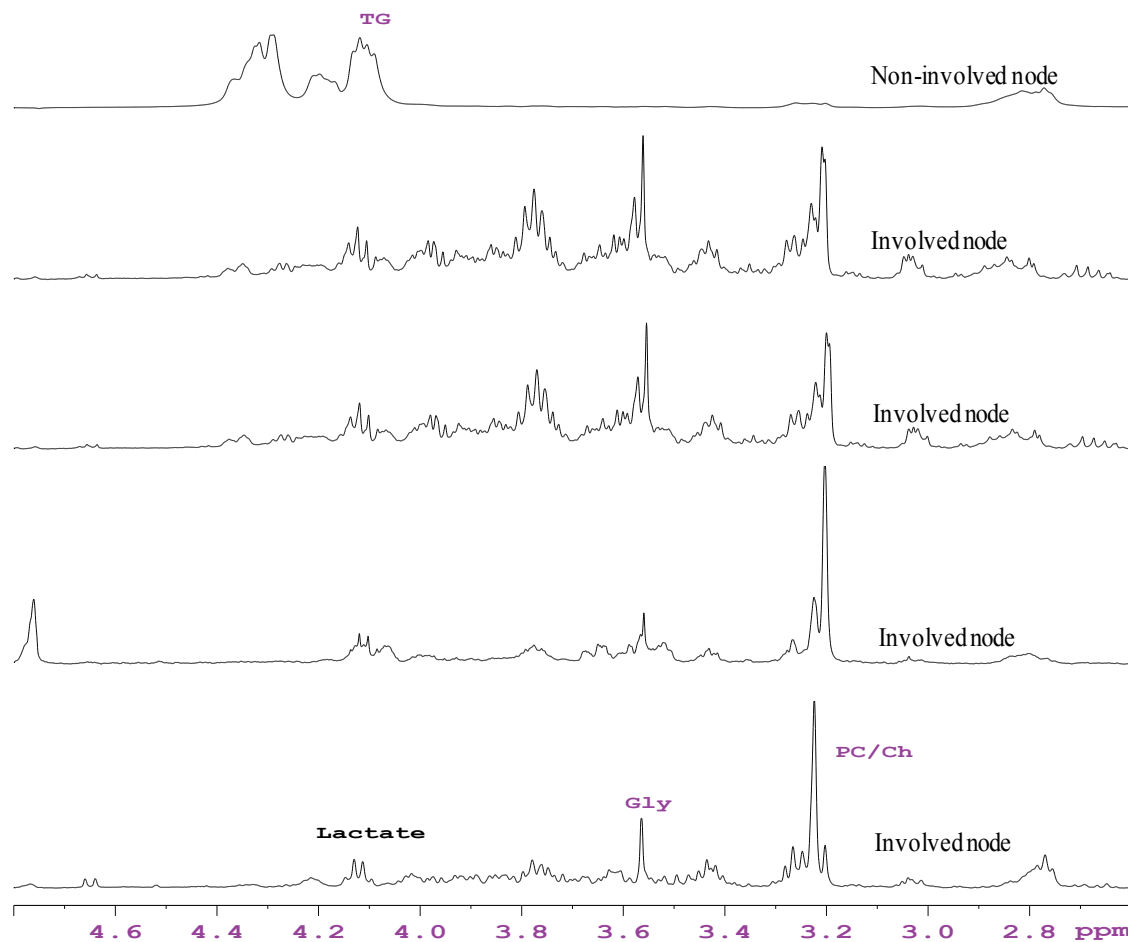


Figure 4: A Portion of CPMG Stack Plot Spectra of Various Lymph Nodes

Table 1: Test Results of HRMAS Compared with Histopathological Examination

	HPE +ve	HPE -ve	Total
HR MAS +ve	7	0	7
HR MAS -ve	0	10	10
Total	7	10	

Sensitivity: 100%

Specificity: 100%

Positive predictive value: 100%

Negative predictive value: 100%

herself goes into the MRI machine and the metabolic profile of the tumor is recorded with the help of special software has been evaluated in axillary nodes in breast cancer in 2 studies [23,24]. Firstly, in 35 nodes using the basis of choline signals alone the authors identified metastasis with sensitivity, specificity and accuracy of 82%, 100% and 90% [23]. In the second study, using the similar technique in 20 nodes the sensitivity, specificity and accuracy of 80%, 91%, and 88% were described. It was further noted in these studies conducted longitudinally on these patients that neo-adjuvant chemotherapy lowered the concentration of various metabolites e.g. choline, phosphocholine, etc. [24]. As these patients were undergoing chemotherapy and longitudinal assessment by MRI these describe a different subset of patients in whom rather large axillary lymph nodes were present as opposed to the non palpable nodes in the axillae of the patients that have been included in the present study. The ability of MRI to detect a sentinel node in clinically N0 axillae of early breast carcinoma has not been evaluated. The MRI along with *in vivo* MRS in clinically N0 axillae can theoretically be a tool for pre operative detection of malignancy in a sentinel node.

The molecular diagnosis of malignant change as opposed to histomorphological diagnosis is an emerging field of medical research. Metabonomics by MRS equipments are generally expensive commodity largely available in big public sector hospitals and corporate pharmaceuticals industries so far. There is relative paucity of trained man power in the field of MRS. Pharmacological companies and chemical industries routinely use MRS to test the purity of their products. Clinician's interest however, in this field has been tardy. An increasing interest and availability of the MRS equipment is being witnessed world over. The MRS or *in vivo* MRS and MRI as common facility can become cost effective in future with increasing usage. The *in-vitro* per-operative expeditious assessment HRMAS of an intact axillary lymph node slice or other tissues as a central facility for a number of hospitals in the vicinity of the HMRS facility can be promoted as a cost effective technology. Several other applications of *in-vitro* HRMAS studies include detection of micro-metastases, ability to detect residuals in the tumor bed and tumor margins, metabolites in tumor aspirate, exfoliated cells metabonomics, brush cytology specimens and, fluids like bronchial lavage, ascitic tap and nipple discharge. The initial fixed cost of the equipment though high is also likely to go down with widespread use in the future. Though at present the discriminating metabolites for a particular type of carcinoma or sarcoma are not available, the study designs like the present study offer a useful application of this technology within the present level of knowledge. This study was conducted to mimic the scenario of sentinel node assessment in the operating theatre. To this effect HMR spectroscopy emerged as an efficient and reliable method for the evaluation of the sentinel node as compared with routine histopathology. A larger sample size on a even higher frequency MRS (800 MHz) equipment may further enhance these results.

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